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<p>(54) Title: SYNTHETIC PEPTIDE INHIBITORS OF HIV TRANSMISSION</p>		
<p>(57) Abstract</p>		
<p>The present invention relates to peptides which exhibit potent anti-retroviral activity. The peptides of the invention comprise DP-178 (SEQ ID:1) peptide corresponding to amino acids 638 to 673 of the HIV-1_{LAI} gp41 protein, and fragments, analogs and homologs of DP-178. The invention further relates to the uses of such peptides as inhibitory of human and non-human retroviral, especially HIV, transmission to uninfected cells.</p>		

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SYNTHETIC PEPTIDE INHIBITORS OF HIV TRANSMISSION1. INTRODUCTION

The present invention relates to DP-178 (SEQ ID:1), a peptide corresponding to amino acids 638 to 673 of the HIV-1_{LAI} transmembrane protein (TM) gp41, and portions, analogs, and homologs of DP-178 (SEQ ID:1), all of which exhibit anti-viral activity. Such anti-viral activity includes, but is not limited to, the inhibition of HIV transmission to uninfected CD-4⁺ cells. Further, the invention relates to the use of DP-178 (SEQ ID:1) and DP-178 fragments and/or analogs or homologs as inhibitors of human and non-human retroviral, especially HIV, transmission to uninfected cells. Still further, the invention relates to the use of DP-178 as a HIV subtype-specific diagnostic. The present invention also relates to antiviral peptides analogous to DP-107, a peptide corresponding to amino acids 558 to 595 of the HIV-1_{LAI} transmembrane protein (TM) gp41, that are present in other enveloped viruses. The present invention further relates to methods for identifying antiviral compounds that disrupt the interaction between DP-178 and DP-107, and/or between DP-107-like and DP-178-like peptides. The invention is demonstrated by way of a working example wherein DP-178 (SEQ ID:1), and a peptide whose sequence is homologous to DP-178 are each shown to be potent, non-cytotoxic inhibitors of HIV-1 transfer to uninfected CD-4⁺ cells. The invention is further demonstrated by working examples wherein peptides having antiviral and/or structural similarity to DP-107 and DP-178 are identified.

2. BACKGROUND OF THE INVENTION

2.1. THE HUMAN IMMUNODEFICIENCY VIRUS

The human immunodeficiency virus (HIV) has been implicated as the primary cause of the slowly degenerative immune system disease termed acquired
5 immune deficiency syndrome (AIDS) (Barre-Sinoussi, F. et al., 1983, Science 220:868-870; Gallo, R. et al., 1984, Science 224:500-503). there are at least two distinct types of HIV: HIV-1 (Barre-Sinoussi, F. et al., 1983, Science 220:868-870; Gallo R. et al., 1984,
10 Science 224:500-503) and HIV-2 (Clavel, F. et al., 1986, Science 233:343-346; Guyader, M. et al., 1987, Nature 326:662-669). Further, a large amount of genetic heterogeneity exists within populations of each of these types. Infection of human CD-4⁺ T-
15 lymphocytes with an HIV virus leads to depletion of the cell type and eventually to opportunistic infections, neurological dysfunctions, neoplastic growth, and ultimately death.

HIV is a member of the lentivirus family of
20 retroviruses (Teich, N. et al., 1984, RNA Tumor Viruses, Weiss, R. et al., eds., CSH-Press, pp. 949-956). Retroviruses are small enveloped viruses that contain a diploid, single-stranded RNA genome, and replicate via a DNA intermediate produced by a
25 virally-encoded reverse transcriptase, an RNA-dependent DNA polymerase (Varmus, H., 1988, Science 240:1427-1439). Other retroviruses include, for example, oncogenic viruses such as human T-cell leukemia viruses (HTLV-I, -II, -III), and feline
30 leukemia virus.

The HIV viral particle consists of a viral core, composed of capsid proteins, that contains the viral RNA genome and those enzymes required for early
35 replicative events. Myristylated Gag protein forms an

outer viral shell around the viral core, which is, in turn, surrounded by a lipid membrane envelope derived from the infected cell membrane. The HIV envelope surface glycoproteins are synthesized as a single 160 Kd precursor protein which is cleaved by a cellular protease during viral budding into two glycoproteins, gp41 and gp120. gp41 is a transmembrane protein and gp120 is an extracellular protein which remains non-covalently associated with gp41, possibly in a trimeric or multimeric form (Hammarskjold, M. and Rekosh, D., 1989, Biochem. Biophys. Acta 989:269-280).

HIV is targeted to CD-4⁺ cells because the CD-4 cell surface protein acts as the cellular receptor for the HIV-1 virus (Dalglish, A. *et al.*, 1984, Nature 312:763-767; Klatzmann *et al.*, 1984, Nature 312:767-768; Maddon *et al.*, 1986, Cell 47:333-348). Viral entry into cells is dependent upon gp120 binding the cellular CD-4⁺ receptor molecules (McDougal, J.S. *et al.*, 1986, Science 231:382-385; Maddon, P.J. *et al.*, 1986, Cell 47:333-348) and thus explains HIV's tropism for CD-4⁺ cells, while gp41 anchors the envelope glycoprotein complex in the viral membrane.

2.2. HIV TREATMENT

HIV infection is pandemic and HIV associated diseases represent a major world health problem. Although considerable effort is being put into the successful design of effective therapeutics, currently no curative anti-retroviral drugs against AIDS exist. In attempts to develop such drugs, several stages of the HIV life cycle have been considered as targets for therapeutic intervention (Mitsuya, H. *et al.*, 1991, FASEB J. 5:2369-2381). For example, virally encoded reverse transcriptase has been one focus of drug development. A number of reverse-transcriptase-

targeted drugs, including 2',3'-dideoxynucleoside analogs such as AZT, ddI, ddC, and d4T have been developed which have been shown to be active against HIV (Mitsuya, H. et al., 1991, Science 249:1533-1544). While beneficial, these nucleoside analogs are not
5 curative, probably due to the rapid appearance of drug resistant HIV mutants (Lander, B. et al., 1989, Science 243:1731-1734). In addition, the drugs often exhibit toxic side effects such as bone marrow suppression, vomiting, and liver function
10 abnormalities.

Attempts are also being made to develop drugs which can inhibit viral entry into the cell, the earliest stage of HIV infection. Here, the focus has thus far been on CD4, the cell surface receptor for
15 HIV. Recombinant soluble CD4, for example, has been shown to inhibit infection of CD-4⁺ T-cells by some HIV-1 strains (Smith, D.H. et al., 1987, Science 238:1704-1707). Certain primary HIV-1 isolates, however, are relatively less sensitive to inhibition
20 by recombinant CD-4 (Daar, E. et al., 1990, Proc. Natl. Acad. Sci. USA 87:6574-6579). In addition, recombinant soluble CD-4 clinical trials have produced inconclusive results (Schooley, R. et al., 1990, Ann. Int. Med. 112:247-253; Kahn, J.O. et al., 1990, Ann.
25 Int. Med. 112:254-261; Yarchoan, R. et al., 1989, Proc. Vth Int. Conf. on AIDS, p. 564, MCP 137).

The late stages of HIV replication, which involve crucial virus-specific secondary processing of certain viral proteins, have also been suggested as possible
30 anti-HIV drug targets. Late stage processing is dependent on the activity of a viral protease, and drugs are being developed which inhibit this protease (Erickson, J., 1990, Science 249:527-533). The

clinical outcome of these candidate drugs is still in question.

Attention is also being given to the development of vaccines for the treatment of HIV infection. The HIV-1 envelope proteins (gp160, gp120, gp41) have been shown to be the major antigens for anti-HIV antibodies present in AIDS patients (Barin, et al., 1985, Science 228:1094-1096). Thus far, therefore, these proteins seem to be the most promising candidates to act as antigens for anti-HIV vaccine development. To this end, several groups have begun to use various portions of gp160, gp120, and/or gp41 as immunogenic targets for the host immune system. See for example, Ivanoff, L. et al., U.S. Pat. No. 5,141,867; Saith, G. et al., WO 92/22,654; Shafferman, A., WO 91/09,872; Formoso, C. et al., WO 90/07,119. Clinical results concerning these candidate vaccines, however, still remain far in the future.

Thus, although a great deal of effort is being directed to the design and testing of anti-retroviral drugs, a truly effective, non-toxic treatment is still needed.

3. SUMMARY OF THE INVENTION

The present invention relates to DP-178 (SEQ ID:1), a 36-amino acid synthetic peptide corresponding to amino acids 638 to 673 of the transmembrane protein (TM) gp41 from the HIV-1 isolate LAI, which exhibits potent anti-HIV-1 activity. As evidenced by the example presented below, in Section 6, the DP-178 (SEQ ID:1) anti-viral activity is so high that, on a weight basis, no other known anti-HIV agent is effective at concentrations as low as those at which DP-178 (SEQ ID:1) exhibits its inhibitory effects. The invention further relates to those portions, analogs, and

homologs of DP-178 which also show such antiviral activity. The antiviral activity of such DP-178 portions, analogs, and homologs, includes, but is not limited to the inhibition of HIV transmission to uninfected CD-4⁺ cells. The invention relates to the use of DP-178 (SEQ ID:1) and DP-178 fragments and/or analogs or homologs. Such uses may include, but are not limited to, the use of the peptides as inhibitors of human and non-human retroviral, especially HIV, transmission to uninfected cells, and as type and/or subtype-specific diagnostic tools.

An embodiment of the invention is demonstrated below wherein an extremely low concentration of DP-178 (SEQ ID:1), and very low concentrations of a DP-178 homolog (SEQ ID:3) are shown to be potent inhibitors of HIV-1 mediated CD-4⁺ cell-cell fusion (i.e., syncytial formation) and infection of CD-4⁺ cells by cell-free virus. Further, it is shown that DP-178 (SEQ ID:1) is not toxic to cells, even at concentrations 3 logs higher than the inhibitory DP-178 (SEQ ID:1) concentration.

The invention also relates to analogous DP178 peptides in other enveloped viruses that demonstrate similar antiviral properties.

The invention further relates to peptides analogous to DP-107, a peptide corresponding to amino acids 558-595 of the HIV-1_{LAI} transmembrane protein (TM) of gp41, that are present in other enveloped viruses, and demonstrate antiviral properties. The present invention is based, in part, on the surprising discovery that the DP-107 and DP-108 domains of the gp41 protein non-covalently complex with each other, and that their interaction is necessary for the normal activity of the virus. The invention, therefore, further relates to methods for identifying antiviral

compounds that disrupt the interaction between DP-107 and DP-178, and/or between DP-107-like and DP-178-like peptides.

Embodiments of the invention are demonstrated, below, wherein peptides having structural and/or similarity to DP-107 and DP-178 are identified.

3.1. DEFINITIONS

Peptides are defined herein as organic compounds comprising two or more amino acids covalently joined by peptide bonds. Peptides may be referred to with respect to the number of constituent amino acids, i.e., a dipeptide contains two amino acid residues, a tripeptide contains three, etc. Peptides containing ten or fewer amino acids may be referred to as oligopeptides, while those with more than ten amino acid residues are polypeptides.

Peptide sequences defined herein are represented by one-letter symbols for amino acid residues as follows:

A (alanine)
R (arginine)
N (asparagine)
D (aspartic acid)
C (cysteine)
Q (glutamine)
E (glutamic acid)
G (glycine)
H (histidine)
I (isoleucine)
L (leucine)
K (lysine)
M (methionine)
F (phenylalanine)
P (proline)

S (serine)
T (threonine)
W (tryptophan)
Y (tyrosine)
V (valine)

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4. BRIEF DESCRIPTION OF THE FIGURES

FIG. 1. Amino acid sequence of DP-178 (SEQ ID:1) derived from HIV_{LAI}; DP-178 homologs derived from HIV-1_{SF2} (DP-185; SEQ ID:3), HIV-1_{RF} (SEQ ID:4), and
10 HIV-1_{MN} (SEQ ID:5); DP-178 homologs derived from amino acid sequences of two prototypic HIV-2 isolates, namely, HIV-2_{rod} (SEQ ID:6) and HIV-2_{NH2} (SEQ ID:7); control peptides: DP-180 (SEQ ID:2), a peptide incorporating the amino acid residues of DP-178 in a
15 scrambled sequence; DP-118 (SEQ ID:10) unrelated to DP-178, which inhibits HIV-1 cell free virus infection; DP-125 (SEQ ID:8), unrelated to DP-178, was also previously shown to inhibit HIV-1 cell free virus infection (Wild *et al.*, 1992, Proc. Natl. Acad. Sci
20 USA 89:10,537-10,541); DP-116 (SEQ ID:9), unrelated to DP-178 had previously been shown to be negative for inhibition of HIV-1 infection using the cell-free virus infection assay (Wild, *et al.*, 1992, Proc. Natl. Acad. Sci USA 89:10,537-10,541). Throughout the
25 figures, the one letter amino acid code is used.

FIG. 2. Inhibition of HIV-1 cell-free virus infection by synthetic peptides. IC50 refers to the concentration of peptide that inhibits RT production from infected cells by 50% compared to the untreated
30 control. Control: the level of RT produced by untreated cell cultures infected with the same level of virus as treated cultures.

FIG. 3. Inhibition of HIV-1 and HIV-2 cell-free virus infection by the synthetic peptide DP-178 (SEQ
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ID:1). IC50: concentration of peptide that inhibits RT production by 50% compared to the untreated control. Control: Level of RT produced by untreated cell cultures infected with the same level of virus as treated cultures.

5 FIG. 4A. Fusion Inhibition Assay. DP-178 (SEQ ID:1) inhibition of HIV-1 prototypic isolate-mediated syncytia formation. Data represents the number of virus-induced syncytia per cell.

10 FIG. 4B. Fusion Inhibition Assay. DP-180 (SEQ ID:2): scrambled control peptide. DP-185 (SEQ ID:3): DP-178 homolog derived from HIV-1_{SP2} isolate. Control: number of syncytia produced in the absence of peptide.

15 FIG. 5. Fusion inhibition assay: HIV-1 vs. HIV-2. Data represents the number of virus-induced syncytia per well. ND: not done.

 FIG. 6. Cytotoxicity study of DP-178 (SEQ ID:1) and DP-116 (SEQ ID:9) on CEM cells. Cell proliferation data is shown.

20 FIG. 7. Schematic representation of HIV-gp41 and maltose binding protein (MBP)-gp41 fusion proteins. DP107 and DP178 are synthetic peptides based on the two putative helices of gp41. The letter P in the DP107 boxes denotes an Ile to Pro mutation at amino acid number 578. Amino acid residues are
25 numbered according to Meyers et al., Human Retroviruses and AIDS, 1991, Theoret. Biol. and Biophys. Group, Los Alamos Natl. Lab., Los Alamos, NM.

30 FIG. 8. A point mutation alters the conformation and anti-HIV activity of M41.

 FIG. 9. Abrogation of DP178 anti-HIV activity. Cell fusion assays were carried out in the presence of 10 nM DP178 and various concentrations of M41Δ178 or M41PA178.

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FIG. 10. Binding of DP178 to leucine zipper of gp41 analyzed by ELISA.

FIG. 11A-B. Models for a structural transition in the HIV-1 TM protein. Two models are proposed which indicate a structural transition from a native oligomer to a fusogenic state following a trigger event (possibly gp120 binding to CD4). Common features of both models include (1) the native state is held together by noncovalent protein-protein interactions to form the heterodimer of gp120/41 and other interactions, principally through gp41 interactive sites, to form homo-oligomers on the virus surface of the gp120/41 complexes; (2) shielding of the hydrophobic fusogenic peptide at the N-terminus (F) in the native state; and (3) the leucine zipper domain (DP107) exists as a homo-oligomer coiled coil only in the fusogenic state. The major differences in the two models include the structural state (native or fusogenic) in which the DP107 and DP178 domains are complexed to each other. In the first model (A; FIG. 11A) this interaction occurs in the native state and in B during the fusogenic state. When triggered, the fusion complex in the model depicted in (A) is generated through formation of coiled-coil interactions in homologous DP107 domains resulting in an extended α -helix. This conformational change positions the fusion peptide for interaction with the cell membrane. In the second model (B; FIG. 11B), the fusogenic complex is stabilized by the association of the DP178 domain with the DP107 coiled-coil.

FIG. 12. Motif design using heptad repeat positioning of amino acids of known coiled-coils.

FIG. 13. Motif design using proposed heptad repeat positioning of amino acids of DP-107 and DP-178.

FIG. 14. Hybrid motif design crossing GCN4 and DP-107.

FIG. 15. Hybrid motif design crossing GCN4 and DP-178.

5 FIG. 16. Hybrid motif design 107x178x4, crossing DP-107 and DP-178. This motif was found to be the most consistent at identifying relevant DP-107-like and DP-178-like peptide regions.

10 FIG. 17. Hybrid motif design ALLMOTI5, crossing GCN4, DP-107, and DP-178.

FIG. 18. Hybrid motif design crossing GCN4, DP-107, DP-178, c-Fos c-Jun, c-Myc, and Flu Loop 36.

FIG. 19. Motifs designed to identify N-terminal proline-leucine zipper motifs.

15 FIG. 20. Search results for HIV-1 (BRU isolate) envelope protein gp41. Sequence search motif designations: Spades (♠): 107x178x4; Hearts (♥) ALLMOTI5; Clubs (♣): PLZIP; Diamonds (♦): transmembrane region (the putative transmembrane domains were identified using a PC/Gene program
20 designed to search for such peptide regions). Asterisk (*): Lupas method. The amino acid sequences identified by each motif are bracketed by the respective characters. Representative sequences chosen based on all searches are underlined and in
25 bold. DP-107 and DP-178 sequences are marked, and additionally double-underlined and italicized.

30 FIG. 21. Search results for human respiratory syncytial virus (RSV) strain A2 fusion glycoprotein F1. Sequence search motif designations are as in FIG. 20.

35 FIG. 22. Search results for simian immunodeficiency virus (SIV) envelope protein gp41 (AGM3 isolate). Sequence search motif designations are as in FIG. 20.

FIG. 23. Search results for canine distemper virus (strain Onderstepoort) fusion glycoprotein 1. Sequence search motif designations are as in FIG. 20.

5 FIG. 24. Search results for newcastle disease virus (strain Australia-Victoria/32) fusion glycoprotein F1. Sequence search motif designations are as in FIG. 20.

10 FIG. 25. Search results for human parainfluenza 3 virus (strain NIH 47885) fusion glycoprotein F1. Sequence search motif designations are as in FIG. 20.

15 FIG. 26. Search results for influenza A virus (strain A/AICHI/2/68) hemagglutinin precursor HA2. Sequence search designations are as in FIG. 20.

20 FIG. 27. Coiled-coil structural similarity and anti-RSV antiviral activity of 35-mer peptides synthesized utilizing the sequence of a 48-amino acid RSV F2 peptide which spans sequences identified utilizing the computer-assisted searches described herein. For the exact location and motifs utilized, see FIG. 21. "+" symbols are relative indicators of either structural similarity or antiviral activity, with a greater number of "+" symbols indicating a higher relative similarity or antiviral activity.

25 FIG. 28. Coiled-coil structural similarity and anti-RSV antiviral activity of 35-mer peptides synthesized utilizing the sequence of a 53-amino acid RSV F1 peptide which spans sequences identified utilizing the computer-assisted searches described herein. See FIG. 21 for the exact location and motifs used. "+" symbols are as described for FIG. 27.

30 FIG. 29. Coiled-coil structural similarity and anti-human parainfluenza 3 virus (HPF3) antiviral activity of 35-mer peptides synthesized utilizing the

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sequence of a 56-amino acid HPF3 peptide which spans sequences identified utilizing computer-assisted searches described herein. For the exact location and motifs utilized, see FIG. 25. "+" symbols are as described in FIG. 27.

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FIG. 30. Coiled-coil structural similarity and anti-HPF3 antiviral activity of 35-mer peptides synthesized utilizing the sequence of a 70-amino acid HPF3 peptide which spans sequences identified utilizing the computer-assisted searches described herein. For the exact location and motifs utilized, see FIG. 25. "+" symbols are as described in FIG. 27.

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5. DETAILED DESCRIPTION OF THE INVENTION

Described herein are peptides that exhibit potent antiviral activity. These peptides include DP-178 (SEQ ID:1), a gp41-derived 36 amino acid peptide, fragments and/or analogs of DP-178, and peptides which are homologous to DP-178. In addition, these peptides may include peptides exhibiting anti-viral activity which are analogous to DP-107, a 38 amino acid peptide corresponding to residues 558 to 595 of the HIV-1_{LAI} transmembrane (TM) gp41 protein, and which are present in other enveloped viral proteins. Also described here are assays for testing the antiviral activities of such peptides. The present invention is based, in part, of the surprising discovery that the DP-107 and DP-178 domains of the gp41 protein complex with each other via non-covalent protein-protein interactions which are necessary for normal activity of the virus. As such, methods are described for the identification of antiviral compounds that disrupt the interaction between DP-107 and DP-178 peptides, and between DP-107-like and DP-178-like peptides. Finally, the use of the peptides of the invention as inhibitors of non-

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human and human viral and retroviral, especially HIV, transmission are detailed, as is the use of the peptides as diagnostic indicators of the presence of specific, viruses, especially retroviruses.

5 While not limited to any theory of operation, the following model is proposed to explain the potent anti-HIV activity of DP178, based, in part, on the experiments described in the working examples, infra. In the viral protein, gp41, DP178 corresponds to a putative α -helix region located in the C-terminal end
10 of the gp41 ectodomain, and appears to associate with a distal site on gp41 whose interactive structure is influenced by the leucine zipper motif, a coiled-coil structure, referred to as DP107. The association of these two domains may reflect a molecular linkage or
15 "molecular clasp" intimately involved in the fusion process. It is of interest that mutations in the C-terminal α -helix motif of gp41 (i.e., the D178 domain) tend to enhance the fusion ability of gp41, whereas mutations in the leucine zipper region (i.e.,
20 the DP107 domain) decrease or abolish the fusion ability of the viral protein. It may be that the leucine zipper motif is involved in membrane fusion while the C-terminal α -helix motif serves as a molecular safety to regulate the availability of the
25 leucine zipper during virus-induced membrane fusion.

On the basis of the foregoing, two models are proposed of gp41-mediated membrane fusion which are schematically shown in FIG. 11A-B. The reason for
30 proposing two models is that the temporal nature of the interaction between the regions defined by DP107 and DP178 cannot, as yet, be pinpointed. Each model envisions two conformations for gp41 - one in a "native" state as it might be found on a resting virion. The other in a "fusogenic" state to reflect
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conformational changes triggered following binding of gp120 to CD4 and just prior to fusion with the target cell membrane. The strong binding affinity between gp120 and CD4 may actually represent the trigger for the fusion process obviating the need for a pH change such as occurs for viruses that fuse within intracellular vesicles. The two major features of both models are: (1) the leucine zipper sequences (DP107) in each chain of oligomeric envelope are held apart in the native state and are only allowed access to one another in the fusogenic state so as to form the extremely stable coiled-coils, and (2) association of the DP178 and DP107 sites as they exist in gp41 occur either in the native or fusogenic state. FIG. 11A depicts DP178/DP107 interaction in the native state as a molecular class. On the other hand, if one assumes that the most stable form of the envelope occurs in the fusogenic state, the model in FIG. 11B can be considered.

When synthesized as peptides, both DP107 and DP178 are potent inhibitors of HIV infection and fusion, probably by virtue of their ability to form complexes with viral gp41 and interfere with its fusogenic process; e.g., during the structural transition of the viral protein from the native structure to the fusogenic state, the DP178 and DP107 peptides may gain access to their respective binding sites on the viral gp41, and exert a disruptive influence. DP107 peptides which demonstrate anti-HIV activity are described in Applicants' co-pending application Serial No. 07/927,532, filed August 7, 1992, which is incorporated by reference herein in its entirety.

As shown in the working examples, infra, a truncated recombinant gp41 protein corresponding the

ectodomain of gp41 containing both DP107 and DP178 domains (excluding the fusion peptide, transmembrane region and cytoplasmic domain of gp41) did not inhibit HIV-1 induced fusion. However, when a single mutation was introduced to disrupt the coiled-coil structure of the DP107 domain -- a mutation which results in a total loss of biological activity of DP107 peptides -- the inactive recombinant protein was transformed to an active inhibitor of HIV-1 induced fusion. This transformation may result from liberation of the potent DP178 domain from a molecular clasp with the leucine zipper, DP107 domain.

For clarity of discussion, the invention will be described for DP178 peptide inhibitors of HIV. However, the principles may be analogously applied to other fusogenic enveloped viruses, including but not limited to those viruses containing the peptides listed in Tables V through X, below.

5.1. DP-178 AND DP-178-LIKE PEPTIDES

The peptide DP-178 (SEQ ID:1) of the invention corresponds to amino acid residues 638 to 673 of the transmembrane protein gp41 from the HIV-1_{LAI} isolate, and has the 36 amino acid sequence (reading from amino to carboxy terminus):

NH₂-YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID:1)

In addition to the full-length DP-178 (SEQ ID:1) 36-mer, the peptides of the invention may include truncations of the DP-178 (SEQ ID:1) peptide which exhibit antiviral activity. Such truncated DP-178 (SEQ ID:1) peptides may comprise peptides of between 3 and 36 amino acid residues (i.e., peptides ranging in size from a tripeptide to a 36-mer polypeptide), and

may include but are not limited to those listed in
Tables I and II, below. Peptide sequences in these
tables are listed from amino (left) to carboxy (right)
terminus. "X" may represent an amino group ($-NH_2$) and
"Z" may represent a carboxyl ($-COOH$) group.

5 Alternatively, as described below, "X" and/or "Z" may
represent a hydrophobic group, an acetyl group, a FMOC
group, an amido group, or a covalently attached
macromolecule.

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TABLE I
DP-178 (SEQ ID:1) CARBOXY TRUNCATIONS

X-YTS-Z
 X-YTSL-Z
 X-YTSLI-Z
 X-YTSLIH-Z
 5 X-YTSLIHS-Z
 X-YTSLIHSL-Z
 X-YTSLIHSLI-Z
 X-YTSLIHSLIE-Z
 X-YTSLIHSLIEE-Z
 X-YTSLIHSLIEES-Z
 X-YTSLIHSLIEESQ-Z
 10 X-YTSLIHSLIEESQN-Z
 X-YTSLIHSLIEESQNNQ-Z
 X-YTSLIHSLIEESQNNQQ-Z
 X-YTSLIHSLIEESQNNQQE-Z
 X-YTSLIHSLIEESQNNQQEK-Z
 X-YTSLIHSLIEESQNNQQEKN-Z
 X-YTSLIHSLIEESQNNQQEKNE-Z
 X-YTSLIHSLIEESQNNQQEKNEQ-Z
 15 X-YTSLIHSLIEESQNNQQEKNEQE-Z
 X-YTSLIHSLIEESQNNQQEKNEQEL-Z
 X-YTSLIHSLIEESQNNQQEKNEQELL-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLE-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLEL-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELD-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELDK-Z
 20 X-YTSLIHSLIEESQNNQQEKNEQELLELDKW-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWA-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASL-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLW-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLWN-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLWNW-Z
 X-YTSLIHSLIEESQNNQQEKNEQELLELDKWASLWNWF-Z
 25

The one letter amino acid code is used.

Additionally,

"X" may represent an amino group, a hydrophobic group,
 including but not limited to carbobenzoxyl, dansyl, or
 30 T-butyloxycarbonyl; an acetyl group; a 9-
 fluorenylmethoxy-carbonyl (Fmoc) group; a
 macromolecular carrier group including but not limited
 to lipid-fatty acid conjugates, polyethylene glycol,
 or carbohydrates.

"Z" may represent a carboxyl group; an amido group; a
 T-butyloxycarbonyl group; a macromolecular carrier
 35 group including but not limited to lipid-fatty acid
 conjugates, polyethylene glycol, or carbohydrates.

TABLE II
DP-178 (SEQ ID:1) AMINO TRUNCATIONS

	X-NWF-Z
	X-WNWF-Z
	X-LWNWF-Z
5	X-SLWNWF-Z
	X-ASLWNWF-Z
	X-WASLWNWF-Z
	X-KWASLWNWF-Z
	X-DKWASLWNWF-Z
	X-LDKWASLWNWF-Z
	X-ELDKWASLWNWF-Z
10	X-LELDKWASLWNWF-Z
	X-LLELDKWASLWNWF-Z
	X-ELLELDKWASLWNWF-Z
	X-QELLELDKWASLWNWF-Z
	X-EQELLELDKWASLWNWF-Z
	X-NEQELLELDKWASLWNWF-Z
	X-KNEQELLELDKWASLWNWF-Z
	X-EKNEQELLELDKWASLWNWF-Z
15	X-QEKNEQELLELDKWASLWNWF-Z
	X-QQEKNEQELLELDKWASLWNWF-Z
	X-NQQEKNEQELLELDKWASLWNWF-Z
	X-QNQQEKNEQELLELDKWASLWNWF-Z
	X-SQNQQEKNEQELLELDKWASLWNWF-Z
	X-ESQNQQEKNEQELLELDKWASLWNWF-Z
	X-EESQNQQEKNEQELLELDKWASLWNWF-Z
	X-IEESQNQQEKNEQELLELDKWASLWNWF-Z
20	X-LIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-SLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-HSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-IHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-LIHSIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-SLIHSIEESQNQQEKNEQELLELDKWASLWNWF-Z
	X-TSLIHSIEESQNQQEKNEQELLELDKWASLWNWF-Z
25	X-YTSLIHSIEESQNQQEKNEQELLELDKWASLWNWF-Z

The one letter amino acid code is used.

Additionally,

30 "X" may represent an amino group, a hydrophobic group, including but not limited to carbobenzoxyl, dansyl, or T-butyloxycarbonyl; an acetyl group; a 9-fluorenylmethoxy-carbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

35 "Z" may represent a carboxyl group; an amido group; a T-butyloxycarbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

The antiviral peptides of the invention also include analogs of DP-178 and/or DP-178 truncations which may include, but are not limited to, peptides comprising the DP-178 (SEQ ID:1) sequence, or DP-178 truncated sequence, containing one or more amino acid
5 substitutions, insertions and/or deletions. Analogs of DP-178 homologs, described below, are also within the scope of the invention. The DP-178 analogs of the invention exhibit antiviral activity, and may, further, possess additional advantageous features,
10 such as, for example, increased bioavailability, and/or stability, or reduced host immune recognition.

HIV-1 and HIV-2 envelope proteins are structurally distinct, but there exists a striking amino acid conservation within the DP-178-
15 corresponding regions of HIV-1 and HIV-2. The amino acid conservation is of a periodic nature, suggesting some conservation of structure and/or function. Therefore, one possible class of amino acid substitutions would include those amino acid changes
20 which are predicted to stabilize the structure of the DP-178 peptides of the invention.

Amino acid substitutions may be of a conserved or non-conserved nature. Conserved amino acid substitutions consist of replacing one or more amino
25 acids of the DP-178 (SEQ ID:1) peptide sequence with amino acids of similar charge, size, and/or hydrophobicity characteristics, such as, for example, a glutamic acid (E) to aspartic acid (D) amino acid substitution. When only conserved substitutions are
30 made, the resulting peptide is functionally equivalent to DP-178 (SEQ ID:1) or the DP-178 peptide from which it is derived. Non-conserved substitutions consist of replacing one or more amino acids of the DP-178 (SEQ
ID:1) peptide sequence with amino acids possessing
35 dissimilar charge, size, and/or hydrophobicity

characteristics, such as, for example, a glutamic acid (E) to valine (V) substitution.

5 Amino acid insertions may consist of single amino acid residues or stretches of residues ranging from 2 to 15 amino acids in length. One or more insertions may be introduced into DP-178 (SEQ ID:1), DP-178 fragments, analogs and/or DP-178 homologs (described below).

10 Deletions of DP-178 (SEQ ID:1), DP-178 fragments, analogs, and/or DP-178 homologs (described below) are also within the scope of the invention. Such deletions consist of the removal of one or more amino acids from the DP-178 or DP-178-like peptide sequence, with the lower limit length of the resulting peptide sequence being 4 to 6 amino acids. Such deletions may 15 involve a single contiguous or greater than one discrete portion of the peptide sequences.

The peptides of the invention may further include homologs of DP-178 (SEQ ID:1) and/or DP-178 truncations which exhibit antiviral activity. Such 20 DP-178 homologs are peptides whose amino acid sequences are comprised of the amino acid sequences of peptide regions of other (*i.e.*, other than HIV-1_{LAI}) viruses that correspond to the gp41 peptide region from which DP-178 (SEQ ID:1) was derived. Such 25 viruses may include, but are not limited to, other HIV-1 isolates and HIV-2 isolates. DP-178 homologs derived from the corresponding gp41 peptide region of other (*i.e.*, non HIV-1_{LAI}) HIV-1 isolates may include, for example, peptide sequences as shown below.

30 NH₂-YTNTIYTLLEESQNQQEKNEQEELLELDKWASLWNWF-COOH (DP-185; SEQ ID:3);

35 NH₂-YTGIIYNLLEESQNQQEKNEQEELLELDKWANLWNWF-COOH (SEQ ID:4);

NH₂-YTSLIYSLLEKSQIQQEKNEQELLELDKWASLWNWF-COOH (SEQ ID:5).

SEQ ID:3 (DP-185), SEQ ID:4, and SEQ ID:5 are derived from HIV-1_{SP2}, HIV-1_{RP}, and HIV-1_{MN} isolates, respectively. Underlined amino acid residues refer to those residues that differ from the corresponding position in the DP-178 (SEQ ID:1) peptide. One such DP-178 homolog, DP-185 (SEQ ID:3), is described in the Working Example presented in Section 6, below, where it is demonstrated that DP-185 (SEQ ID:3) exhibits antiviral activity. The DP-178 homologs of the invention may also include truncations, amino acid substitutions, insertions, and/or deletions, as described above.

In addition, striking similarities, as shown in FIG. 1, exist within the regions of HIV-1 and HIV-2 isolates which correspond to the DP-178 sequence. A DP-178 homolog derived from the HIV-2_{NH2} isolate has the 36 amino acid sequence (reading from amino to carboxy terminus):

NH₂-LEANISQSLEQAQIQQEKMYELQKLNSWDVFTNWL-COOH (SEQ ID:7)

Table III and Table IV show some possible truncations of the HIV-2_{NH2} DP-178 homolog, which may comprise peptides of between 3 and 36 amino acid residues (*i.e.*, peptides ranging in size from a tripeptide to a 36-mer polypeptide). Peptide sequences in these tables are listed from amino (left) to carboxy (right) terminus. "X" may represent an amino group (-NH₂) and "Z" may represent a carboxyl (-COOH) group. Alternatively, as described below, "X" and/or "Z" may represent a hydrophobic group, an acetyl group, a FMOC group, an amido group, or a covalently attached macromolecule, as described below.

TABLE III

HIV-2_{NPZ} DP-178 homolog carboxy truncations.

	X-LEA-Z
	X-LEAN-Z
	X-LEANI-Z
	X-LEANIS-Z
5	X-LEANISQ-Z
	X-LEANISQS-Z
	X-LEANISQSL-Z
	X-LEANISQSLE-Z
	X-LEANISQSLEQ-Z
	X-LEANISQSLEQA-Z
	X-LEANISQSLEQAQ-Z
10	X-LEANISQSLEQAQI-Z
	X-LEANISQSLEQAQIQ-Z
	X-LEANISQSLEQAQIQQ-Z
	X-LEANISQSLEQAQIQQE-Z
	X-LEANISQSLEQAQIQQEK-Z
	X-LEANISQSLEQAQIQQEKN-Z
	X-LEANISQSLEQAQIQQEKNM-Z
	X-LEANISQSLEQAQIQQEKNMY-Z
15	X-LEANISQSLEQAQIQQEKNMYE-Z
	X-LEANISQSLEQAQIQQEKNMYEL-Z
	X-LEANISQSLEQAQIQQEKNMYELQ-Z
	X-LEANISQSLEQAQIQQEKNMYELQK-Z
	X-LEANISQSLEQAQIQQEKNMYELQKL-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLN-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNS-Z
20	X-LEANISQSLEQAQIQQEKNMYELQKLNSW-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWD-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDV-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVF-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFT-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTN-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNW-Z
	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z

25 The one letter amino acid code is used.

Additionally,

30 "X" may represent an amino group, a hydrophobic group, including but not limited to carbobenzoxy, dansyl, or T-butyloxycarbonyl; an acetyl group; a 9-fluorenylmethoxy-carbonyl (Fmoc) group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

35 "Z" may represent a carboxyl group; an amido group; a T-butyloxycarbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

TABLE IV

HIV-2_{NH2} DP-178 homolog amino truncations.

	X-NWL-Z
	X-TNWL-Z
	X-FTNWL-Z
5	X-VFTNWL-Z
	X-DVFTNWL-Z
	X-WDVFTNWL-Z
	X-SWDVFTNWL-Z
	X-NSWDVFTNWL-Z
	X-LNSWDVFTNWL-Z
	X-KLNSWDVFTNWL-Z
10	X-QKLNSWDVFTNWL-Z
	X-LQKLNSWDVFTNWL-Z
	X-ELQKLNSWDVFTNWL-Z
	X-YELQKLNSWDVFTNWL-Z
	X-MYELQKLNSWDVFTNWL-Z
	X-NMYELQKLNSWDVFTNWL-Z
	X-KNMYELQKLNSWDVFTNWL-Z
	X-EKNMYELQKLNSWDVFTNWL-Z
15	X-QEKNMYELQKLNSWDVFTNWL-Z
	X-QQEKNMYELQKLNSWDVFTNWL-Z
	X-IQQEKNMYELQKLNSWDVFTNWL-Z
	X-QIQQEKNMYELQKLNSWDVFTNWL-Z
	X-AQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-QAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-EQAQIQQEKNMYELQKLNSWDVFTNWL-Z
20	X-LEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-SLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-QSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-ISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-NISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-ANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
	X-EANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z
25	X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z

The one letter amino acid code is used.

Additionally,

30 "X" may represent an amino group, a hydrophobic group, including but not limited to carbobenzoxyl, dansyl, or T-butyloxycarbonyl; an acetyl group; a 9-fluorenylmethoxy-carbonyl (Fmoc) group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

35 "Z" may represent a carboxyl group; an amido group; a T-butyloxycarbonyl group; a macromolecular carrier group including but not limited to lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates.

5.2. DP-107 and DP-178 ANALOGOUS
ANTIVIRAL PEPTIDES

Peptide sequences functionally corresponding, and thus analogous to, the DP-178 sequences of the invention, described, above, in Section 5.1 may be found in other, non-HIV-1 envelope viruses. Further, peptide sequences functionally corresponding, and thus analogous to, DP-107, an HIV-1-derived antiviral peptide, may also be found in other, non-HIV-1 envelope viruses. DP-107 is a 38 amino acid peptide corresponding to residues 558 to 595 of HIV-1_{LAI} transmembrane (TM) gp41 protein, which exhibits potent anti-viral activity. DP-107 is more fully described in Applicant's co-pending U.S. Patent Application Ser. No. 07/927,532. These DP-107-like and DP-178-like analogous peptides and present in TM proteins of envelope viruses and preferably exhibit antiviral activity, most preferably antiviral activity which is specific to the virus in which their native sequences are found.

DP-107-like and DP-178-like peptides may be identified, for example, by utilizing a computer-assisted search strategy such as that described and demonstrated, below, in the Examples presented in Sections 9 through 16. The search strategy identifies regions in other viruses that are similar in predicted secondary structure to DP-107 and DP-178.

This search strategy is described fully, below, in the Example presented in Section 9. While this search strategy is based, in part, on a primary amino acid motif deduced from DP-107 and DP-178, it is not based solely on searching for primary amino acid sequence homologies, as such protein sequence homologies exist within, but not between major groups of viruses. For example, primary amino acid sequence homology is high within the TM protein of different

strains of HIV-1 or within the TM protein of different isolates of simian immunodeficiency virus (SIV).

Primary amino acid sequence homology between HIV-1 and SIV, however, is low enough so as not to be useful.

5 It is not possible, therefore, to find DP-107 or DP-178-like peptides within other viruses, whether structurally, or otherwise, based on primary sequence homology, alone.

Further, while it would be potentially useful to identify primary sequence arrangements of amino acids
10 based on the physical chemical characteristics of different classes of amino acids rather than based on the specific amino acids themselves, for instance, a by concentrating on the coiled-coil nature of the peptide sequence, a computer algorithm designed by
15 Lupas et al. to identify such coiled-coil propensities of regions within proteins (Lupas, A., et al., 1991 Science 252:1162-1164) is inadequate for identifying protein regions analogous to DP-107 or DP-178.

Specifically, analysis of HIV-1 gp160 (containing
20 both gp120 and gp41) using the Lupas algorithm does not identify the coiled-coil region within DP-107. It does, however, identify a region within DP-178 beginning eight amino acids N-terminal to the start of DP-178 and ending eight amino acids from the C-
25 terminus. The DP-107 peptide has been shown experimentally to form a stable coiled coil. A search based on the Lupas search algorithm, therefore, would not have identified the DP-107 coiled-coil region. Conversely, the Lupas algorithm identified the DP-178
30 region as a potential coiled-coil motif. However, the peptide DP-178 derived from this region failed to form a coiled coil in solution. A possible explanation for the inability of the Lupas search algorithm to
35 accurately identify coiled-coil sequences within the HIV-1 TM, is that the Lupas algorithm is based on the

structure of coiled coils from proteins that are not structurally or functionally similar to the TM proteins of viruses, antiviral peptides (e.g. DP-107 and DP-178) of which are an object of this invention.

5 The computer search strategy of the invention, as demonstrated in the Examples presented below, in Sections 9 through 16, successfully identifies regions of viral TM proteins similar to DP-107 or DP-178. This search strategy was designed to be used with a commercially-available sequence database packages,
10 preferably PC/Gene. A series of motifs were designed and engineered to range in stringency from very strict to very broad, as discussed in Section 9.

Among the protein sequence search motifs which may be utilized in such a computer-assisted DP-107-like
15 and DP-178-like antiviral peptide search are the 107x178x4 motif, the ALLMOTI5 motif, and the PLZIP series of motifs, each of which is described in the Example presented in Section 9, below, with 107x178x4 being preferred.

20 Coiled-coiled sequences are thought to consist of heptad amino acid repeats. For ease of description, the amino acid positions within the heptad repeats are sometimes referred to as A through G, with the first position being A, the second B, etc. The motifs used
25 to identify DP-107-like and DP-178-like sequences herein are desined to specifically search for and identify such heptad repeats. In the descriptions of each of the motifs described, below, amino acids enclosed by brackets , i.e., [], designate the only
30 amino acid residues that are acceptable at the given position, while amino acids enclosed by braces, i.e., {}, designate the only amino acids which are unacceptable at the given heptad position. When a set of bracketed or braced amino acids is followed by a
35 number in parentheses i.e., (), it refers to the

number of subsequent amino acid positions for which the designated set of amino acids hold, e.g, a (2) means "for the next two heptad amino acid positions.

The ALLMOTI5 is written as follows:

5 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-
 {CDGHP}-{CFP}(2)-{CDGHP}-{CFP}(3)-

Translating this motif, it would read: "at the first (A) position of the heptad, any amino acid
 10 residue except C, D, G, H, or P is acceptable, at the next two (B,C) amino acid positions, any amino acid residue except C, F, or P is acceptable, at the fourth heptad position (D), any amino acid residue except C,
 D, G, H, or P is acceptable, at the next three (E, F,
 15 G) amino acid positions, any amino acid residue except C, F, or P is acceptable. This motif is designed to search for five consecutive heptad repeats (thus the repeat of the first line five times), meaning that it searches for 35-mer sized peptides. It may also be
 20 designed to search for 28-mers, by only repeating the initial motif four times. With respect to the ALLMOTI5 motif, a 35-mer search is preferred. Those viral sequences identified via such an ALLMOTI5 motif are listed in Table V, below, at the end of this
 25 Section. The viral sequences listed in Table V potentially exhibit antiviral activity, may be useful in the the identification of antiviral compounds, and are intended to be within the scope of the invention.

The 107x178x4 motif is written as follows:

30 [EFIKLNQSTVWY]-{CFMP}(2)-[EFIKLNQSTVWY]-{CFMP}(3)-
 [EFIKLNQSTVWY]-{CFMP}(2)-[EFIKLNQSTVWY]-{CFMP}(3)-
 [EFIKLNQSTVWY]-{CFMP}(2)-[EFIKLNQSTVWY]-{CFMP}(3)-
 [EFIKLNQSTVWY]-{CFMP}(2)-[EFIKLNQSTVWY]-{CFMP}(3)-

Translating this motif, it would read: "at the first (A) position of the heptad, any amino acid
 35 residue except E, F, I, K, L, N, Q, S, T, V, W, or Y

is acceptable, at the next two (B,C) amino acid positions, any amino acid residue except C, F, M or P is acceptable, at the fourth position (D), any amino acid residue except E, F, I, K, L, N, Q, S, T, V, W, or Y is acceptable, at the next three (E, F, G) amino acid positions, any amino acid residue except C, F, M or P is acceptable. This motif is designed to search for four consecutive heptad repeats (thus the repeat of the first line four times), meaning that it searches for 28-mer sized peptides. It may also be designed to search for 35-mers, by repeating the initial motif five times. With respect to the 107x178x4 motif, a 28-mer search is preferred. Those viral sequences identified via such a 107x178x4 motif are listed in Table V, below, at the end of this Section. The viral sequences listed in Table V potentially exhibit antiviral activity, may be useful in the the identification of antiviral compounds, and are intended to be within the scope of the invention.

The PLZIP series of motifs are as listed in FIG. 19. These motifs are designed to identify leucine zipper coiled-coil like heptads wherein at least one proline residue is present at some predefined distance N-terminal to the repeat. These PLZIP motifs find regions of proteins with similarities to HIV-1 DP-178 generally located just N-terminal to the transmembrane anchor. These motifs may be translated according to the same convention described above. Each line depicted in FIG. 19 represents a single, complete search motif. "X" in these motifs refers to any amino acid residue. In instances wherein a motif contains two numbers within parentheses, this refers to a variable number of amino acid residues. For example, X (1,12) is translated to "the next one to twelve amino acid residues, inclusive, may be any amino acid".

Tables VI through X, below, at the end of this

Section, list hits from such PLZIP motifs. The viral sequences listed in Table VI through X potentially exhibit antiviral activity, may be useful in the the identification of antiviral compounds, and are intended to be within the scope of the invention.

5 The Examples presented in Sections 17 and 18, below, demonstrate that respiratory syncytial virus and parainfluenza virus sequences identified via such a computer search exhibit antiviral and/or structural characteristics similar to those of DP-107 or DP-178.

10 The DP-107-like and DP-178-like analogous peptides may, further, contain any of the additional groups described for DP-178, above, in Section 5.1. For example, these peptides may include any of the additional amino-terminal groups which "X" of Tables I
15 through IV may represent, and may also include any of the carboxy-terminal groups which "Z" of Tables I through IV may represent.

 Additionally, such DP-107-like and DP-178-like peptides may further include DP-107-like or DP-178-like
20 peptides, such as those listed in Tables V through X, above, containing one or more amino acid substitutions, insertions, and/or deletions. Also, analogs of such DP-107-like and DP-178-like peptides are intended to be within the scope of the invention.
25 Such analogs of the invention may exhibit increased antiviral activity, and may, further, possess increased bioavailability, and/or stability, or reduced immune recognition.

 The DP-107-like and DP-178-like amino acid
30 substitutions, insertions and deletions, are as described for DP-178, above, in Section 5.1. Analog modifications are as described, below, in Section 5.3.

35

TABLE V

Search Results Summary for 107x178x4 and
ALLMOTI5 Motifs

[illegible]

PENV HV1H3	545-584	631-683	701-818			PENV HV1BN	601-580	608-708	763-831		
PENV HV1J3	558-605	642-684	802-829			PENV HV1BR	510-588	615-717	772-841		
PENV HV1JR		622-675	783-811			PENV HV1C4	510-808	626-724	778-855		
PENV HV1KB	555-588	637-677	778-824			PENV HV1EL	502-581	607-708	768-829		
PENV HV1MA	547-585	633-707	794-826			PENV HV1H2	505-584	610-712	767-838		
PENV HV1MF	543-582	628-681	788-816			PENV HV1H3	505-584	610-712	767-843		
PENV HV1MN	567-585	632-684	791-819			PENV HV1J3	517-805	622-723	778-843		
PENV HV1ND	538-583	621-673	783-813			PENV HV1JR	487-588	603-704	759-835		
PENV HV1OV	544-583	630-704	799-820			PENV HV1KB	511-545	656-589	818-718	772-848	
PENV HV1PV	545-584	631-683	791-818			PENV HV1MA	507-588	617-714	770-825		
PENV HV1RH	554-602	640-692	800-832			PENV HV1MF	503-582	622-710	768-841		
PENV HV1S1	538-585	622-674	782-809			PENV HV1MN	508-585	617-713	774-841		
PENV HV1S3	541-589	627-679	787-815			PENV HV1ND	486-584	601-702	767-825		
PENV HV1SC	545-583	631-683				PENV HV1OV	487-583	610-711	768-842		
PENV HV1W1	545-583	631-683	791-818			PENV HV1PV	505-584	610-712	767-843		
PENV HV1W2	538-584	622-674	782-809			PENV HV1RH	507-603	618-721	776-852		
PENV HV1Z2	542-581	628-680	790-820			PENV HV1S1	489-585	602-703	758-830		
PENV HV1Z8	545-583	630-682	782-822			PENV HV1S3	484-580	607-708	763-837		
PENV HV1ZH	573-601	634-678	787-828			PENV HV1SC	489-584	611-712	767-834		
PENV HV1ZH	545-584	627-668	781-823			PENV HV1W1	488-584	611-712	767-838		
PENV HV2BE	532-581	621-648	655-688			PENV HV1W2	489-584	602-703	758-827		
PENV HV2CA	534-583	623-650	655-688			PENV HV1Z2	502-581	607-709	764-831		
PENV HV2D1	523-580	555-582	644-688			PENV HV1Z8	504-583	608-711	768-840		
PENV HV2Q1	524-581	556-583	613-640	645-693		PENV HV1Z8	512-601	617-675	682-719	774-831	
PENV HV2NZ	524-581	556-583	613-640	682-688		PENV HV1ZH	522-584	612-712	777-839		
PENV HV2RO	533-582	622-688				PENV HV2BE	510-585	617-680			
PENV HV2R2	527-584	559-588	648-682			PENV HV2CA	512-587	618-709			
PENV HV2SB	557-584	614-673				PENV HV2D1	501-586	608-688			
PENV HV2ST	527-584	559-588	648-682			PENV HV2Q1	502-587	608-688			
PENV MCF3	473-512					PENV HV2NZ	488-587	608-688			
PENV MCF3	488-516					PENV HV2RO	511-588	618-708			
PENV MLVAV	517-544					PENV HV2B2	505-580	612-702			
PENV MLVCB	510-538					PENV HV2SB	528-588	614-700			
PENV MLVFS	523-583					PENV HV2ST	505-580	612-702			
PENV MLVFF	523-583					PENV PHAE	387-422	485-527			
PENV MLVFP	523-583					PENV JBRV	403-455	571-605			
PENV MLVHO	510-540					PENV MCF3	473-525	537-571			
PENV MLVK1	40-81					PENV MCF3	474-528	538-572			
PENV MLVMO	502-543					PENV MLVAV	503-555	587-601			
PENV MLVRD	487-538					PENV MLVCB	488-550	582-585			
PENV MLVRK	487-538					PENV MLVFS	520-584	576-610			
PENV MMTV8	458-485	582-589				PENV MLVFF	520-584	576-610			
PENV MMTVG	458-485	582-589				PENV MLVFP	520-584	576-610			
PENV MPNV	422-470					PENV MLVHO	504-551	583-587			
PENV M8VFB	57-84					PENV MLVK1	40-82	104-138			
PENV DMVV8	42-68	186-223	780-807			PENV MLVMO	502-554	586-600			
PENV RMCFV	487-517				PENV MLVRD	487-548	581-585			

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	PHEMA IADU3	387-463					PHEMA IADH2	384-440				
	PHEMA IAEN7	387-463					PHEMA IADH3	384-440				
	PHEMA IAFPR	384-442					PHEMA IADH4	384-440				
	PHEMA IAGRE	381-461					PHEMA IADH5	384-440				
	PHEMA IAGU2	505-532					PHEMA IADH8	384-440				
	PHEMA IAGUA	504-631					PHEMA IADH7	384-440				
	PHEMA IAHAL	386-462					PHEMA IADIR	379-471	509-551			
	PHEMA IACH6	388-467					PHEMA IADM1	21-55				
	PHEMA IACH7	388-467					PHEMA IADM2	380-458				
	PHEMA IAHCD	388-467					PHEMA IADNY	21-55				
	PHEMA IAHDE	388-467					PHEMA IADNZ	378-464				
	PHEMA IAHFO	388-462					PHEMA IADU1	21-55				
	PHEMA IAHK0	388-462					PHEMA IADU3	380-458				
	PHEMA IAHK7	388-462					PHEMA IAEN7	380-458				
	PHEMA IAHLE	388-467					PHEMA IAFPR	377-477				
	PHEMA IAHLO	388-467					PHEMA IAGRE	378-454				
	PHEMA IAHMI	388-462					PHEMA IAGU2	378-473				
	PHEMA IAHNM	388-462					PHEMA IAGUA	377-478				
	PHEMA IAHNN	388-467					PHEMA IAHAL	379-455				
	PHEMA IAHNR	388-467					PHEMA IAHCB	112-148	380-484	503-537		
	PHEMA IAHRO	388-462					PHEMA IAHCT	112-148	380-484	503-537		
	PHEMA IAHSA	388-462					PHEMA IAHCD	380-484	503-537			
	PHEMA IAHSP	388-467					PHEMA IAHDE	380-484	503-537			
	PHEMA IAHSW	388-467					PHEMA IAHFO	378-455				
	PHEMA IAHTE	388-462					PHEMA IAHKB	379-455				
	PHEMA IAHTO	388-466					PHEMA IAHKT	378-455				
	PHEMA IAHUR	388-462					PHEMA IAHLE	112-148	380-484	503-537		
	PHEMA IAKIE	426-478					PHEMA IAHLO	112-148	380-484	503-537		
	PHEMA IALEN	426-478					PHEMA IAHMI	379-455				
	PHEMA IAMAA	380-460					PHEMA IAHNM	378-455				
	PHEMA IAMAB	385-468					PHEMA IAHNN	112-148	380-484	503-537		
	PHEMA IAMAO	387-463					PHEMA IAHNR	112-148	380-484	503-537		
	PHEMA IAME1	387-463					PHEMA IAHRO	379-455				
	PHEMA IAME2	387-463					PHEMA IAHBA	378-455				
	PHEMA IAME6	371-437					PHEMA IAHBP	112-148	380-484	503-537		
	PHEMA IAMIN	382-441					PHEMA IAHBW	112-148	380-484	503-537		
	PHEMA IANT6	387-463					PHEMA IAHTE	378-455				
	PHEMA IAPIL	506-534					PHEMA IAHTO	378-455				
	PHEMA IAPIE	426-478					PHEMA IAHUR	378-455				
	PHEMA IARUD	381-461					PHEMA IAJAP	378-487	502-547			
	PHEMA IASE2	381-461					PHEMA IAKIE	378-478	508-541			
	PHEMA IASH2	508-647					PHEMA IALEN	378-478	508-648			
	PHEMA IASTA	384-443					PHEMA IAMAA	377-463				
	PHEMA IATKI	416-446					PHEMA IAMAB	382-468				
	PHEMA IATKM	381-461					PHEMA IAMAO	380-468				
	PHEMA IATKP	507-634					PHEMA IAME1	380-466				
	PHEMA IATPP	493-539				PHEMA IAME2	380-468				

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PHEMA P1HW	79-110	388-393					PHEMA INC2	471-559				
PHEMA P1B	68-83						PHEMA INC3	471-559				
PHEMA P1H4	27-61						PHEMA INC4	471-559				
PHEMA P1HA	27-61						PHEMA INC5A	471-559				
PHEMA P1HT	27-76						PHEMA MEABE	49-80				
PHEMA P1HU	23-70						PHEMA MEABH	49-80				
PHEMA P1HV	27-61						PHEMA MEASI	49-87				
PHEMA P1HW	27-61						PHEMA MEABY	49-87				
PHEMA P1HX	27-61						PHEMA MUMPM	34-86				
PHEMA RACVI	186-214	256-283					PHEMA MUMPR	34-86				
PHEMA SEND5	79-106						PHEMA MUMPS	34-86				
PHEMA SENDF	79-106						PHEMA NDVA	8-52	477-628			
PHEMA SENDH	79-106						PHEMA NDVB	1-49				
PHEMA SENDJ	79-106						PHEMA NDVD	1-49				
PHEMA SENDZ	79-106						PHEMA NDVM	1-49				
PHEMA 6V41	22-62	394-421					PHEMA NDVQ	1-49				
PHEMA VACCC	119-149	175-202	216-243				PHEMA NDVTG	1-49				
PHEMA VACCI	109-148	175-202	216-243				PHEMA NDVU	1-49				
PHEMA VACCT	119-148	175-202	216-243				PHEMA PHODV	39-73				
PHEMA VACCV	109-148	175-202	216-242				PHEMA PITHW	66-110				
PVENV DHV11	318-368						PHEMA P1ZH	247-281				
PVENV EAV	120-147						PHEMA P1ZHT	247-281				
PVENV THOGV	313-347						PHEMA P1B	39-93				
PVF03 VACCC	71-110	186-212					PHEMA P1H4	13-110	394-428			
PVF03 VACCV	71-110	186-212					PHEMA P1HA	20-110	394-428			
PVF05 VACCP	33-60						PHEMA P1HT	13-110	394-428			
PVF05 VACCV	33-60						PHEMA P1HU	13-110	394-428			
PVF11 VACCC	274-321						PHEMA P1HV	13-110	394-428			
PVF11 VACCP	270-317						PHEMA P1HW	13-110	394-428			
PVF12 VACCC	10-37	113-140	554-581				PHEMA P1HX	13-110	394-428			
PVF12 VACCP	10-37	113-140	554-581				PHEMA P1HA	54-88				
PVF18 VACCC	36-82	152-178					PHEMA RACVI	186-214	256-280			
PVF18 VACCP	36-82	152-178					PHEMA RINDK	49-87				
PVFP4 FOWPV	148-173						PHEMA RINDL	49-87	191-226			
PVFUS OFNZ	59-86						PHEMA SEND5	57-110				
PVFUS VACCC	37-84						PHEMA SENDF	57-110				
PVFUS VACCV	37-84						PHEMA SENDH	57-110				
PVG01 VACCC	226-252	301-336					PHEMA SENDJ	57-110				
PVG01 VACCV	184-181	240-274					PHEMA SENDZ	57-110				
PVG01 VARV	226-252	301-336					PHEMA BV41	19-62	387-421			
PVG02 VACCV	89-123						PHEMA BV6	27-82				
PVG02 VARV	89-123						PHEMA BV6LN	27-82				
PVG03 HSEB	148-176						PVENV BEV	186-229				
PVG03 HSEK	148-176						PVENV DHV11	318-368				
PVG05 VACCC	48-76	131-161	226-289	356-389			PVENV MCV1	252-289				
PVG05 VARV	48-76	124-161	226-289	356-389			PVENV MCV2	252-289				
PVG07 H8V11	71-98						PVENV THOGV	313-354				

PVGL2 CVH22	770-787	808-875	1056-1112				PVG1 SPV4	287-321				
PVGL2 CVM4	843-884	1030-1082					PVG22 HSVI1	117-168	437-828	680-882	888-1056	
PVGL2 CVM45	39-63	581-832	978-1040				PVG24 HSVI1	7-72	74-108			
PVGL2 CVMJH	502-543	889-851					PVG27 HSVI1	184-219				
PVGL2 CVPF3	69-110	882-733	1072-1145	1353-1380			PVG28 HSVI1	253-280				
PVGL2 CVPF4	89-107	880-731	1087-1143	1351-1387			PVG2R AMEPV	28-83	184-218			
PVGL2 CVPF8	488-508	845-821	1128-1165				PVG2 SPV1R	222-258	285-326			
PVGL2 CVPF8M	488-508	845-821	1128-1165				PVG2 BPV4	255-310				
PVGL2 EBV	68-102						PVG33 HSVI1	148-189				
PVGL2 FIPV	189-233	454-481	708-738	1072-1148	1356-1392		PVG34 HSVI1	345-378				
PVGL2 IBV6	808-836	878-903	1057-1091				PVG36 HSVI1	17-90				
PVGL2 IBV8	808-836	878-902	1056-1090				PVG37 HSVI1	435-472				
PVGL2 IBVD2	808-836	878-903	1057-1091				PVG38 HSVI1	84-118				
PVGL2 IBVK	808-835	875-902	1056-1090				PVG39 HSVI1	124-158	286-300			
PVGL2 IBVM	808-835	875-902	1056-1090				PVG3 SPV1R	8-49	182-196	203-244		
PVGLB EBV	56-122	631-658					PVG3 SPV4	6-54	87-121			
PVGLB HCMVA	25-88	387-424	440-487	851-878			PVG43 HSVI1	116-150	282-286	324-381	843-877	
PVGLB HCMVT	50-88	397-424	435-482	852-878			PVG45 HSVSA	121-162				
PVGLB HSB1	427-464						PVG48 HSVI1	46-88	938-1078	1251-1321		
PVGLB HSB2	447-474						PVG48 HSVI1	168-207				
PVGLB HSB3C	428-463						PVG48 HSBVA	360-417	911-988	733-787		
PVGLB HSBV1	443-470	934-961					PVG49 HSBVA	68-102				
PVGLB HSBV4	486-513	918-843					PVG4R AMEPV	4-38				
PVGLB HSBV4	443-470	934-961					PVG4 SPV4	88-130				
PVGLB HSBV8	443-470	934-961					PVG51 HSVI1	34-73	89-123			
PVGLB HSBV8	443-470	933-960					PVG51 HSBVA	28-70	123-157	162-186		
PVGLB HSBVMD	93-120	362-378					PVG53 HSVI1	87-127				
PVGLB MCVS	381-408	441-475					PVG54 HSVI1	355-386				
PVGLC HSB11	489-510						PVG55 HSVI1	101-135				
PVGLC HSBV1K	489-510						PVG55 HSBVA	126-178				
PVGLC HSBV8	124-151						PVG56 HSVI1	151-182	678-812	844-878	750-784	846-880
PVGLC HSBV8	63-97						PVG58 HSVI1	10-72	88-123			
PVGLC HSBV8M	63-97						PVG59 HSBVA	188-209				
PVGLC HSBV8M	63-97						PVG5 SPV1R	65-103				
PVGLC HSBV8M	295-322						PVG51 HSVI1	286-289				
PVGLC HSBV8	295-322						PVG53 HSVI1	546-584				
PVGLC HSBV2	111-148						PVG55 HSBVA	805-839	1213-1254			
PVGLF BRSVA	38-65	154-202	216-243	442-489	488-531		PVG58 HSVI1	154-188	328-410			
PVGLF BRSVC	38-65	154-202	216-243	444-471	488-533		PVG57 HSVI1	378-413	601-546	1321-1369	1478-1541	
PVGLF BRSVR	38-65	154-202	216-243	444-471	488-533		PVG58 HSBVA	245-288				
PVGLF CDVO	252-283	340-387					PVG72 HSVI1	447-484	723-767	912-949		
PVGLF HRSV1	38-65	154-203	442-471	488-515			PVG75 HSBVA	271-306	388-422			
PVGLF HRSVA	38-65	154-202	213-243	488-518			PVG8 SPV1R	6-51				
PVGLF HRSVL	38-65	154-202	216-243	444-471	488-515		PVG81 HSBVA	142-178	1233-1267	2118-2156	3388-3424	3517-3558
PVGLF HRSVR	38-65	154-202	213-243	442-471	488-518		PVG83 HCMVA	10-44				
PVGLF MEA5E	228-282						PVG82 CVBF	842-878	850-885	893-1088	1263-1305	
PVGLF MEA6I	231-286						PVG82 CVBL9	850-885	893-1109	1283-1306		

PVGLF MEARY	228-282					PVGL2 CVBLY	642-878	850-885	893-1109	1283-1305		
PVGLF MUMPM	20-54	447-488				PVGL2 CVBVM	642-878	850-885	893-1109	1283-1305		
PVGLF MUMPR	20-54	447-488				PVGL2 CVBQ	642-878	850-885	893-1109	1283-1305		
PVGLF MUMPS	151-178	428-511				PVGL2 CVBVB	642-878	850-885	893-1109	1283-1305		
PVGLF NDVA	151-178	428-512				PVGL2 CVH22	770-918	1055-1112				
PVGLF NDVB	151-178	428-512				PVGL2 CVH4	643-884	1001-1117	1270-1315			
PVGLF NDVI	151-178	428-512				PVGL2 CVMA5	581-832	848-1078	1218-1263			
PVGLF NDVM	151-178	428-512				PVGL2 CVMAH	502-843	890-878	1128-1174			
PVGLF NDVT	151-178	428-512				PVGL2 CVPF5	68-110	448-482	892-733	889-923	1040-1188	1352-1388
PVGLF NDVTG	151-178	428-512				PVGL2 CVPFU	68-110	448-480	890-731	887-921	1038-1184	1351-1387
PVGLF NDVU	151-178	428-512				PVGL2 CVPFR	224-258	488-509	895-898	818-882	1128-1185	
PVGLF PHODV	38-63	221-282	308-338			PVGL2 CVPFRM	224-258	488-508	895-898	818-882	1128-1185	
PVGLF PIHC	147-174	210-268				PVGL2 EBV	68-102					
PVGLF PI2H	80-117	141-175	238-268	483-528		PVGL2 FIPV	188-245	451-485	895-738	892-928	1043-1188	1355-1382
PVGLF PI2HG	80-117	141-175	238-268	483-528		PVGL2 IBV8	781-805	1057-1091				
PVGLF PI2HT	80-117	141-175	238-268	483-528		PVGL2 IBVB	437-478	772-804	1058-1080			
PVGLF PI3B	115-182	207-241	458-487			PVGL2 IBVD2	773-805	1057-1091				
PVGLF PI3H4	115-182	207-241	457-487			PVGL2 IBVK	437-478	772-804	1058-1080			
PVGLF RINDK	224-265	458-508				PVGL2 IBVM	437-478	772-804	1058-1080			
PVGLF RINDL	224-265	458-508				PVGLB HCMVA	43-88	128-182	438-484	844-878		
PVGLF SEND5	122-149	211-245	480-507			PVGLB HCMVT	22-88	128-182	437-485	845-878		
PVGLF SENDF	122-149	211-245	480-507			PVGLB HSBV1	828-880					
PVGLF SENDH	122-149	211-245	480-507			PVGLB HSBVF	827-889					
PVGLF SENDJ	122-149	211-245	480-507			PVGLB HSBVK	827-889					
PVGLF SENDZ	122-149	211-245	480-507			PVGLB HSBVP	828-880					
PVGLF SV41	144-185	241-269	458-488			PVGLB HSBV23	828-880					
PVGLF SV5	137-171	417-444				PVGLB HSBV2H	828-880					
PVGLF TRTV	124-161	183-200	457-484			PVGLB HSBV25	817-871					
PVGLG BEFV	823-857					PVGLB HSBV8U	37-71	185-223				
PVGLG BR5VC	82-123					PVGLB HSBV81	858-913					
PVGLG HR5V1	83-83					PVGLB HSBV2	440-474	848-892				
PVGLG HR5V4	88-107					PVGLB HSBVC	883-900					
PVGLG HR5V5	243-273					PVGLB HSBVE1	542-578	911-981				
PVGLG HR5V8	88-93					PVGLB HSBVE4	474-515	847-890				
PVGLG HSBVE4	271-288					PVGLB HSBVEA	542-578	911-981				
PVGLG HSBVEB	383-410					PVGLB HSBVEB	542-578	911-981				
PVGLG RABVT	488-518					PVGLB HSBVEL	542-578	910-980				
PVGLG V5VIG	472-498					PVGLB HSBVMD	380-435	648-683				
PVGLH EBV	548-578	818-848				PVGLB HSBVSA	240-288	408-447				
PVGLH HCMVA	107-136	270-297				PVGLB HCMV8	208-280	427-475	883-778	880-884		
PVGLH HCMVT	108-135					PVGLB PRVIF	847-881					
PVGLH HSBV8	82-88	388-403				PVGLB VZVD	82-133	588-630	808-887			
PVGLH HSBVA	388-415					PVGLC HSBV11	488-510					
PVGLH HCMVA	47-111					PVGLC HSBVK	488-510					
PVGLM BUNGE	512-548	814-941	1128-1255			PVGLC HSBV2	442-478					
PVGLM BUNL7	813-950					PVGLC HSBV23	443-477					
PVGLM BUNYW	340-374	504-535	682-708			PVGLC HSBVC	235-288					

PVGLM DUGBV	945-872						PVGLC HSEVB	182-218				
PVGLM HANTB	73-100	883-720					PVGLC HSEVB	83-87				
PVGLM HANTH	75-102						PVGLC HSEVB	82-98				
PVGLM HANTL	75-102						PVGLC HSEVB	83-87				
PVGLM HANTV	75-102						PVGLC HSEVB	183-235				
PVGLM PHV	88-88						PVGLC HSEVB	280-321				
PVGLM PUMH	72-110						PVGLC HSEVB	280-321				
PVGLM PUMS	72-110						PVGLC HSEVB	88-123				
PVGLM SEOUR	73-100	813-540	884-721				PVGLD HSEVB	138-173				
PVGLM SEOUR	73-100	513-540	884-721				PVGLD HSEVB	138-173				
PVGLM BEFV	623-584						PVGLD HSEVB	111-145				
PVGLP BEV	48-82	1145-1178	1184-1211	1506-1532			PVGLD HSEVB	111-159				
PVGLX HSEVB	17-44	413-444					PVGLF HSEVB	146-202	504-545			
PVGLX PRVRI	427-481						PVGLF HSEVB	146-202	267-302	508-547		
PVGLY JUNIN	14-41						PVGLF HSEVB	146-202	267-302	508-554		
PVGLY LA66Q	88-113						PVGLF HSEVB	228-287	340-381	588-602		
PVGLY MOREI	88-113	310-346					PVGLF HSEVB	116-203	267-302	508-549		
PVGLY PIARV	334-376						PVGLF HSEVB	116-202	267-302	508-549		
PVGLY TACV	108-138	315-350					PVGLF HSEVB	116-202	267-302	508-547		
PVGLY TACV6	303-338						PVGLF HSEVB	116-202	267-302	508-549		
PVGLY TACV7	302-337						PVGLF HSEVB	116-184	228-268	452-500		
PVGLY TACV7	303-338						PVGLF HSEVB	119-187	231-272	455-503		
PVGLZ HSEVB	17-44						PVGLF HSEVB	116-184	228-268	452-500		
PVGNM BPMV	403-430						PVGLF HSEVB	20-54	103-178	235-272	447-502	
PVGNM CP5MV	182-221						PVGLF HSEVB	20-54	103-178	235-272	447-502	
PVGPB BEV	104-148						PVGLF HSEVB	20-54	103-178	235-272	447-502	
PVM1 REOVL	200-317						PVGLF HSEVB	117-182	231-272	428-512		
PVM21 REOVD	825-882						PVGLF HSEVB	122-182	231-272	428-517		
PVM22 REOVD	824-881						PVGLF HSEVB	133-182	238-272	428-517		
PVM2 REOVJ	824-881						PVGLF HSEVB	117-182	231-272	428-512		
PVM3 REOVD	188-188	343-370	458-483	631-680			PVGLF HSEVB	117-182	231-272	428-517		
PVMA2 BR6VA	124-152						PVGLF HSEVB	122-182	231-272	428-517		
PVMA2 HRSVA	124-151						PVGLF HSEVB	122-182	231-272	428-512		
PVMAT BR6VA	218-248						PVGLF HSEVB	28-63	187-266	308-350	533-581	
PVMAT HRSVA	218-248						PVGLF HSEVB	123-174	207-267	458-503		
PVMAT INCJJ	151-185						PVGLF HSEVB	83-183	477-528			
PVMAT NDVA	247-274						PVGLF HSEVB	83-183	477-528			
PVMAT PI2HT	88-123						PVGLF HSEVB	83-183	477-528			
PVMAT PI3B	201-231						PVGLF HSEVB	117-182	207-241	458-518		
PVMAT PI3H4	201-231						PVGLF HSEVB	117-182	207-241	482-532		
PVMAT SV41	323-353						PVGLF HSEVB	112-180	224-265	448-483		
PVME1 CVBM	176-209						PVGLF HSEVB	112-180	224-265	448-508		
PVME1 CVTKE	176-209						PVGLF HSEVB	127-188	211-271	483-533		
PVME1 IBV6	21-48	184-218					PVGLF HSEVB	127-188	211-271	483-533		
PVME1 IBV6	21-48	184-218					PVGLF HSEVB	127-188	211-271	483-533		
PVME1 IBV62	21-48	184-218					PVGLF HSEVB	127-188	211-271	483-533		
PVME1 IBVK		184-218					PVGLF HSEVB	127-188	211-271	483-533		

PVMP CAMVC	220-264	273-324		PVGLF SV41	96-188	454-508			
PVMP CAMVD	28-58	220-264	273-324	PVGLF SV6	103-171	241-276	451-487		
PVMP CAMVE		227-264	273-324	PVGLF TRTV	105-161	180-224	457-488		
PVMP CAMVN		220-264	273-324	PVGLG BEFV	508-812				
PVMP CAMVS		220-264	273-324	PVGLG BRVC	30-70	104-138			
PVMP CAMVW		220-264	273-324	PVGLG HRSV1	30-81				
PVMP CERV	28-53	100-127		PVGLG HRSV2	30-85				
PVMP SOCMV	4-31	78-118		PVGLG HRSV3	30-85				
PVMSA HPBHE	284-328			PVGLG HRSV4	30-107				
PVMT1 DHV11	38-65	237-284		PVGLG HRSV5	30-85				
PVMT8 MYXVL	183-180			PVGLG HRSV6	30-85				
PVMT8 MYXVL	485-482			PVGLG HRSV7	30-85				
				PVGLG HRSV8	30-81				
				PVGLG HRSVA	30-87				
				PVGLG HRSVL	25-85				
				PVGLG HRSV4	271-305				
				PVGLG SIGMA	344-381	464-488			
				PVGLG SYN	485-523				
				PVGLG VHSV0	383-387				
				PVGLG VSVIG	475-510				
				PVGLH EB	53-87	160-201	338-380	863-884	
				PVGLH HCMVA	103-137	270-311	683-741		
				PVGLH HCMVT	102-138	682-740			
				PVGLH HSV11	447-481				
				PVGLH HSV1E	447-481				
				PVGLH HSV6G	367-408				
				PVGLH HSVBC	384-418				
				PVGLH HSV4	334-378	414-455			
				PVGLH HSVB	327-372	407-448			
				PVGLH HSV8A	32-88	374-453	684-712		
				PVGLH MCMV8	440-474				
				PVGLH PRVKA	226-260				
				PVGLH PRVNG	226-280				
				PVGLH PRVRI	226-260				
				PVGLH VZVD	455-508				
				PVGLI HCMVA	47-111	323-358			
				PVGLM BUNGE	512-587	686-737	1228-1282		
				PVGLM BUNL7	643-677	818-850			
				PVGLM BUNSH	843-877				
				PVGLM BUNYW	340-374	504-563	805-839		
				PVGLM DUGBV	837-889	1239-1300			
				PVGLM HANTB	683-727				
				PVGLM HANTH	72-108				
				PVGLM HANTL	72-108				
				PVGLM HANTV	72-108				
				PVGLM PHV	73-111				
				PVGLM PTPV	148-251				

	PVGLM SEOUR	894-728							
	PVGLM SEOUS	893-730							
	PVGLN BEFV	377-414	613-669						
	PVGLP BEV	43-82	80-124	822-658	1128-1236				
	PVGLX HSVEB	177-282							
	PVGLX PRVRI	420-481							
	PVGLY JUNIN	301-349							
	PVGLY LASSG	317-380	388-422						
	PVGLY LASSJ	318-381	388-423						
	PVGLY LYCVA	333-387	395-432						
	PVGLY LYCVW	124-168	333-387	395-432					
	PVGLY MOPEI	316-368							
	PVGLY PIARV	334-375							
	PVGLY TACV	315-383							
	PVGLY TACV5	303-361	382-418						
	PVGLY TACV7	302-360	381-415						
	PVGLY TACVT	303-361	382-418						
	PVGNB CPMV	835-889							
	PVGNM BPMV	143-177	403-437						
	PVGNN CPMV	160-201							
	PVGNM CPBMV	192-228	758-792	874-915					
	PVGNN RCNV	837-871	812-848						
	PVGP8 EBV	84-149							
	PVM01 VACC	5-56							
	PVM1 REOVL	287-321							
	PVM21 REOVD	418-450	618-683						
	PVM22 REOVD	418-450	618-682						
	PVM2 REOVJ	418-450	618-682						
	PVM2 REOVL	418-450	618-682						
	PVM3 REOVD	135-190	337-371	523-558	618-680				
	PVMA2 BR6VA	42-80							
	PVMA2 HRBVA	42-80							
	PVMAT CDVO	193-234							
	PVMAT INCJJ	73-114	151-208						
	PVMAT NDVA	310-358							
	PVMAT NDVB	324-358							
	PVMAT PJ3B	99-133	204-252						
	PVMAT PJ3H4	99-133	204-252						
	PVMAT RABVA	68-103							
	PVMAT RABVC	68-103							
	PVMAT RABVE	68-103							
	PVMAT RABVN	68-103							
	PVMAT RABVP	68-103							
	PVMAT RABVS	68-103							
	PVMAT SYN	248-280							
	PVMAT VS6IQ	188-232							
	PVME1 CVBM	175-208							

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TABLE VI

Search Results Summary for PCTLZIP,
P1CTLZIP, and P2CTLZIP Motifs

[illegible]

PHEMA MUMPM	133-148	PHEMA IABAN	221-237			PHEMA CVHOC	391-408
PHEMA MUMPR	133-148	PHEMA IABUD	234-250			PHEMA IAAC	322-338
PHEMA MUMPS	133-148	PHEMA JACKA	234-250			PHEMA IABAN	306-323
PHEMA PI1HW	345-380	PHEMA JACKG	231-247			PHEMA IABUD	320-337
PHEMA P12H	65-80	PHEMA JACKV	230-246			PHEMA JACKA	320-337
PHEMA P12HT	65-80	PHEMA JADA1	234-250			PHEMA JACKG	316-333
PHEMA RINDK	398-383	PHEMA IADA3	237-253			PHEMA JACKP	302-319
PHEMA SVS	7-84	PHEMA IADCZ	234-250			PHEMA JACKQ	302-319
PHEMA SV6CM	7-84	PHEMA IADH1	221-237			PHEMA JACK8	319-336
PHEMA SV6CP	7-84	PHEMA IADH2	221-237			PHEMA JACKV	316-332
PHEMA SV6LN	7-84	PHEMA IADH3	221-237			PHEMA IADA1	320-337
PVENV DRV11	42-57	PHEMA IADH4	221-237			PHEMA IADA3	322-338
PVPF7 CAPVK	69-104	PHEMA IADH5	221-237			PHEMA IADCZ	320-337
PVVU5 VACC8	72-87	PHEMA IADH6	221-237			PHEMA IADH1	306-323
PVG01 BPP22	242-267	PHEMA IADH7	221-237			PHEMA IADH2	306-323
PVG01 HSEB	168-184	PHEMA IADM2	237-253			PHEMA IADH3	306-323
PVG01 HSV11	210-225	PHEMA IADNZ	234-250			PHEMA IADH4	306-323
PVG06 BPT4	184-188	PHEMA IAEN6	221-237			PHEMA IADH6	306-323
PVG07 BPT4	885-900	PHEMA IAEN7	237-253			PHEMA IADH7	306-323
PVG08 HSV11	134-149	PHEMA IAFPR	230-246			PHEMA IADM2	322-338
PVG10 BPPII2	183-188	PHEMA IAHAL	238-252			PHEMA IADNZ	320-337
PVG10 BPPZA	183-188	PHEMA IAHAR	235-251			PHEMA IADU3	322-338
PVG10 HSVSA	109-124	PHEMA IAHCB	230-246			PHEMA IAEN6	306-323
PVG16 BPP1	81-86	PHEMA IAHCB	230-246			PHEMA IAEN7	322-338
PVG16 BPT4	488-483	PHEMA IAHCD	230-246			PHEMA IAFPR	315-332
PVG25 BPT4	67-112	PHEMA IAHDE	230-246			PHEMA IAGRE	320-337
PVG28 HSV11	20-36	PHEMA IAHFO	230-252			PHEMA IAGU2	320-337
PVG30 BPH8	11-84	PHEMA IAHK6	238-252			PHEMA IAGUA	318-336
PVG38 BPOX2	22-37	PHEMA IAHK7	238-252			PHEMA IAHAL	321-338
PVG38 HSV6A	108-123	PHEMA IAHLE	230-246			PHEMA IAHCB	315-332
PVG37 BPT2	1253-1268	PHEMA IAHLO	230-246			PHEMA IAHCB	315-332
PVG37 HSV11	284-288	PHEMA IAHMI	238-252			PHEMA IAHCD	315-332
PVG65 HSV11	22-37	PHEMA IAHNM	238-252			PHEMA IAHDE	315-332
PVG68 HSV11	288-283	PHEMA IAHRO	238-252			PHEMA IAHFO	321-338
PVG68 HSV11	102-117	PHEMA IAHGA	238-252			PHEMA IAHK6	321-338
PVG68 HSV11	267-282	PHEMA IAH8P	230-246			PHEMA IAHK7	321-338
PVG65 HSV11	518-533	PHEMA IAHSW	230-246			PHEMA IAHLE	315-332
PVG8 BPH2	234-248	PHEMA IAHTE	238-252			PHEMA IAHLO	315-332
PVG8 BPPZA	234-249	PHEMA IAHTO	238-252			PHEMA IAHMI	321-338
PVG8 GPVR	67-72	PHEMA IAHUR	238-252			PHEMA IAHNM	321-338
PVG8 BPHX	234-249	PHEMA IAKIE	235-251			PHEMA IAHNN	315-332
PVL2 CVBF	264-278	PHEMA IALEN	235-251			PHEMA IAHNR	321-338
PVL2 CVBL9	264-278	PHEMA IAMAA	238-254			PHEMA IAHSA	315-332
PVL2 CVBL9	264-278	PHEMA IAMAB	238-254			PHEMA IAHSP	315-332
PVL2 CVBM	264-278	PHEMA IMAAO	237-253			PHEMA IAH8W	315-332
PVL2 CVBQ	264-278	PHEMA IAME1	237-253			PHEMA IAHTE	321-338
PVL2 CVBR	264-278	PHEMA IAME2	237-253				

PVGL2 CVPR9	442-457		PHEMA JAME8	221-237				PHEMA IAH70	321-338	
PVGL2 CVPPU	440-455	604-619	PHEMA IAMIN	85-101	231-247			PHEMA IAHUR	321-338	
PVGL2 CVPR8	218-233		PHEMA IANT6	237-253				PHEMA IAJAP	317-334	
PVGL2 CVPRM	218-233		PHEMA IAJQ7	221-237				PHEMA IAMAA	319-338	
PVGL2 BV8	1058-1071		PHEMA IARUD	234-250				PHEMA IAMAB	324-341	
PVGL2 BV8	1058-1070		PHEMA IASE2	234-250				PHEMA IAMAD	322-338	
PVGL2 BV2	1058-1071		PHEMA IASH2	234-250				PHEMA IAME1	322-339	
PVGL2 BVK	1058-1070		PHEMA IASTA	230-248				PHEMA IAME2	322-338	
PVGL2 BVW	1058-1070		PHEMA IATAI	235-251				PHEMA IAME8	306-323	
PVGLB H5VA	701-718		PHEMA IATKM	234-250				PHEMA IAMIN	316-333	
PVGLB PRVF	203-218		PHEMA IATKO	233-249				PHEMA IANT6	322-339	
PVGLC H5BC	478-490		PHEMA IATKR	230-248				PHEMA IAPIL	320-337	
PVGLC H5VE4	444-459		PHEMA IATKW	229-245				PHEMA IAPIL	306-323	
PVGLC H5VEB	427-442		PHEMA IAUDD	237-253				PHEMA IARUD	320-337	
PVGLC PRVF	446-461		PHEMA IAUS9	235-251				PHEMA IASE2	320-337	
PVGLD H5V11	79-84		PHEMA IAVI7	238-254				PHEMA IASH2	321-338	
PVGLD H5V2	79-84		PHEMA IAXIA	235-251				PHEMA IASTA	315-332	
PVGLF BR5VA	285-280		PHEMA IAZCO	237-253				PHEMA IATKM	320-337	
PVGLF BR5VC	285-280		PHEMA IAZH2	221-237				PHEMA IAUDD	322-339	380-387
PVGLF BR5VR	285-280		PHEMA IAZH3	221-237				PHEMA IAVI7	323-340	
PVGLF HR8V1	285-280		PHEMA IAZUK	237-253				PHEMA IAZCO	322-339	
PVGLF HR5VA	285-280		PHEMA INBAA	115-131	285-310			PHEMA IAZH2	306-323	
PVGLF HR5VL	285-280		PHEMA INB8E	123-139	303-318			PHEMA IAZH3	306-323	
PVGLF HR5VR	285-280		PHEMA INB8Q	118-132	283-308			PHEMA IAZUK	322-339	
PVGLF MUMPS	5-84		PHEMA INBEN	123-138	301-318			PHEMA MUMPM	101-118	
PVGLI VZVD	728-283		PHEMA INBFL	108-124	288-301			PHEMA MUMPR	101-118	
PVGLM HANTB	800-815		PHEMA INBGL	119-135	288-311			PHEMA MUMPS	101-118	
PVGLM PTPV	743-758		PHEMA INBHK	118-132	283-308			PHEMA NOVA	83-110	
PVGLM 5E0UR	801-818		PHEMA INBIB	108-124	288-303			PHEMA NDVB	83-110	
PVGLM 5E0UB	800-815		PHEMA INBID	120-136	289-314			PHEMA NDVD	83-110	
PVGLY LA98G	428-441		PHEMA INBLE	123-138	302-317			PHEMA NDVH	83-110	
PVGLY LASEJ	427-442		PHEMA INBMD	113-128	282-307			PHEMA NDVI	83-110	
PVGLY MOPEI	425-440		PHEMA INBME	118-132	286-311			PHEMA NDVM	83-110	
PVM3 REOVD	521-538		PHEMA INBNA	108-124	288-303			PHEMA NDVQ	83-110	
PVM9A HPGG9	380-385		PHEMA INBOR	123-139	301-318			PHEMA NDVTG	83-110	
PVM9A HPGV9	187-202		PHEMA INBBI	123-139	301-318			PHEMA NDVU	83-110	
PVM9A WHV1	378-383		PHEMA INBSJ	118-135	289-313			PHEMA PHODV	38-53	
PVM9A WHV69	383-388		PHEMA INBUS	118-132	284-309			PHEMA PIHW	488-503	
PVM9A WHV7	383-388		PHEMA INBVI	118-132	288-311			PHEMA PI8B	111-128	
PVM9A WHV8	383-388		PHEMA INBVK	123-139	303-318			PHEMA PI8H4	111-128	
PVM9A WHV8I	383-388		PHEMA INBY8	108-124	288-301			PHEMA PI8HA	111-128	
PVM9A WHVW8	234-249		PHEMA MUMPM	133-148				PHEMA PI8HT	111-128	
PVM2 IAAAN	25-40		PHEMA MUMPR	133-148				PHEMA PI8HU	111-128	
PVM2 IABAN	25-40		PHEMA MUMPS	133-148				PHEMA PI8HV	111-128	
PVM2 IAFOW	25-40		PHEMA PI1HW	345-360				PHEMA PI8HW	111-128	
PVM2 IAFPR	25-40		PHEMA PI2H	85-81				PHEMA PI8HX	111-128	
PVM2 IAFPW	25-40		PHEMA PI2HT	85-81				PHEMA PI8HA	60-67	

PVMT2 IALE1	25-40		PHEMA P13B	324-340					PHEMA 8V41	86-102	
PVMT2 IALE2	25-40		PHEMA P13H4	324-340					PHEMA 8V5	84-101	
PVMT2 IAMAN	25-40		PHEMA P13HA	324-340					PHEMA 8V6CM	84-101	
PVMT2 IAPUE	25-40		PHEMA P13HT	324-340					PHEMA 8V6CP	84-101	
PVMT2 IASIN	25-40		PHEMA P13HU	324-340					PHEMA 8V6LN	84-101	
PVMT2 IAUDO	25-40		PHEMA P13HV	324-340					PVFO5 VACCC	280-287	
PVMT2 IAWIL	25-40		PHEMA P13HW	324-340					PVFO5 VACCP	280-287	
PVMT9 MYXVL	228-241		PHEMA P13HX	324-340					PVFO5 VACCV	281-288	
			PHEMA RINDK	308-383					PVFO8 VACCC	178-193	
			PHEMA 8V6	7-94					PVFO8 VACCV	178-193	
			PHEMA 8V6CM	7-94					PVG27 HSVSA	208-228	
			PHEMA 8V6CP	7-94					PVG28 HSVI1	173-180	
			PHEMA 8V6LN	7-94					PVG39 HSVI1	648-665	
			PVENV DHV11	42-57					PVG43 HSVI1	109-128	621-638
			PVENV EAV	25-41					PVG67 HSVI1	171-188	
			PVFP2 FOWPV	89-104					PVG72 HSVI1	1252-1289	
			PVFP7 CAPVK	89-104					PVGFI IBVB	3073-3080	
			PVFUS VACC6	72-87					PVGL2 IBV8	1094-1111	
			PVG01 HSVEB	189-184					PVGLB HSVI1	738-753	
			PVG01 HSVI1	208-225	317-332				PVGLB HSVI1	675-682	
			PVG08 HSVI1	134-149					PVGLB HSVI1	738-753	
			PVG10 HSVSA	108-124					PVGLB HSVI1	738-753	
			PVG11 HSVI1	103-119					PVGLB HSVI1	738-753	
			PVG12 HSVI1	270-286					PVGLB LTV8	587-614	
			PVG1 SPVIR	76-92					PVGLB LTV8	607-624	
			PVG28 HSVI1	20-35					PVGLB LTVT	607-624	
			PVG36 BPOX2	22-37					PVGLC PRVIF	180-197	
			PVG36 HSVSA	108-123					PVGLF VZVD	489-488	
			PVG37 HSVI1	284-289					PVGLF 8V6	401-418	
			PVG41 HSVI1	244-260					PVGLH HCMVA	385-382	
			PVG46 HSVI1	1244-1260					PVGLH HCMVT	384-381	
			PVG55 HSVI1	22-37	143-158				PVGLH HSVI1	245-282	803-820
			PVG58 HSVI1	268-283					PVGLH HSVI1E	245-282	803-820
			PVG58 HSVI1	101-117					PVGLI HSVI1	43-60	
			PVG58 HSVI1	130-146	330-346				PVGLM BUNL7	81-98	
			PVG58 HSVSA	267-282					PVGLM BUNSH	81-98	
			PVG59 HSVI1	362-378	618-633				PVGLM PUUMH	712-728	
			PVG65 HSVI1	88-106					PVGLM PUUMS	712-728	
			PVG71 HSVSA	234-249					PVGLM RVFV	344-361	
			PVG8 BPPH2	234-249					PVGLM RVFVZ	344-381	
			PVG8 BPPZA	234-249					PVGLY LASSG	12-94	
			PVG8 SPVIR	57-72					PVGLY LASSJ	12-94	
			PVGFI IBVB	2210-2228					PVGLY LACVA	12-94	
			PVGL2 CVBF	123-139	174-180	264-279			PVGLY LYCVW	12-94	
			PVGL2 CVBL9	123-139	174-180	264-279			PVGLY MOPEI	12-94	
			PVGL2 CVBL1	123-139	174-180	264-279			PVM1 REOVD	280-297	
			PVGL2 CVBM	123-139	174-180	264-279			PVM1 REOVL	280-297	
			PVGL2 CVBQ	31-47	123-139	174-180	264-279				

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	PVGLF RINDK	282-288							
	PVGLF RINDL	282-288							
	PVGLF TRIV	176-191							
	PVGLI VZVD	278-283							
	PVGLM HANTB	366-371	900-916						
	PVGLM HANTH	489-516							
	PVGLM HANTL	499-516							
	PVGLM HANTY	489-516							
	PVGLM PTRV	743-768							
	PVGLM PUUMH	608-626							
	PVGLM PUUMS	608-626							
	PVGLM SEOUR	366-371	901-816						
	PVGLM SEOUS	366-371	900-916						
	PVGLM ULUK	828-842							
	PVGLP BEV	888-885							
	PVGLY LASSG	12-84	426-441						
	PVGLY LASSJ	12-84	427-442						
	PVGLY LYCVA	12-84							
	PVGLY LYCVW	12-84							
	PVGLY MOPEI	12-84	426-440						
	PVGLY PIARV	12-84							
	PVGMM CPMV	1021-1037							
	PVM3 REOVD	621-638							
	PVMAT MUMPS	181-207							
	PVMAT NDVA	136-161							
	PVMAT NDVB	136-161							
	PVMAT PIZHT	189-208							
	PVMAT SV41	189-206							
	PVMAT SV6	89-114	132-148						
	PVMP CAMVC	118-134							
	PVMP CAMVD	118-134							
	PVMP CAMVE	118-134							
	PVMP CAMVN	118-134							
	PVMP CAMV6	118-134							
	PVMP CAMVW	118-134							
	PVMP FMAVD	116-131							
	PVMSA HPBG9	380-396							
	PVMSA HPBV8	187-202							
	PVMSA WHV1	378-383							
	PVMSA WHV59	383-388							
	PVMSA WHV7	383-388							
	PVMSA WHV8	383-388							
	PVMSA WHV81	383-388							
	PVMSA WHVW6	234-248							
	PVMT2 IAANN	26-40							
	PVMT2 IABAN	26-40							
	PVMT2 LAEOW	26-40							

[illegible]

TABLE VII

Search Results Summary for P3CTLZIP, P4CTLZIP,
P5CTLZIP, and P6CTLZIP Motifs

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PVMO1 VACCV	83-101	128-144	PVGL2 CVM4	889-1018	PVENV THQCV	358-378	PHEMA P12H	13-34	
PVM1 REOVD	227-245		PVGL2 CVM45	947-868	PVG01 VACCV	288-318	PHEMA P12H	13-34	
PVM1 REOVL	227-245		PVGL2 CVM4H	868-877	PVG01 VACCV	237-267	PHEMA SV6	7-28	378-400
PVMAT HRSVA	44-82		PVGL2 CVPF8	64-83	PVG01 VARV	288-318	PHEMA SV6CM	7-28	378-400
PVMAT NDVA	180-208		PVGL2 CVPPU	84-83	PVG08 VACCV	31-51	PHEMA SV6CP	7-28	378-400
PVMAT NDVB	180-208		PVGL2 CVPR8	814-833	PVG08 VARV	31-51	PHEMA SV6LN	7-28	378-400
PVMP CAMVC	183-201		PVGL2 CVPRM	814-833	PVG08 BPFF1	26-45	PVG01 HSEVB	188-180	
PVMP CAMVD	183-201		PVGL2 FIPV	1041-1080	PVG12 HSN11	161-171	PVG01 HSN11	588-610	
PVMP CAMVE	183-201		PVGL2 IBV8	588-607	PVG22 HSN11	300-320	PVG23 HSN11	314-335	
PVMP CAMVN	183-201		PVGL2 IBV8	587-806	PVG38 HSN11	648-668	PVG37 BPOX2	65-88	
PVMP CAMV5	183-201		PVGL2 IBVD2	588-607	PVG51 HSN11	28-49	PVG43 HSN11	167-178	
PVMP CAMVW	183-201		PVGL2 IBVK	587-806	PVG63 HSN11	338-358	PVG55 HSN11	288-309	
PVMP FMVD	180-188		PVGL2 IBVM	587-806	PVG65 HSN11	117-137	PVG55 HSN11	85-108	
			PVGLB HCMVA	708-725	PVG74 HSN11	124-144	PVG56 HSN11	1155-1178	
			PVGLB HCMVT	707-726	PVGL2 IBV8	328-348	PVG58 HSN11	288-287	
			PVGLB HSNV8U	117-138	PVGL2 IBV8	327-347	PVG80 HSN11	30-51	
			PVGLB ILTV8	268-275	PVGL2 IBVD2	328-348	PVG83 HSN11	238-259	
			PVGLB ILTVS	268-285	PVGL2 IBVD3	328-348	PVG81 IBV8	1868-1877	
			PVGLB ILTVT	268-285	PVGL2 IBVK	327-347	PVG83 HCMVA	167-178	
			PVGLC HSN11	3-84	PVGL2 IBVM	327-347	PVGL2 CVBF	1259-1280	
			PVGLC HSN1K	3-84	PVGL2 IBV2	310-330	PVGL2 CVBL8	1259-1280	
			PVGLC HSNVC	476-484	PVGLB EBV	732-752	PVGL2 CVBL1	1259-1280	
			PVGLG CHAV	438-455	PVGLB HCMVA	750-770	PVGL2 CVBM	1259-1280	
			PVGLG RABVH	372-381	PVGLB HCMVT	751-771	PVGL2 CVBQ	1259-1280	
			PVGLI HSEVB	44-63	PVGLB HSN23	78-89	PVGL2 CVBV	1259-1280	
			PVGLI VZVD	278-287	PVGLB HSN2H	78-89	PVGL2 CVM4	1317-1338	
			PVGLM BUNGE	117-136	PVGLB HSN2S	65-85	PVGL2 CVM45	1265-1288	
			PVGLM PHV	152-171	PVGLB HSN2U	72-82	PVGL2 CVM4H	1178-1187	
			PVGLM PTPV	887-1016	PVGLB HSNB2	278-289	PVGLB HSN11	83-104	
			PVGLM PUUMH	155-174	PVGLB HSN6A	63-83	PVGLB HSN1F	82-103	
			PVGLM PUUMS	156-174	PVGLB MCMV9	738-758	PVGLB HSN1K	82-103	
			PVGLM RVFV	830-849	PVGLG P13H4	283-303	PVGLB HSN1P	83-104	
			PVGLM RVFVZ	830-849	PVGLG RABVE	454-474	PVGLB MCMV9	135-158	
			PVGLM UUK	655-674	PVGLG RABVH	454-474	PVGLC PRVIF	448-487	
			PVGLY LYCVW	89-108	PVGLG RABVP	454-474	PVGLF CDVO	338-357	
			PVGLB CPMV	1185-1184	PVGLG RABVT	454-474	PVGLF MEASE	224-245	
			PVM3 REOVD	521-540	PVGLH RABV8	454-474	PVGLF MEASI	227-248	
			PVME1 CVH22	136-155	PVGLH MCMV8	670-680	PVGLF MEASY	224-245	
			PVME1 CVBM	171-180	PVGLM BUNL7	1326-1345	PVGLF MUMPM	448-487	
			PVME1 CVPF8	174-183	PVGLM BUNH	1325-1345	PVGLF MUMPR	448-487	
			PVME1 CVPPU	174-183	PVGLM BUNYW	898-1016	PVGLF MUMPS	448-487	
			PVME1 CVPRM	174-183	PVGLM HANTB	898-1019	PVGLF PHODV	305-328	
			PVME1 CVTKE	171-180	PVGLM HANTH	1000-1020	PVGLF P11HC	458-477	
					PVGLM HANTL	1001-1021	PVGLF P12H	450-471	
					PVGLM HANTV	1001-1021	PVGLF P12HG	450-471	
					PVGLM RVFVZ	1158-1178	PVGLF P12HT	450-471	
					PVGLM SEOUR	1000-1020	PVGLF P13B	408-428	453-474

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TABLE VIII

Search Results Summary for P7CTLZIP,
P8CTLZIP, and P9CTLZIP Motifs

P7CTLZIP		P8CTLZIP		P8CTLZIP	
LIBRARY FILE	LIBRARY FILE	LIBRARY FILE	LIBRARY FILE	LIBRARY FILE	LIBRARY FILE
202-224	PENV1 FRSFV	380-403	PENV1 FRSFV	303-327	
488-520	PENV1 FRSFV	380-403	PENV BLVAF	303-327	
493-516	PENV BIV08	178-201	PENV BLVAU	303-327	
494-518	PENV BIV27	207-230	PENV BLVAV	303-327	
603-626	PENV FOAMV	884-887	PENV BLVB2	303-327	
495-517	PENV HV123	176-188	PENV BLVB5	303-327	
488-520	PENV HV2BE	3-28	PENV BLVJ	303-327	
488-520	PENV HV2CA	760-773	PENV FVPE	781-805	
510-532	PENV HV2D1	3-28	PENV FV5D	778-803	
490-512	PENV HV2G1	772-785	PENV FV2T	780-804	
504-526	PENV HV2N2	777-800	PHEMA CVBLY	391-415	
500-522	PENV JSRV	541-564	PHEMA CVBM	391-415	
488-518	PENV SFV1	884-887	PHEMA CVHOC	391-415	
498-510	PENV SFV3L	881-804	PHEMA INCCA	442-466	
488-520	PENV SINM1	803-828	PHEMA INCEN	430-454	
488-511	PENV SINMK	802-825	PHEMA INCOL	430-454	
123-145	PENV SINML	801-824	PHEMA INCY	428-453	
497-519	PENV SIN64	808-828	PHEMA INCJH	443-487	
505-527	PENV SIN3P	810-833	PHEMA INCKY	428-453	
498-520	PHEMA CDVO	200-223	PHEMA INCM1	428-453	
378-388	PHEMA PI2H	65-88	PHEMA INCMNA	428-453	
213-235	PHEMA PI2HT	65-88	PHEMA INCP1	430-454	
213-235	PVF11 VACCC	181-184	PHEMA NCP2	430-454	
37-58	PVF16 VACCC	25-48	PHEMA NCP3	430-454	
21-43	PVF15 VACCP	3-28	PHEMA NCTA	430-454	
37-58	PVG11L AMEPV	313-336	PHEMA NCYA	430-454	
21-43	PVG28 HSV11	491-514	PHEMA MUMPM	101-125	
21-43	PVG43 HSV11	322-346	PHEMA MUMPR	101-125	
21-43	PVG52 HSV11	228-252	PHEMA MUMPS	101-125	
21-43	PHEMA IADH5	722-745	PHEMA PITHW	28-53	
21-43	PVGL2 CVBFE	10-33	PVENV BEV	82-88	
21-43	PVGL2 CVBL8	861-874	PVF05 VACCC	280-304	
21-43	PVGL2 CVBLY	10-33	PVF05 VACCP	280-304	
28-50	PVGL2 CVMA5	1287-1280	PVF05 VACCV	281-305	
37-58	PVGL2 CVMA5	1216-1238	PVF08 VACCC	178-200	
21-43	PVGL2 CVMJH	1128-1148	PVF08 VACCV	178-200	
37-58	PVGL2 CVPF5	1274-1287	PVG01 VZVD	58-82	
37-58	PVGL2 CVPPU	1272-1286	PVG10 HBSVA	355-378	
37-58	PVGL2 CVPR8	1050-1073	PVG12 HSV5A	85-82	
37-58	PVGL2 CVPRM	1050-1073	PVG18 HSV11	88-112	
21-43	PVGL2 FIPV	1277-1300	PVG28 HSV11	173-187	
37-58	PVGL2 IBV6	185-218	PVG43 HSV11	108-133	
21-43	PVGL2 IBVB	185-218	PVG67 HSV11	108-132	1006-1029
37-58	PVGL2 IBVD2	188-218	PVG72 HSV11	720-744	
37-58	PVGL2 IBVD3	188-218	PVG71 IBVB	3801-3825	

PHEMA IAV17	38-60		PVGL2 IBVK	185-218	PVGLB HSMVD	588-613			
PHEMA IAX31	37-58		PVGL2 IBVM	185-218	PVGLB ILTV6	587-621			
PHEMA IAZC0	37-58		PVGL2 IBVU1	178-201	PVGLB ILTV8	607-631			
PHEMA IAZH2	21-43		PVGL2 IBVU2	178-201	PVGLB ILTVT	607-631			
PHEMA IAZH3	21-43		PVGL2 IBVU3	178-201	PVGLB HSMV11	413-437			
PHEMA IAZUK	37-58		PVGLB HCMVA	535-558	PVGLB VZVD	488-483			
PHEMA PHODV	38-68		PVGLB HCMVT	538-558	PVGLF SV6	401-425			
PHEMA PI2H	85-87		PVGLB H8V6A	483-508	PVGLH HCMVA	574-588			
PHEMA PI2HT	85-87		PVGLB MCMV8	608-608	PVGLH HCMVT	573-587			
PVF77 CAPVK	88-111		PVGLC H8V11	487-480	PVGLH HSMV11	443-487	803-827		
PVFUS VACC6	72-84		PVGLC H8V2	435-458	PVGLH HSMV1E	443-487	803-827		
PVG01 HSMV11	317-338		PVGLC H8V23	438-458	PVGLM BUNL7	31-55			
PVG03 VACC	50-72		PVGLM BUNL7	1387-1410	PVGLM BUNSH	31-55			
PVG04 VACC	11-33		PVGLM BUNSH	1387-1410	PVGLM RVFV	344-388			
PVG04 VARV	11-33		PVGLM UUK	688-688	PVGLM RVFVZ	344-388			
PVG18 HSMV11	88-110		PVGLY JUNIN	12-35	PVGLM UUK	591-585			
PVG28 HSMV11	173-185		PVGLY LASSG	12-35	PVGNM CPMV	311-335			
PVG28 HSMV1	20-42		PVGLY LASSJ	12-35	PVGP2 EBV	657-681			
PVG48 HSMV1	134-158		PVGLY LYCVA	12-35	PVGP3 EBV	854-878			
PVG48 HSMVA	71-83		PVGLY LYCVW	12-35	PVM1 REOVD	280-304			
PVG68 HSMVA	288-288		PVGLY MOPEI	12-35	PVM1 REOVL	280-304			
PVG68 HSMV1	287-288		PVGLY TACV5	12-35	PVM21 REOVD	188-192			
PVG5 SPV4	42-64		PVGLY TACV6	12-35	PVM22 REOVD	188-192			
PVG80 HSMV11	53-75		PVGLY TACV7	12-35	PVM2 REOVL	188-192			
PVG85 HSMV11	1347-1388		PVGLY TACVT	12-35	PVM2 REOVL	188-192			
PVG8 SPV1R	60-82		PVGNM CPMV	741-784	PVMAT MEAS1	87-111			
PVGL2 IBV8	1058-1078		PVM1 REOVD	324-347	PVMAT SSPVB	314-338			
PVGL2 IBV8	1056-1077		PVM1 REOVL	464-477	PVME1 CVBM	137-181			
PVGL2 IBVD2	1058-1078		PVMAT MUMPS	227-250	PVME1 CVHOC	137-181			
PVGL2 IBVK	1055-1077		PVMSA HPBDB	269-282	PVME1 CVTKE	137-181			
PVGL2 IBVM	1055-1077		PVMSA HPBDC	269-281	PVME1 IBV8	74-98			
PVGLB H8VB2	117-138		PVMSA HPBDU	231-254	PVME1 IBV8	74-98			
PVGLB H8VB2	745-767		PVMSA HPBDW	269-282	PVME1 IBV2	74-98			
PVGLC H8VMB	389-421		PVMSA HPBHE	238-259	PVME1 IBVK	74-98			
PVGLC H8VMD	388-420				PVMSA HPBGS	271-285			
PVGLC H8VMM	389-421				PVMSA WHV1	269-283			
PVGLF BR5VA	285-287	482-504			PVMSA WHV59	274-288			
PVGLF BR5VC	484-508				PVMSA WHV7	274-288			
PVGLF BR5VR	484-508				PVMSA WHV8	274-288			
PVGLF HR8V1	484-508				PVMSA WHV8I	274-288			
PVGLF HR8VA	484-508				PVMSA WHVW8	125-149			
PVGLF HR8VL	484-508								
PVGLF HR5VR	484-508								
PVGLF TRTV	452-474								
PVGLG IHNV	77-89								
PVGLG VH8V0	408-428								

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TABLE IX

Search Results Summary for P12CTLZIP Motif

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TABLE X.

Search Results Summary for P23CTLZIP Motif

[illegible]

PENV HV1W1	730-783				
PENV HV1W2	721-754				
PENV HV1Z2	264-286	727-760			
PENV HV1Z3	250-281				
PENV HV1Z6	255-280	729-762			
PENV HV1Z8	265-288				
PENV HV2BE	781-811				
PENV HV2D1	772-802				
PENV HV2G1	772-802				
PENV HV2N2	777-814				
PENV HV2SB	743-776				
PENV JSRV	288-332	484-516			
PENV MMTVB	435-472				
PENV MMTVG	435-472				
PENV RSPV	533-570				
PENV 6FV1	44-78	482-530			
PENV 8FV3L	48-82	550-588			
PENV BIVC2	745-776				
PENV BIVGB	247-277	353-388			
PENV BIVM1	788-800				
PENV BIVMK	785-788				
PENV BIVML	511-545	784-788			
PENV BIV84	458-488				
PENV BIV8P	482-480	810-840			
PHEMA CDVO	200-234				
PHEMA IABUD	23-55				
PHEMA IACKA	23-55				
PHEMA IACKV	517-547				
PHEMA IADA1	23-55				
PHEMA IADC2	23-55				
PHEMA IADH5	283-323				
PHEMA IADNZ	23-55				
PHEMA IAFPR	15-51				
PHEMA IAGRE	23-55				
PHEMA IAMAA	22-54				
PHEMA IAMAB	27-58				
PHEMA IARUD	23-55				
PHEMA IASE2	23-55				
PHEMA IASTA	517-547				
PHEMA MUMPM	19-52	101-132			
PHEMA MUMPR	19-52	101-132			
PHEMA MUMPB	19-52	101-132			
PHEMA NDVA	60-88				
PHEMA NDVB	60-88				
PHEMA NDVD	60-88				
PHEMA NDVH	60-88				
PHEMA NDVI	60-88				

PHEMA_NDVM	60-88								
PHEMA_NDVO	60-88								
PHEMA_NDVTG	60-88								
PHEMA_NDVU	60-88								
PHEMA_PI1HW	28-60					186-233			
PHEMA_PI2H	13-48					334-389			
PHEMA_PI2HT	13-48					334-389			
PHEMA_PI3B	184-231								
PHEMA_PI3H4	184-231								
PHEMA_PI3HA	184-231								
PHEMA_PI3HT	184-231								
PHEMA_PI3HU	184-231								
PHEMA_PI3HV	184-231								
PHEMA_PI3HW	184-231								
PHEMA_PI3HX	184-231								
PHEMA_PI4HA	245-280								
PHEMA_RACVI	255-283					338-376			
PHEMA_RINDL	282-313								
PHEMA_SEND6	16-54					186-233			
PHEMA_SENDF	16-54					186-233			
PHEMA_SENDH	16-54					186-233			
PHEMA_SENDJ	16-54					186-233			
PHEMA_SEND2	23-54					186-233			
PHEMA_SV41	55-84					330-385			
PHEMA_SV6	7-35								
PHEMA_SV6CM	7-41								
PHEMA_SV6CP	7-41								
PHEMA_SV6LN	7-35								
PHEMA_VACCC	258-284								
PHEMA_VACCI	258-284								
PHEMA_VACGT	258-284								
PHEMA_VACCV	258-284								
PVENV_BEV	16-51					87-117			
PVENV_DHV11	287-335								
PVENV_MCV1	203-238								
PVENV_MCV2	203-238								
PVENV_VACCC	208-241								
PVENV_VACCI	208-241								
PVENV_VACCP	208-241								
PVENV_VACCV	208-241								
PVFO3_VACCC	2-40					61-83			
PVFO3_VACCV	2-40					61-83			
PVFP1_FOWPV	287-330								
PVFP4_FOWPV	237-267								
PVFP7_CAPVK	88-118								
PVRUB_VACCC	28-61								
PVRUS_VACCV	28-61								

PVG01 HSV11	317-346				
PVG02 HSEB	163-198				
PVG02 VACCV	92-120				
PVG02 VARV	92-120				
PVG03 HSV11	106-136				
PVG06 HSV11	54-83				
PVG06 VACCV	88-136				
PVG06 VARV	88-136				
PVG07 VACCV	113-145				
PVG07 VARV	113-145				
PVG09 VACCV	303-338				
PVG09 VACCV	266-301				
PVG09 VARV	303-338				
PVG11 HSV11	150-183				
PVG12 HSV11	206-243				
PVG12 HSV5A	68-106				
PVG1 SPV1R	254-282	303-337	414-462		
PVG22 HSV11	300-337	647-678			
PVG23 HSV11	70-108				
PVG26 HSV11	84-125				
PVG27 HSV5A	38-74				
PVG28 HSV11	491-521				
PVG28 HSV5A	7-40				
PVG2R AMEPV	180-217				
PVG2 SPV4	209-244				
PVG35 HSV11	15-48	180-226			
PVG36 HSV5A	151-185				
PVG38 HSV11	543-577	648-682			
PVG40 HSV5A	167-216				
PVG41 HSV11	11-46	202-233			
PVG42 HSV11	91-126				
PVG43 HSV11	108-140	167-185			
PVG46 HSV11	888-926				
PVG48 HSV5A	328-357				
PVG50 HSV5A	113-141				
PVG51 HSV11	28-64	84-120			
PVG52 HSV11	88-134				
PVG55 HSV11	100-129				
PVG58 HSV11	831-867	1081-1126			
PVG58 HSV11	342-376	480-508			
PVG58 HSV5A	25-60	195-233			
PVG59 HSV11	92-118				
PVG61 HSV11	76-109				
PVG64 HSV11	55-89	363-401	420-462		
PVG65 HSV11	801-838	1290-1328			
PVG67 HSV11	150-188	1160-1185			
PVG8 SPV1R	60-80				

PVG71 H8V9A	128-168					
PVG72 H8V11	445-478	720-751	1158-1189	1252-1285		
PVG75 H8V11	283-291	387-422				
PVG78 H8V11	187-221					
PVG7 8PV1R	18-48					
PVGFT1 IBVB	1718-1747	1856-1891	2108-2148	3601-3633		
PVGH3 HCMVA	80-116	157-186				
PVGL2 CVB8F	1258-1284					
PVGL2 CVB8G	651-681	1258-1284				
PVGL2 CVBLY		1258-1284				
PVGL2 CVB8M		1258-1284				
PVGL2 CVB8Q		1258-1284				
PVGL2 CVB8V		1258-1284				
PVGL2 CVH22	1053-1088					
PVGL2 CVH4	1257-1304					
PVGL2 CVMA5	1215-1262					
PVGL2 CVMJH	1128-1163					
PVGL2 CVPF6	632-665	736-784	1328-1383			
PVGL2 CVPPU	630-663	734-782	1328-1381			
PVGL2 CVPR8	512-540	1104-1139				
PVGL2 CVPRM	408-441	1104-1139				
PVGL2 FIPV	635-688	738-787	1331-1388			
PVGL2 IBVB	153-188					
PVGLB HCMVA	116-147	706-743				
PVGLB HCMVT	118-147	707-744				
PVGLB H8VB8U	72-110					
PVGLB H8VB1	254-288					
PVGLB H8VB2	284-288	745-774				
PVGLB H8VBC	253-287					
PVGLB ILTV8	442-472					
PVGLB ILTV8	452-482					
PVGLB ILTVT	452-482					
PVGLB MCMV8	135-163	738-778				
PVGLC H8V11	487-500					
PVGLC H8V1K	487-500					
PVGLC H8V2	435-465					
PVGLC H8V23	438-468					
PVGLC H8VBC	476-507					
PVGLC VZVD	351-388	513-548				
PVGLC VZV8	351-388	513-548				
PVGLD H8VEA	340-370					
PVGLD H8VEB	41-70	380-420				
PVGLD H8VEK	41-70	380-420				
PVGLD H8VE4	85-125					
PVGLE H8VEB	63-100	380-420				
PVGLE H8VEL	63-100	382-422				
PVGLE PRVRI	332-369					

PVGLF BR8VA	285-301	482-511		
PVGLF BR8VC	484-513			
PVGLF BR8VR	484-513			
PVGLF CDVO	562-588			
PVGLF HRSVI	484-513			
PVGLF HRSVA	484-513			
PVGLF HRSVL	484-513			
PVGLF HRSVR	484-513			
PVGLF MEASE	224-258	451-484		
PVGLF MEAGI	227-258	454-487		
PVGLF MEASY	224-258	451-484		
PVGLF MUMPM	446-474			
PVGLF MUMPR	446-474			
PVGLF MUMPS	5-38	446-474		
PVGLF NDVI	132-185			
PVGLF PHODV	531-585			
PVGLF P11HC	456-484			
PVGLF P13B	453-481			
PVGLF P13H4	453-481			
PVGLF RINDK	220-252	447-480		
PVGLF RINDL	220-252	447-480		
PVGLF SEND5	480-488			
PVGLF SENDF	480-488			
PVGLF SENDH	480-488			
PVGLF SENDJ	480-488			
PVGLF SENDZ	480-488			
PVGLF SV5	446-474			
PVGLF TRTV	452-481			
PVGLQ H8VEB	327-384			
PVGLQ 6YNV	524-553			
PVGLQ V8VIQ	450-488			
PVGLQ V8VJO	457-482			
PVGLQ V8VO	450-488			
PVGLQ V8V6J	450-488			
PVGLH HCMVA	681-719			
PVGLH HCMVT	680-718			
PVGLH H8V6G	640-677			
PVGLH H8VE4	814-850			
PVGLH H8VEB	807-843			
PVGLI HCMVA	158-184			
PVGLM BUNGE	187-227	438-488	982-1020	1049-1084
PVGLM BUNL7	180-220			
PVGLM BUNSH	180-220	344-381		
PVGLM BUNYW	183-228	434-472	823-854	
PVGLM DUGBV	244-273	837-872	880-915	935-965
PVGLM HANTB	810-841	1081-1119		
PVGLM HANTH	188-222	812-843	1082-1120	

PVGLM HANTL	188-222	612-643	1083-1121	
PVGLM HANTV	188-222	612-643	1083-1121	
PVGLM PHV	816-849	1088-1121		
PVGLM PTPV	848-882	1275-1308		
PVGLM PUUMH	820-853	1092-1125		
PVGLM PUUMS	820-853	1092-1125		
PVGLM RVFV	820-853	830-863		
PVGLM RVFVZ	820-853	830-863	1160-1185	
PVGLM SEOUR	805-841	1082-1120		
PVGLM SEOUS	810-841	1081-1118		
PVGLM UUK	431-468	966-995		
PVGLP BEV	1481-1628			
PVGLY JUNIN	12-45			
PVGLY LASSG	237-285			
PVGLY LASSJ	236-268			
PVGLY PIARV	12-50			
PVGLY TACV	12-50			
PVGLY TACV5	12-50	89-124		
PVGLY TACV7	12-50	88-124		
PVGLY TACVT	12-50	88-124		
PVGNB CPMV	1527-1555			
PVGNM BPMV	137-167	280-327	837-868	
PVGNM CPMV	208-242	741-771		
PVGNM CFSMV	50-88	478-515		
PVGNM RCMV	766-789			
PVGP2 EBV	78-111			
PVGP3 EBV	78-111			
PVM1 REOVD	280-318	324-361		
PVM1 REOVL	280-318			
PVM21 REOVD	188-199			
PVM22 REOVD	188-199			
PVM2 REOVJ	188-199			
PVM2 REOVL	188-199			
PVM3 REOVD	333-384			
PVMAT SV5	305-342			
PVMAT TRTV	122-150			
PVME1 CVBM	64-102			
PVME1 CVHOC	84-102			
PVME1 CVMAS	65-103			
PVME1 CVMJH	65-103			
PVME1 CVTKE	64-102			
PVMEM EBV	178-213			
PVMP CERV	83-126			
PVMP SOCMV	68-88	273-303		
PVMSA HPBDB	201-238	288-302		
PVMSA HPBDC	184-227	288-301		
PVMSA HPBDU	157-190	231-264		

[illegible]

5.3. SYNTHESIS OF PEPTIDES

The peptides of the invention may be synthesized or prepared by techniques well known in the art. See, for example, Creighton, 1983, Proteins: Structures and Molecular Principles, W.H. Freeman and Co., NY, 5 which is incorporated herein by reference in its entirety. Short peptides, for example, can be synthesized on a solid support or in solution. Longer peptides may be made using recombinant DNA techniques. Here, the nucleotide sequences encoding the peptides 10 of the invention may be synthesized, and/or cloned, and expressed according to techniques well known to those of ordinary skill in the art. See, for example, Sambrook, et al., 1989, Molecular Cloning, A Laboratory Manual, Vols. 1-3, Cold Spring Harbor 15 Press, NY.

The peptides of the invention may alternatively be synthesized such that one or more of the bonds which link the amino acid residues of the peptides are non-peptide bonds. These alternative non-peptide 20 bonds may be formed by utilizing reactions well known to those in the art, and may include, but are not limited to imino, ester, hydrazide, semicarbazide, and azo bonds, to name but a few. In yet another embodiment of the invention, peptides comprising the 25 sequences described above may be synthesized with additional chemical groups present at their amino and/or carboxy termini, such that, for example, the stability, bioavailability, and/or inhibitory activity of the peptides is enhanced. For example, hydrophobic 30 groups such as carbobenzoxyl, dansyl, or t-butyloxycarbonyl groups, may be added to the peptides' amino termini. Likewise, an acetyl group or a 9-fluorenylmethoxy-carbonyl group may be placed at the peptides' amino termini. (See "X" in Tables I to IV, 35 above.) Additionally, the hydrophobic group, t-

butyloxycarbonyl, or an amido group may be added to the peptides' carboxy termini. (See "Z" in Tables I to IV, above.) Further, the peptides of the invention may be synthesized such that their steric configuration is altered. For example, the D-isomer of one or more of the amino acid residues of the peptide may be used, rather than the usual L-isomer. Still further, at least one of the amino acid residues of the peptides of the invention may be substituted by one of the well known non-naturally occurring amino acid residues. Alterations such as these may serve to increase the stability, bioavailability and/or inhibitory action of the peptides of the invention.

Any of the peptides described above may, additionally, have a non-peptide macromolecular carrier group covalently attached to their amino and/or carboxy termini. Such macromolecular carrier groups may include, for example, lipid-fatty acid conjugates, polyethylene glycol, or carbohydrates. "X", in Tables I to IV, above, may therefore additionally represent any of the above macromolecular carrier groups covalently attached to the amino terminus of a peptide. Likewise, "Z", in Tables I to IV, may additionally represent any of the macromolecular carrier groups described above.

5.4. ASSAYS FOR ANTIVIRAL ACTIVITY

The antiviral activity exhibited by the peptides of the invention may be measured, for example, by easily performed in vitro assays, such as those described below, which can test the peptides' ability to inhibit syncytia formation, or their ability to inhibit infection by cell-free virus. Using these assays, such parameters as the relative antiviral activity of the peptides, exhibit against a given strain of virus and/or the strain specific inhibitory

activity of the peptide can be determined. A cell fusion assay may be utilized to test the peptides' ability to inhibit HIV-induced syncytia formation in vitro. Such an assay may comprise culturing uninfected CD-4⁺ cells (such as Molt or CEM cells, for example) in the presence of chronically HIV-infected cells and a peptide to be assayed. For each peptide, a range of peptide concentrations may be tested. This range should include a control culture wherein no peptide has been added. Standard conditions for culturing, well known to those of ordinary skill in the art, are used. After incubation for an appropriate period (24 hours at 37°C, for example) the culture is examined microscopically for the presence of multinucleated giant cells, which are indicative of cell fusion and syncytia formation.

A reverse transcriptase (RT) assay may be utilized to test the peptides' ability to inhibit infection of CD-4⁺ cells by cell-free HIV. Such an assay may comprise culturing an appropriate concentration (i.e., TCID₅₀) of virus and CD-4⁺ cells in the presence of the peptide to be tested. Culture conditions well known to those in the art are used. As above, a range of peptide concentrations may be used, in addition to a control culture wherein no peptide has been added. After incubation for an appropriate period (e.g., 7 days) of culturing, a cell-free supernatant is prepared, using standard procedures, and tested for the presence of RT activity as a measure of successful infection. The RT activity may be tested using standard techniques such as those described by, for example, Goff et al. (Goff, S. et al., 1981, J. Virol. 38:239-248) and/or Willey et al. (Willey, R. et al., 1988, J. Virol. 62:139-147). These references are incorporated herein by reference in their entirety.

Standard methods which are well-known to those of skill in the art may be utilized for assaying non-retroviral activity. See, for example, Pringle et al. (Pringle, C.R. et al., 1985, J. Medical Virology 17:377-386) for a discussion of respiratory syncytial virus and parainfluenza virus activity assay techniques. Further, see, for example, "Zinsser Microbiology", 1988, Joklik, W.K. et al., eds., Appleton & Lange, Norwalk, CT, 19th ed., for a general review of such techniques. These references are incorporated by reference herein in its entirety.

5.5. USES OF THE PEPTIDES OF THE INVENTION

The DP-178 (SEQ ID:1) peptides of the invention, and DP-178 fragments, analogs, and homologs, exhibit potent antiviral activity. The DP-107-like and DP-178-like peptides of the invention preferably exhibit antiviral activity. As such, the peptides may be used as inhibitors of human and non-human viral and retroviral, especially HIV, transmission to uninfected cells.

The human retroviruses whose transmission may be inhibited by the peptides of the invention include, but are not limited to all strains of HIV-1 and HIV-2 and the human T-lymphocyte viruses (HTLV-I and II). The non-human retroviruses whose transmission may be inhibited by the peptides of the invention include, but are not limited to bovine leukosis virus, feline sarcoma and leukemia viruses, simian immunodeficiency, sarcoma and leukemia viruses, and sheep progress pneumonia viruses.

Non retroviral viruses whose transmission may be inhibited by the peptides of the invention include, but are not limited to human respiratory syncytial virus, canine distemper virus, newcastle disease virus, human parainfluenza virus, and influenza

viruses. Further, any virus or retrovirus containing peptides listed in Tables V through X above, may be inhibited by the peptides of the invention.

As discussed more fully, below, in Section 5.5.1 and in the Example presented, below, in Section 8, DP-107 and DP-178, and DP-107-like and DP-178-like peptides form non-covalent protein-protein interactions which are required for normal activity of the virus. Thus, the peptides of the invention may also be utilized as components in assays for the identification of compounds that interfere with such protein-protein interactions and may, therefore, act as antiviral agents. These assays are discussed, below, in Section 5.5.1.

5.5.1. ANTIVIRAL COMPOUND SCREENING SCREENING ASSAYS FOR COMPOUNDS THAT INTERACT WITH THE PKD1 GENE PRODUCT

As demonstrated in the Example presented in Section 8, below, DP-107 and DP-178 portions of the TM protein gp41 form non-covalent protein-protein interactions. As also demonstrated, the maintenance of such interactions is necessary for normal viral infectivity. Thus, compounds which bind DP-107, bind DP-178, and/or act to disrupt normal DP-107/DP-178 protein-protein interactions may act as potent antiviral agents. Described below are assays for the identification of such compounds. Note that, while, for ease and clarity of discussion, DP-107 and DP-178 peptides will be used as components of the assays described, but it is to be understood that any of the DP-107-like or DP-178-like peptides described, above, in Sections 5.1 and 5.2 may also be utilized as part of these screens for antiviral compounds.

Compounds which may be tested for an ability to bind DP-107, DP-178, and/or disrupt DP-107/DP-178 interactions, and which therefore, potentially

represent antiviral compounds, include, but are not limited to, peptides made of D- and/or L-configuration amino acids (in, for example, the form of random peptide libraries; see Lam, K.S. et al., 1991, Nature 354:82-84), phosphopeptides (in, for example, the form of random or partially degenerate, directed phosphopeptide libraries; see, for example, Songyang, Z. et al., 1993, Cell 72:767-778), antibodies, and small organic or inorganic molecules. Synthetic compounds, natural products, and other sources of potentially effective materials may be screened in a variety of ways, as described in this Section. The compounds, antibodies, or other molecules identified may be tested for an ability to inhibit viral activity, utilizing, for example, viral assays such as those described, above, in Section 5.4.

Among the peptides which may be tested are soluble peptides comprising DP-107 and/or DP-178 domains, and peptides comprising DP-107 and/or DP-178 domains having one or more mutations within one or both of the domains, such as the M41-P peptide described, below, in the Example presented in Section 8, which contains a isoleucine to proline mutation within the DP-178 sequence.

In one embodiment of such screening methods is a method for identifying a compound to be tested for antiviral ability comprising:

- (a) exposing at least one compound to a peptide comprising a DP-107 peptide for a time sufficient to allow binding of the compound to the DP-107 peptide;
- (b) removing non-bound compounds; and
- (c) determining the presence of the compound bound to the DP-107 peptide, thereby identifying an agent to be tested for antiviral ability.

In a second embodiment of such screening methods is a method for identifying a compound to be tested for antiviral ability comprising:

(a) exposing at least one compound to a peptide comprising a DP-178 peptide for a time
5 sufficient to allow binding of the compound to the DP-178 peptide;

(b) removing non-bound compounds; and

(c) determining the presence of the compound bound to the DP-178 peptide,
10 thereby identifying an agent to be tested for antiviral ability.

One method utilizing these types of approaches that may be pursued in the isolation of such DP-107-binding or DP-178-binding compounds is an assay which
15 would include the attachment of either the DP-107 or the DP-178 peptide to a solid matrix, such as, for example, agarose or plastic beads, microtiter plate wells, petri dishes, or membranes composed of, for example, nylon or nitrocellulose. In such an assay
20 system, either the DP-107 or DP-178 protein may be anchored onto a solid surface, and the compound, or test substance, which is not anchored, is labeled, either directly or indirectly. In practice, microtiter plates are conveniently utilized. The
25 anchored component may be immobilized by non-covalent or covalent attachments. Non-covalent attachment may be accomplished simply by coating the solid surface with a solution of the protein and drying.

Alternatively, an immobilized antibody, preferably a
30 monoclonal antibody, specific for the protein may be used to anchor the protein to the solid surface. The surfaces may be prepared in advance and stored.

In order to conduct the assay, the labeled compound is added to the coated surface containing the
35 anchored DP-107 or DP-178 peptide. After the reaction

is complete, unreacted components are removed (e.g., by washing) under conditions such that any complexes formed will remain immobilized on the solid surface. The detection of complexes anchored on the solid surface can be accomplished in a number of ways.

5 Where the compound is pre-labeled, the detection of label immobilized on the surface indicates that complexes were formed. Where the labeled component is not pre-labeled, an indirect label can be used to detect complexes anchored on the surface; e.g., using
10 a labeled antibody specific for the compound (the antibody, in turn, may be directly labeled or indirectly labeled with a labeled anti-Ig antibody).

Alternatively, such an assay can be conducted in a liquid phase, the reaction products separated from
15 unreacted components, and complexes detected; e.g., using an immobilized antibody specific for DP-107 or DP-178, whichever is appropriate for the given assay, or an antibody specific for the compound, i.e., the test substance, in order to anchor any complexes
20 formed in solution, and a labeled antibody specific for the other member of the complex to detect anchored complexes.

By utilizing procedures such as this, large numbers of types of molecules may be simultaneously
25 screened for DP-107 or DP-178-binding capability, and thus potential antiviral activity.

Further, compounds may be screened for an ability to inhibit the formation of or, alternatively, disrupt DP-107/DP-178 complexes. Such compounds may then be
30 tested for antiviral capability. For ease of description, DP-107 and DP-178 will be referred to as "binding partners." Compounds that disrupt such interactions may exhibit antiviral activity. Such compounds may include, but are not limited to
35

molecules such as antibodies, peptides, and the like described above.

The basic principle of the assay systems used to identify compounds that interfere with the interaction between the DP-107 and DP-178 peptides involves
5 preparing a reaction mixture containing peptides under conditions and for a time sufficient to allow the two peptides to interact and bind, thus forming a complex. In order to test a compound for disruptive activity, the reaction is conducted in the presence and absence
10 of the test compound, i.e., the test compound may be initially included in the reaction mixture, or added at a time subsequent to the addition of one of the binding partners; controls are incubated without the test compound or with a placebo. The formation of any
15 complexes between the binding partners is then detected. The formation of a complex in the control reaction, but not in the reaction mixture containing the test compound indicates that the compound interferes with the interaction of the DP-107 and
20 DP-178 peptides.

The assay for compounds that interfere with the interaction of the binding partners can be conducted in a heterogeneous or homogeneous format. Heterogeneous assays involve anchoring one of the
25 binding partners onto a solid phase and detecting complexes anchored on the solid phase at the end of the reaction. In homogeneous assays, the entire reaction is carried out in a liquid phase. In either approach, the order of addition of reactants can be
30 varied to obtain different information about the compounds being tested. For example, test compounds that interfere with the interaction between the binding partners, e.g., by competition, can be identified by conducting the reaction in the presence
35 of the test substance; i.e., by adding the test

substance to the reaction mixture prior to or simultaneously with the binding partners. On the other hand, test compounds that disrupt preformed complexes, e.g. compounds with higher binding constants that displace one of the binding partners
5 from the complex, can be tested by adding the test compound to the reaction mixture after complexes have been formed. The various formats are described briefly below.

10 In a heterogeneous assay system, one binding partner, e.g., either the DP-107 or DP-178 peptide, is anchored onto a solid surface, and its binding partner, which is not anchored, is labeled, either directly or indirectly. In practice, microtiter
15 plates are conveniently utilized. The anchored species may be immobilized by non-covalent or covalent attachments. Non-covalent attachment may be accomplished simply by coating the solid surface with a solution of the protein and drying. Alternatively,
20 an immobilized antibody specific for the protein may be used to anchor the protein to the solid surface. The surfaces may be prepared in advance and stored.

In order to conduct the assay, the binding partner of the immobilized species is added to the coated surface with or without the test compound.
25 After the reaction is complete, unreacted components are removed (e.g., by washing) and any complexes formed will remain immobilized on the solid surface. The detection of complexes anchored on the solid surface can be accomplished in a number of ways.
30 Where the binding partner was pre-labeled, the detection of label immobilized on the surface indicates that complexes were formed. Where the binding partner is not pre-labeled, an indirect label can be used to detect complexes anchored on the
35 surface; e.g., using a labeled antibody specific for

the binding partner (the antibody, in turn, may be directly labeled or indirectly labeled with a labeled anti-Ig antibody). Depending upon the order of addition of reaction components, test compounds which inhibit complex formation or which disrupt preformed complexes can be detected.

Alternatively, the reaction can be conducted in a liquid phase in the presence or absence of the test compound, the reaction products separated from unreacted components, and complexes detected; e.g., using an immobilized antibody specific for one binding partner to anchor any complexes formed in solution, and a labeled antibody specific for the other binding partner to detect anchored complexes. Again, depending upon the order of addition of reactants to the liquid phase, test compounds which inhibit complex or which disrupt preformed complexes can be identified.

In an alternate embodiment of the invention, a homogeneous assay can be used. In this approach, a preformed complex of the DP-107 and DP-178 peptides is prepared in which one of the binding partners is labeled, but the signal generated by the label is quenched due to complex formation (see, e.g., U.S. Patent No. 4,109,496 by Rubenstein which utilizes this approach for immunoassays). The addition of a test substance that competes with and displaces one of the binding partners from the preformed complex will result in the generation of a signal above background. In this way, test substances which disrupt DP-107/DP-178 protein-protein interaction can be identified.

5.5 PHARMACEUTICAL FORMULATIONS, DOSAGES AND MODES OF ADMINISTRATION

With respect to HIV, the peptides of the invention may be used as a therapeutic in the

treatment of AIDS. The peptides of the invention may be administered using techniques well known to those in the art. Preferably, agents are formulated and administered systemically. Techniques for formulation and administration may be found in "Remington's
5 Pharmaceutical Sciences", 18th ed., 1990, Mack Publishing Co., Easton, PA. Suitable routes may include oral, rectal, transmucosal, or intestinal administration; parenteral delivery, including
10 intramuscular, subcutaneous, intramedullary injections, as well as intrathecal, direct intraventricular, intravenous, intraperitoneal, intranasal, or intraocular injections, just to name a few. Most preferably, administration is intravenous. For injection, the agents of the invention may be
15 formulated in aqueous solutions, preferably in physiologically compatible buffers such as Hanks' solution, Ringer's solution, or physiological saline buffer. For such transmucosal administration, penetrants appropriate to the barrier to be permeated
20 are used in the formulation. Such penetrants are generally known in the art.

In addition, the peptides may be used as a prophylactic measure in previously uninfected individuals after acute exposure to an HIV virus.
25 Examples of such prophylactic use of the peptides may include, but are not limited to, prevention of virus transmission from mother to infant and other settings where the likelihood of HIV transmission exists, such as, for example, accidents in health care settings
30 wherein workers are exposed to HIV-containing blood products. The peptides of the invention in such cases may serve the role of a prophylactic vaccine, wherein the host raises antibodies against the peptides of the invention, which then serve to neutralize HIV viruses
35 by, for example, inhibiting further HIV infection.

Administration of the peptides of the invention as a prophylactic vaccine, therefore, would comprise administering to a host a concentration of peptides effective in raising an immune response which is sufficient to neutralize HIV, by, for example,
5 inhibiting HIV ability to infect cells. The exact concentration will depend upon the specific peptide to be administered, but may be determined by using standard techniques for assaying the development of an immune response which are well known to those of
10 ordinary skill in the art. The peptides to be used as vaccines are usually administered intramuscularly.

The peptides may be formulated with a suitable adjuvant in order to enhance the immunological response. Such adjuvants may include, but are not
15 limited to mineral gels such as aluminum hydroxide; surface active substances such as lysolecithin, pluronic polyols, polyanions; other peptides; oil emulsions; and potentially useful human adjuvants such as BCG and Corynebacterium parvum. Many methods may
20 be used to introduce the vaccine formulations described here. These methods include but are not limited to oral, intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, and intranasal routes.

25 Alternatively, an effective concentration of polyclonal or monoclonal antibodies raised against the peptides of the invention may be administered to a host so that no uninfected cells become infected by HIV. The exact concentration of such antibodies will
30 vary according to each specific antibody preparation, but may be determined using standard techniques well known to those of ordinary skill in the art. Administration of the antibodies may be accomplished using a variety of techniques, including, but not
35 limited to those described in this section.

Effective dosages of the peptides of the invention to be administered may be determined through procedures well known to those in the art which address such parameters as biological half-life, bioavailability, and toxicity. Given the data
5 presented below in Section 6, DP-178, for example, may prove efficacious in vivo at doses required achieve circulating levels of 10ng per ml of peptide.

A therapeutically effective dose refers to that amount of the compound sufficient to result in
10 amelioration of symptoms or a prolongation of survival in a patient. Toxicity and therapeutic efficacy of such compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD50
15 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Compounds
20 which exhibit large therapeutic indices are preferred. The data obtained from these cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of
25 circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the
30 therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (i.e., the concentration of the test compound which
35 achieves a half-maximal disruption of the PTK/adaptor

protein complex, or a half-maximal inhibition of the cellular level and/or activity of a complex component) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for
5 example, by high performance liquid chromatography (HPLC).

The exact formulation, route of administration and dosage can be chosen by the individual physician in view of the patient's condition. (See e.g. Fingl
10 et al., 1975, in "The Pharmacological Basis of Therapeutics", Ch. 1 p1).

It should be noted that the attending physician would know how to and when to terminate, interrupt, or adjust administration due to toxicity, or to organ
15 dysfunctions. Conversely, the attending physician would also know to adjust treatment to higher levels if the clinical response were not adequate (precluding toxicity). The magnitude of an administered dose in the management of the oncogenic disorder of interest
20 will vary with the severity of the condition to be treated and to the route of administration. The dose and perhaps dose frequency, will also vary according to the age, body weight, and response of the individual patient. A program comparable to that
25 discussed above may be used in veterinary medicine.

As demonstrated in the Example presented below in Section 6, the antiviral activity of the peptides of the invention may show a pronounced type and subtype specificity, i.e., specific peptides may be effective
30 in inhibiting the activity of only specific viruses. This feature of the invention presents many advantages. One such advantage, for example, lies in the field of diagnostics, wherein one can use the antiviral specificity of the peptide of the invention
35 to ascertain the identity of a viral isolate. With

respect to HIV, one may easily determine whether a viral isolate consists of an HIV-1 or HIV-2 strain. For example, uninfected CD-4⁺ cells may be co-infected with an isolate which has been identified as containing HIV the DP-178 (SEQ ID:1) peptide, after
5 which the retroviral activity of cell supernatants may be assayed, using, for example, the techniques described above in Section 5.2. Those isolates whose retroviral activity is completely or nearly completely inhibited contain HIV-1. Those isolates whose viral
10 activity is unchanged or only reduced by a small amount, may be considered to not contain HIV-1. Such an isolate may then be treated with one or more of the other DP-178 peptides of the invention, and subsequently be tested for its viral activity in order
15 to determine the identify of the viral isolate.

Use of pharmaceutically acceptable carriers to formulate the compounds herein disclosed for the practice of the invention into dosages suitable for systemic administration is within the scope of the
20 invention. With proper choice of carrier and suitable manufacturing practice, the compositions of the present invention, in particular, those formulated as solutions, may be administered parenterally, such as by intravenous injection. The compounds can be
25 formulated readily using pharmaceutically acceptable carriers well known in the art into dosages suitable for oral administration. Such carriers enable the compounds of the invention to be formulated as tablets, pills, capsules, liquids, gels, syrups,
30 slurries, suspensions and the like, for oral ingestion by a patient to be treated.

Pharmaceutical compositions suitable for use in the present invention include compositions wherein the active ingredients are contained in an effective
35 amount to achieve its intended purpose. Determination

of the effective amounts is well within the capability of those skilled in the art, especially in light of the detailed disclosure provided herein.

In addition to the active ingredients, these pharmaceutical compositions may contain suitable
5 pharmaceutically acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. The preparations formulated for oral administration may be in the form of tablets,
10 dragees, capsules, or solutions.

The pharmaceutical compositions of the present invention may be manufactured in a manner that is itself known, e.g., by means of conventional mixing, dissolving, granulating, dragee-making, levigating,
15 emulsifying, encapsulating, entrapping or lyophilizing processes.

Pharmaceutical formulations for parenteral administration include aqueous solutions of the active compounds in water-soluble form. Additionally,
20 suspensions of the active compounds may be prepared as appropriate oily injection suspensions. Suitable lipophilic solvents or vehicles include fatty oils such as sesame oil, or synthetic fatty acid esters, such as ethyl oleate or triglycerides, or liposomes.
25 Aqueous injection suspensions may contain substances which increase the viscosity of the suspension, such as sodium carboxymethyl cellulose, sorbitol, or dextran. Optionally, the suspension may also contain suitable stabilizers or agents which increase the
30 solubility of the compounds to allow for the preparation of highly concentrated solutions.

Pharmaceutical preparations for oral use can be obtained by combining the active compounds with solid excipient, optionally grinding a resulting mixture,
35 and processing the mixture of granules, after adding

suitable auxiliaries, if desired, to obtain tablets or dragee cores. Suitable excipients are, in particular, fillers such as sugars, including lactose, sucrose, mannitol, or sorbitol; cellulose preparations such as, for example, maize starch, wheat starch, rice starch, potato starch, gelatin, gum tragacanth, methyl cellulose, hydroxypropylmethyl-cellulose, sodium carboxymethylcellulose, and/or polyvinylpyrrolidone (PVP). If desired, disintegrating agents may be added, such as the cross-linked polyvinyl pyrrolidone, agar, or alginic acid or a salt thereof such as sodium alginate.

Dragee cores are provided with suitable coatings. For this purpose, concentrated sugar solutions may be used, which may optionally contain gum arabic, talc, polyvinyl pyrrolidone, carbopol gel, polyethylene glycol, and/or titanium dioxide, lacquer solutions, and suitable organic solvents or solvent mixtures. Dyestuffs or pigments may be added to the tablets or dragee coatings for identification or to characterize different combinations of active compound doses.

Pharmaceutical preparations which can be used orally include push-fit capsules made of gelatin, as well as soft, sealed capsules made of gelatin and a plasticizer, such as glycerol or sorbitol. The push-fit capsules can contain the active ingredients in admixture with filler such as lactose, binders such as starches, and/or lubricants such as talc or magnesium stearate and, optionally, stabilizers. In soft capsules, the active compounds may be dissolved or suspended in suitable liquids, such as fatty oils, liquid paraffin, or liquid polyethylene glycols. In addition, stabilizers may be added.

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6. EXAMPLE: DP-178 (SEQ ID:1) IS A POTENT
INHIBITOR OF HIV-1 INFECTION

In this example, DP-178 (SEQ ID:1) is shown to be a potent inhibitor of HIV-1 mediated CD-4⁺ cell-cell fusion and infection by cell free virus. In the fusion assay, this peptide completely blocks virus induced syncytia formation at concentrations of from 1-10 ng/ml. In the infectivity assay the inhibitory concentration is somewhat higher, blocking infection at 90ng/ml. It is further shown that DP-178 (SEQ ID:1) shows that the antiviral activity of DP-178 (SEQ ID:1) is highly specific for HIV-1. Additionally, a synthetic peptide, DP-185 (SEQ ID:3), representing a HIV-1-derived DP-178 homolog is also found to block HIV-1-mediated syncytia formation.

6.1. MATERIALS AND METHODS

6.1.1. PEPTIDE SYNTHESIS

Peptides were synthesized using Fast Moc chemistry on an Applied Biosystems Model 431A peptide synthesizer. Amidated peptides were prepared using Rink resin (Advanced Chemtech) while peptides containing free carboxy termini were synthesized on Wang (p-alkoxy-benzyl-alcohol) resin (Bachem). First residues were double coupled to the appropriate resin and subsequent residues were single coupled. Each coupling step was followed by acetic anhydride capping. Peptides were cleaved from the resin by treatment with trifluoroacetic acid (TFA) (10ml), H₂O (0.5ml), thioanisole (0.5ml), ethanedithiol (0.25ml), and crystalline phenol (0.75g). Purification was carried out by reverse phase HPLC. Approximately 50mg samples of crude peptide were chromatographed on a Waters Delta Pak C18 column (19mm x 30cm, 15 μ spherical) with a linear gradient; H₂O/acetonitrile

0.1% TFA. Lyophilized peptides were stored desiccated and peptide solutions were made in water at about 1mg/ml. Electrospray mass spectrometry yielded the following results: DP-178 (SEQ ID:1):4491.87 (calculated 4491.94); DP-180 (SEQ ID:2):4491.45 (calculated 4491.94); DP-185 (SEQ ID:3):not done (calculated 4546.97).

6.1.2. VIRUS

The HIV-1_{LAI} virus was obtained from R. Gallo (Popovic, M. et al., 1984, Science 224:497-508) and propagated in CEM cells cultured in RPMI 1640 containing 10% fetal calf serum. Supernatant from the infected CEM cells was passed through a 0.2 μ m filter and the infectious titer estimated in a microinfectivity assay using the AA5 cell line to support virus replication. For this purpose, 25 μ l of serial diluted virus was added to 75 μ l AA5 cells at a concentration of 2×10^5 /ml in a 96-well microtitre plate. Each virus dilution was tested in triplicate. Cells were cultured for eight days by addition of fresh medium every other day. On day 8 post infection, supernatant samples were tested for virus replication as evidenced by reverse transcriptase activity released to the supernatant. The TCID₅₀ was calculated according to the Reed and Muench formula (Reed, L.J. et al., 1938, Am. J. Hyg. 27:493-497). The titer of the HIV-1_{LAI} and HIV-1_{MN} stocks used for these studies, as measured on the AA5 cell line, was approximately 1.4×10^6 and 3.8×10^4 TCID₅₀/ml, respectively.

6.1.3. CELL FUSION ASSAY

Approximately 7×10^4 Molt cells were incubated with 1×10^4 CEM cells chronically infected with the HIV-1_{LAI} virus in 96-well plates (one-half area cluster plates; Costar, Cambridge, MA) in a final volume of

100 μ l culture medium as previously described (Matthews, T.J. et al., 1987, Proc. Natl. Acad. Sci. USA 84: 5424-5428). Peptide inhibitors were added in a volume of 10 μ l and the cell mixtures were incubated for 24 hr. at 37°C. At that time, multinucleated
5 giant cells were estimated by microscopic examination at a 40x magnification which allowed visualization of the entire well in a single field.

6.1.4. CELL FREE VIRUS INFECTION ASSAY

10 Synthetic peptides were incubated at 37°C with either 247 TCID₅₀ (for experiment depicted in FIG. 2), or 62 TCID₅₀ (for experiment depicted in FIG.3) units of HIV-1_{LAI} virus or 25 TCID₅₀ units of HIV-2_{NIH2} and CEM CD4⁺ cells at peptide concentrations of 0, 0.04, 0.4,
15 4.0, and 40 μ g/ml for 7 days. The resulting reverse transcriptase (RT) activity in counts per minute was determined using the assay described, below, in Section 6.1.5. See, Reed, L.J. et al., 1938, Am. J. Hyg. 27: 493-497 for an explanation of TCID₅₀
20 calculations.

6.1.5. REVERSE TRANSCRIPTASE ASSAY

The micro-reverse transcriptase (RT) assay was adapted from Goff et al. (Goff, S. et al., 1981, J. Virol. 38:239-248) and Willey et al. (Willey, R. et al., 1988, J. Virol. 62:139-147). Supernatants from virus/cell cultures are adjusted to 1% Triton-X100. A
25 10 μ l sample of supernatant was added to 50 μ l of RT cocktail in a 96-well U-bottom microtitre plate and the samples incubated at 37°C for 90 min. The RT
30 cocktail contained 75mM KCl, 2mM dithiothreitol, 5mM MgCl₂, 5 μ g/ml poly A (Pharmacia, cat. No. 27-4110-01), 0.25 units/ml oligo dT (Pharmacia, cat. No. 27-7858-01), 0.05% NP40, 50mM Tris-HCl, pH 7.8, 0.5 μ M non-
35

radioactive dTTP, and 10 μ Ci/ml ³²P-dTTP (Amersham, cat. No. PB.10167).

After the incubation period, 40 μ l of reaction mixture was applied to a Schleicher and Schuell (S+S) NA45 membrane (or DE81 paper) saturated in 2 x SSC buffer (0.3M NaCl and 0.003M sodium citrate) held in a S+S Minifold over one sheet of GB003 (S+S) filter paper, with partial vacuum applied. Each well of the minifold was washed four times with 200 μ l 2xSSC, under full vacuum. The membrane was removed from the minifold and washed 2 more times in a pyrex dish with an excess of 2xSSC. Finally, the membrane was drained on absorbent paper, placed on Whatman #3 paper, covered with Saran wrap, and exposed to film overnight at -70°C.

6.2. RESULTS

6.2.1. PEPTIDE INHIBITION OF INFECTED CELL-INDUCED SYNCYTIA FORMATION

The initial screen for antiviral activity assayed peptides' ability to block syncytium formation induced by overnight co-cultivation of uninfected Molt4 cells with chronically HIV-1 infected CEM cells. The results of several such experiments are presented herein. In the first of these experiments, serial DP-178 (SEQ ID:1) peptide concentrations between 10 μ g/ml and 12.5ng/ml were tested for blockade of the cell fusion process. For these experiments, CEM cells chronically infected with either HIV-1_{LAI}, HIV-1_{MN}, HIV-1_{RF}, or HIV-1_{SF2} virus were cocultivated overnight with uninfected Molt 4 cells. The results (FIG. 4) show that DP-178 (SEQ ID:1) afforded complete protection against each of the HIV-1 isolates down to the lowest concentration of DP-178 (SEQ ID:1) used. For HIV_{LAI} inhibition, the lowest concentration tested was

12.5ng/ml; for all other HIV-1 viruses, the lowest concentration of DP-178 (SEQ ID:1) used in this study was 100ng/ml. A second peptide, DP-180 (SEQ ID:2), containing the same amino acid residues as DP-178 (SEQ ID:1) but arranged in a random order exhibited no
5 evidence of anti-fusogenic activity even at the high concentration of 40µg/ml (FIG. 4). These observations indicate that the inhibitory effect of DP-178 (SEQ ID:1) is primary sequence-specific and not related to non-specific peptide/protein interactions. The actual
10 endpoint (i.e., the lowest effective inhibitory concentration) of DP-178 inhibitory action is within the range of 1-10 ng/ml.

The next series of experiments involved the preparation and testing of a DP-178 (SEQ ID:1) homolog
15 for its ability to inhibit HIV-1-induced syncytia formation. As shown in FIG. 1, the sequence of DP-185 (SEQ ID:3) is slightly different from DP-178 (SEQ ID:1) in that its primary sequence is taken from the HIV-1_{SP2} isolate and contains several amino acid
20 differences relative to DP-178 (SEQ ID:1) near the N terminus. As shown in FIG. 4, DP-185 (SEQ ID:3), exhibits inhibitory activity even at 312.5ng/ml, the lowest concentration tested.

The next series of experiments involved a
25 comparison of DP-178 (SEQ ID:1) HIV-1 and HIV-2 inhibitory activity. As shown in FIG. 5, DP-178 (SEQ ID:1) blocked HIV-1-mediated syncytia formation at peptide concentrations below 1ng/ml. DP-178 (SEQ ID:1) failed, however, to block HIV-2 mediated
30 syncytia formation at concentrations as high as 10µg/ml. This striking 4 log selectivity of DP-178 (SEQ ID:1) as an inhibitor of HIV-1-mediated cell fusion demonstrates an unexpected HIV-1 specificity in the action of DP-178 (SEQ ID:1). DP-178 (SEQ ID:1)
35 inhibition of HIV-1-mediated cell fusion, but the

peptide's inability to inhibit HIV-2 medicated cell fusion in the same cell type at the concentrations tested provides further evidence for the high degree of selectivity associated with the antiviral action of DP-178 (SEQ ID:1).

5

6.2.2. PEPTIDE INHIBITION OF INFECTION BY CELL-FREE VIRUS

DP-178 (SEQ ID:1) was next tested for its ability to block CD-4⁺ CEM cell infection by cell free HIV-1 virus. The results, shown in FIG. 2, are from an experiment in which DP-178 (SEQ ID:1) was assayed for its ability to block infection of CEM cells by an HIV-1_{LAI} isolate. Included in the experiment were three control peptides, DP-116 (SEQ ID:9), DP-125 (SEQ ID:8), and DP-118 (SEQ ID:10). DP-116 (SEQ ID:9) represents a peptide previously shown to be inactive using this assay, and DP-125 (SEQ ID:8; Wild, C. et al., 1992, Proc. Natl. Acad. Sci. USA 89:10,537) and DP-118 (SEQ ID:10) are peptides which have previously been shown to be active in this assay. Each concentration (0, 0.04, 0.4, 4, and 40 μ g/ml) of peptide was incubated with 247 TCID₅₀ units of HIV-1_{LAI} virus and CEM cells. After 7 days of culture, cell-free supernatant was tested for the presence of RT activity as a measure of successful infection. The results, shown in FIG. 2, demonstrate that DP-178 (SEQ ID:1) inhibited the de novo infection process mediated by the HIV-1 viral isolate at concentrations as low as 90ng/ml (IC₅₀=90ng/ml). In contrast, the two positive control peptides, DP-125 (SEQ ID:8) and DP-118 (SEQ ID:10), had over 60-fold higher IC₅₀ concentrations of approximately 5 μ g/ml.

In a separate experiment, the HIV-1 and HIV-2 inhibitory action of DP-178 (SEQ ID:1) was tested with CEM cells and either HIV-1_{LAI} or HIV-2_{NIH}. 62 TCID₅₀

HIV-1_{LAI} or 25 GCID₅₀ HIV-2_{NIH2} were used in these experiments, and were incubated for 7 days. As may be seen in FIG. 3, DP-178 (SEQ ID:1) inhibited HIV-1 infection with an IC50 of about 31ng/ml. In contrast, DP-178 (SEQ ID:1) exhibited a much higher IC50 for HIV-2_{NIH2}, thus making DP-178 (SEQ ID:1) two logs more potent as a HIV-1 inhibitor than a HIV-2 inhibitor. This finding is consistent with the results of the fusion inhibition assays described, above, in Section 6.2.1, and further supports a significant level of selectivity (i.e., for HIV-1 over HIV-2).

7. EXAMPLE: THE HIV-1 INHIBITOR, DP-178 (SEQ ID:1) IS NON-CYTOXIC

In this Example, the 36 amino acid synthetic peptide inhibitor DP-178 (SEQ ID:1) is shown to be non-cytotoxic to cells in culture, even at the highest peptide concentrations (40μg/ml) tested.

7.1. MATERIALS AND METHODS

Cell proliferation and toxicity assay:
Approximately 3.8x10⁵ CEM cells for each peptide concentration were incubated for 3 days at 37°C in T25 flasks. Peptides tested were DP-178 (SEQ ID:1) and DP-116 (SEQ ID:9), as described in FIG. 1. The concentrations of each peptide used were 0, 2.5, 10, and 40μg/ml. Cell counts were taken at incubation times of 0, 24, 48, and 72 hours.

7.2. RESULTS

Whether the potent HIV-1 inhibitor DP-178 (SEQ ID:1) exhibited any cytotoxic effects was assessed by assaying the peptide's effects on the proliferation and viability of cells in culture. CEM cells were incubated in the presence of varying concentrations of DP-178 (SEQ ID:1), and DP-116 (SEQ ID:9), a peptide

previously shown to be ineffective as a HIV inhibitor (Wild, C. et al., 1992, Proc. Natl. Acad. Sci. USA 89:10,537-10,541). Additionally, cells were incubated in the absence of either peptide.

5 The results of the cytotoxicity study demonstrate that DP-178 (SEQ ID:1) exhibits no cytotoxic effects on cells in culture. As can be seen, below, in Table XI, even the proliferation and viability
10 characteristics of cells cultured for 3 days in the presence of the highest concentration of DP-178 (SEQ ID:1) tested (40 μ g/ml) do not significantly differ from the DP-116 (SEQ ID:9) or the no-peptide controls. The cell proliferation data is also represented in
15 graphic form in FIG. 6. As was demonstrated in the Working Example presented above in Section 6, DP-178 (SEQ ID:1) completely inhibits HIV-1 mediated syncytia formation at peptide concentrations between 1 and 10ng/ml, and completely inhibits cell-free viral infection at concentrations of at least 90ng/ml. Thus, this study demonstrates that even at peptide
20 concentrations greater than 3 log higher than the HIV inhibitory dose, DP-178 (SEQ ID:1) exhibits no cytotoxic effects.

TABLE XI

Peptide	Peptide Concentration μ g/ml	% Viability at time (hours)			
		0	24	48	72
DP178 (SEQ ID:1)	40	98	97	95	97
	10	98	97	98	98
	2.5	98	93	96	96

	DP116 (SEQ ID:9)	40	98	95	98	97
		10	98	95	93	98
5		2.5	98	96	98	99
	No Peptide	0	98	97	99	98

10 8. EXAMPLE: THE INTERACTION OF DP178 AND DP107

Soluble recombinant forms of gp41 used in the
 example described below provide evidence that the
 DP178 peptide associates with a distal site on gp41
 whose interactive structure is influenced by the DP107
 leucine zipper motif. A single mutation disrupting
 15 the coiled-coil structure of the leucine zipper domain
 transformed the soluble recombinant gp41 protein from
 an inactive to an active inhibitor of HIV-1 fusion.
 This transformation may result from liberation of the
 potent DP178 domain from a molecular clasp with the
 20 leucine zipper, DP107, determinant. The results also
 indicate that the anti-HIV activity of various gp41
 derivatives (peptides and recombinant proteins) may be
 due to their ability to form complexes with viral gp41
 25 and interfere with its fusogenic process.

8.1. MATERIALS AND METHODS

8.1.1. CONSTRUCTION OF FUSION PROTEINS AND GP41 MUTANTS

30 Construction of fusion proteins and mutants shown
 in FIG. 7 was accomplished as follows: the DNA
 sequence corresponding to the extracellular domain of
 gp41 (540-686) was cloned into the Xmn I site of the
 expression vector pMal-p2 (New England Biolab) to give
 35 M41. The gp41 sequence was amplified from pgtat

(Malim et al., 1988, Nature 355: 181-183) by using polymerase chain reaction (PCR) with upstream primer 5'-ATGACGCTGACGGTACAGGCC-3' (primer A) and downstream primer 5'-TGACTAAGCTTAATACCACAGCCAATTTGTTAT-3' (primer B). M41-P was constructed by using the T7-Gen
5 in vitro mutagenesis kit from United States Biochemicals (USB) following the supplier's instructions. The mutagenic primer (5'-GGAGCTGCTTGGGGCCCCAGAC-3') introduces an Ile to Pro mutation in M41 at position 578. M41Δ107 was made
10 using a deletion mutagenic primer 5'-CCAAATCCCCAGGAGCTGCTCGAGCTGCACTATACCAGAC-3' (primer C) following the USB T7-Gen mutagenesis protocol. M41Δ178 was made by cloning the DNA fragment
15 corresponding to gp41 amino acids 540-642 into the Xmn I site of pMal-p2. Primer A and 5'-ATAGCTTCTAGATTAATTGTTAATTTCTCTGTCCC-3' (primer D) were used in the PCR with the template pgtat to generate the inserted DNA fragments. M41-P was used as the template with primer A and D in PCR to generate M41-
20 PA178. All inserted sequences and mutated residues were checked by restriction enzyme analysis and confirmed by DNA sequencing.

25 8.1.2. PURIFICATION AND CHARACTERIZATION OF FUSION PROTEINS

The fusion proteins were purified according to the protocol described in the manufacturer's brochure of protein fusion and purification systems from New England Biolabs (NEB). Fusion proteins (10 ng) were
30 analyzed by electrophoresis on 8% SDS polyacrylamide gels. Western blotting analysis was performed as described by Sambrook et al, 1989, Molecular Cloning: A Laboratory Manual, 2d Ed, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, Ch. 18,
35 pp. 64-75. An HIV-1 positive serum diluted 1000-fold,

or a human Fab derived from repertoire cloning was used to react with the fusion proteins. The second antibody was HRP-conjugated goat antihuman Fab. An ECL Western blotting detection system (Amersham) was used to detect the bound antibody. A detailed
5 protocol for this detection system was provided by the manufacturer. Rainbow molecular weight marker (Amersham) were used to estimate the size of fusion proteins.

10 8.1.3. CELL FUSION ASSAYS FOR ANTI-HIV ACTIVITY

Cell fusion assays were performed as previously described (Matthews et al., 1987, Proc. Natl. Acad. Sci. USA 84: 5424-5481). CEM cells (7×10^4) were
15 incubated with HIV-1_{MB} chronically infected CEM cells (10^4) in 96-well flat-bottomed half-area plates (Costar) in 100 μ l culture medium. Peptide and fusion proteins at various concentrations in 10 μ l culture medium were incubated with the cell mixtures at 37°C for 24 hours. Multinucleated syncytia were estimated
20 with microscopic examination. Both M41 and M41-P did not show cytotoxicity at the concentrations tested and shown in FIG. 8.

Inhibition of HIV-1 induced cell-cell fusion activity was carried out in the presence of 10 nM
25 DP178 and various concentrations of M41 Δ 178 or M41-PA178 as indicated in FIG. 9. There was no observable syncytia in the presence of 10 nM DP178. No peptide or fusion protein was added in the control samples.

30 8.1.4. ELISA ANALYSIS OF DP178 BINDING TO THE LEUCINE ZIPPER MOTIF OF GP41

The amino acid sequence of DP178 used is:
YTSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF. For enzyme
linked immunoassay (ELISA), M41 Δ 178 or M41-PA178 (5
35 μ g/ml) in 0.1M NaHCO₃, pH 8.6, were coated on 96 wells

Linbro ELISA plates (Flow Lab, Inc.) overnight. Each well was washed three times with distilled water then blocked with 3% bovine serum albumin (BSA) for 2 hours. After blocking, peptides with 0.5% BSA in TBST (40 mM Tris-HCl pH7.5, 150 mM NaCl, 0.05% Tween 20) were added to the ELISA plates and incubated at room temperature for 1 hour. After washing three times with TBST, Fab-d was added at a concentration of 10 ng/ml with 0.5% BSA in TBST. The plates were washed three times with TBST after incubation at room temperature for 1 hour. Horse radish peroxidase (HRP) conjugated goat antihuman Fab antiserum at a 2000 fold dilution in TBST with 0.5% BSA was added to each well and incubated at room temperature for 45 minutes. The plates were then washed four times with TBST. The peroxidase substrate o-phenylene diamine (2.5 mg/ml) and 0.15% H₂O₂ were added to develop the color. The reaction was stopped with an equal volume of 4.5 N H₂SO₄ after incubation at room temperature for 10 minutes. The optical density of the stopped reaction mixture was measured with a micro plate reader (Molecular Design) at 490 nm. Results are shown in FIG. 10.

8.2. RESULTS

25

8.2.1. THE EXPRESSION AND CHARACTERIZATION OF THE ECTODOMAIN OF GP41

As a step toward understanding the roles of the two helical regions in gp41 structure and function, the ectodomain of gp41 was expressed as a maltose binding fusion protein (M41) (Fig. 7). The fusogenic peptide sequence at the N-terminal of gp41 was omitted from this recombinant protein and its derivatives to improve solubility. The maltose binding protein facilitated purification of the fusion proteins under relatively mild, non-denaturing conditions. Because

the M41 soluble recombinant gp41 was not glycosylated, lacked several regions of the transmembrane protein (*i.e.*, the fusion peptide, the membrane spanning, and the cytoplasmic domains), and was expressed in the absence of gp120, it was not expected to precisely reflect the structure of native gp41 on HIV-1 virions. Nevertheless, purified M41 folded in a manner that preserved certain discontinuous epitopes as evidenced by reactivity with human monoclonal antibodies, 98-6, 126-6, and 50-69, previously shown to bind conformational epitopes on native gp41 expressed in eukaryotic cells (Xu et al., 1991, J. Virol. 65: 4832-4838; Chen, 1994, J. Virol. 68:2002-2010). Thus, at least certain regions of native gp41 defined by these antibodies appear to be reproduced in the recombinant fusion protein M41. Furthermore, M41 reacted with a human recombinant Fab (Fab-d) that recognizes a conformational epitope on gp41 and binds HIV-1 virions as well as HIV-1 infected cells but not uninfected cells as analyzed by FACS. Deletion of either helix motif, *i.e.*, DP107 or DP178, of the M41 fusion protein eliminated reactivity with Fab-d. These results indicate that both helical regions, separated by 60 amino acids in the primary sequence, are required to maintain the Fab-d epitope.

8.2.2. ANTI-HIV ACTIVITY OF THE RECOMBINANT ECTODOMAIN OF GP41

The wild type M41 fusion protein was tested for anti-HIV-1 activity. As explained, *supra*, synthetic peptides corresponding to the leucine zipper (DP107) and the C-terminal putative helix (DP178) show potent anti-HIV activity. Despite inclusion of both these regions, the recombinant M41 protein did not affect

HIV-1 induced membrane fusion at concentrations as high as 50 μ M (Table XII, below).

TABLE XII

DISRUPTION OF THE LEUCINE ZIPPER OF
GP41 FREES THE ANTI-HIV MOTIF

	DP107	DP178	M41	M41-P	M41-PA178
Cell fusion (IC ₅₀)	1 μ M	1 nM	> 50 μ M	83 nM	> 50 μ M
Fab-D binding (k _D)	-	-	3.5x10 ⁻⁹	2.5x10 ⁻⁸	-
HIV infectiv- ity (IC ₅₀)	1 μ M	80 nM	> 16 μ M	66 nM	> 8 μ M

The affinity constants of Fab-d binding to the fusion proteins were determined using a protocol described by B. Friguet et al., 1985, J. Immunol. Method. 77:305-319.

- = No detectable binding of Fab-d to the fusion proteins.

Antiviral Infectivity Assays. 20 μ l of serially diluted virus stock was incubated for 60 minutes at ambient temperature with 20 μ l of the indicated concentration of purified recombinant fusion protein in RPMI 1640 containing 10% fetal bovine serum and antibiotics in a 96-well microtiter plate. 20 μ l of CEM4 cells at 6 x 10⁵ cells/ml were added to each well, and cultures were incubated at 37°C in a humidified CO₂ incubator. Cells were cultured for 9 days by the addition of fresh medium every 2 to 30 days. On days 5, 7, and 9 postinfection, supernatant samples were assayed for reverse transcriptase (RT) activity, as described below, to monitor viral replication. The 50% tissue culture infectious dose (TCID₅₀) was calculated for each condition according to the formula of Reed & Muench, 1937, Am. J. Hyg. 27:493-497. RT activity was determined by a modification of the published methods of Goff et al., 1981, J. Virol. 38:239-248 and Willey et al., 1988, J. Virol. 62:139-147 as described in Chen et al., 1993, AIDS Res. Human Retroviruses 9:1079-1086.

Surprisingly, a single amino acid substitution, proline in place of isoleucine in the middle of the leucine zipper motif, yielded a fusion protein (M41-P)

which did exhibit antiviral activity (Table XII and Fig. 8). As seen in Table XII, M41-P blocked syncytia formation by 90% at approximately 85 nM and neutralized HIV-1_{MB} infection by 90% at approximately 70 nM concentrations. The anti-HIV-1 activity of M41-P appeared to be mediated by the C-terminal helical sequence since deletion of that region from M41-P yielded an inactive fusion protein, M41-PA178 (Table XII). That interpretation was reinforced by experiments demonstrating that a truncated fusion protein lacking the DP178 sequence, M41Δ178, abrogated the potent anti-fusion activity of the DP178 peptide in a concentration-dependent manner (FIG. 9). The same truncated fusion protein containing the proline mutation disrupting the leucine zipper, M41-PA178, was not active in similar competition experiments (FIG. 9). The results indicate that the DP178 peptide associates with a second site on gp41 whose interactive structure is dependent on a wild type leucine zipper sequence. A similar interaction may occur within the wild type fusion protein, M41, and act to form an intramolecular clasp which sequesters the DP178 region, making it unavailable for anti-viral activity.

A specific association between these two domains is also indicated by other human monoclonal Fab-d studies. For example, Fab-d failed to bind either the DP178 peptide or the fusion protein M41Δ178, but its epitope was reconstituted by simply mixing these two reagents together (FIG. 10). Again, the proline mutation in the leucine zipper domain of the fusion protein, M41-PA178, failed to reconstitute the epitope in similar mixing experiments.

9. EXAMPLE: METHOD FOR COMPUTER-ASSISTED
IDENTIFICATION OF DP-107-LIKE
AND DP-178-LIKE SEQUENCES

A number of known coiled-coil sequences have been well described in the literature and contain heptad repeat positioning for each amino acid. Coiled-coil nomenclature labels each of seven amino acids of a heptad repeat A through G, with amino acids A and D tending to be hydrophobic positions. Amino acids E and G tend to be charged. These four positions (A, D, E, and G) form the amphipathic backbone structure of a monomeric alpha-helix. The backbones of two or more amphipathic helices interact with each other to form di-, tri-, tetrameric, etc., coiled-coil structures. In order to begin to design computer search motifs, a series of well characterized coiled coils were chosen including yeast transcription factor GCN4, Influenza Virus hemagglutinin loop 36, and human proto-oncogenes c-Myc, c-Fos, and c-Jun. For each peptide sequence, a strict homology for the A and D positions, and a list of the amino acids which could be excluded for the B, C, E, F, and G positions (because they are not observed in these positions) was determined. Motifs were tailored to the DP-107 and DP-178 sequences by deducing the most likely possibilities for heptad positioning of the amino acids of HIV-1 Bru DP-107, which is known to have coiled-coil structure, and HIV-1 Bru DP-178, which is still structurally undefined. The analysis of each of the sequences is contained in FIG. 12. For example, the motif for GCN4 was designed as follows:

1. The only amino acids (using standard single letter amino acid codes) found in the A or D positions of GCN4 were [LMNV].
2. All amino acids were found at B, C, E, F, and G positions except {CFGIMPTW}.

3. The PESEARCH motif would, therefore, be written as follows:

[LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)-
 [LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)-
 [LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)-
 [LMNV]-{CFGIMPTW}(2)-[LMNV]-{CFGIMPTW}(3)

Translating or reading the motif: "at the first A position either L, M, N, or V must occur; at positions B and C (the next two positions) accept everything except C, F, G, I, M, P, T, or W; at the D position either L, M, N, or V must occur; at positions E, F, and G (the next 3 positions) accept everything except C, F, G, I, M, P, T, or W." This statement is contained four times in a 28-mer motif and five times in a 35-mer motif. The basic motif key then would be: [LMNV]-{CFGIMPTW}. The motif keys for the remaining well described coiled-coil sequences are summarized in FIG. 12.

The motif design for DP-107 and DP-178 was slightly different than the 28-mer model sequences described above due to the fact that heptad repeat positions are not defined and the peptides are both longer than 28 residues. FIG. 13 illustrates several possible sequence alignments for both DP-107 and DP-178 and also includes motif designs based on 28^{-mer}, 35^{-mer}, and full-length peptides. Notice that only slight differences occur in the motifs as the peptides are lengthened. Generally, lengthening the base peptide results in a less stringent motif. This is very useful in broadening the possibilities for identifying DP-107-or DP-178-like primary amino acid sequences referred to in this document as "hits".

In addition to making highly specific motifs for each type peptide sequence to be searched, it is also possible to make "hybrid" motifs. These motifs are

made by "crossing" two or more very stringent motifs to make a new search algorithm which will find not only both "parent" motif sequences but also any peptide sequences which have similarities to one, the other, or both "parents". For example, in Table 3 the "parent" sequence of GCN4 is crossed with each of the possible "parent" motifs of DP-107. Now the hybrid motif must contain all of the amino acids found in the A and D positions of both parents, and exclude all of the amino acids not found in either parent at the other positions. The resulting hybrid from crossing GCN4 or [LMNV]{CFGIMPTW} and DP-107 (28-mer with the first L in the D position) or [ILQT]{CDFIMPST}, is [ILMNQTV]{CFIMPT}. Notice that now only two basic hybrid motifs exist which cover both framing possibilities, as well as all peptide lengths of the parent DP-107 molecule. FIG. 15 represents the hybridizations of GCN4 with DP-178. FIG. 16 represents the hybridizations of DP-107 and DP-178. It is important to keep in mind that the represented motifs, both parent and hybrid, are motif keys and not the depiction of the full-length motif needed to actually do the computer search.

Hybridizations can be performed on any combination of two or more motifs. Table 5 summarizes several three-motif hybridizations including GCN4, DP-107 (both frames), and DP-178 (also both frames). Notice that the resulting motifs are now becoming much more similar to each other. In fact, the first and third hybrid motifs are actually subsets of the second and fourth hybrid motifs respectively. This means that the first and third hybrid motifs are slightly more stringent than the second and fourth. It should also be noted that with only minor changes in these four motifs, or by hybridizing them, a single motif could be obtained

which would find all of the sequences. However, it should be remembered that stringency is also reduced. Finally, the most broad-spectra and least-stringent hybrid motif is described in FIG. 18 which summarizes the hybridization of GCN4, DP-107 (both frames), DP-178 (both frames), c-Fos, c-Jun, c-Myc, and Flu loop 36.

A special set of motifs was designed based on the fact that DP-178 is located only approximately ten amino acids upstream of the transmembrane spanning region of gp41 and just C-terminal to a proline which separates DP-107 and DP-178. It has postulated that DP-178 may be an amphipathic helix when membrane associated, and that the proline might aid in the initiation of the helix formation. The same arrangement was observed in Respiratory Syncytial Virus; however, the DP-178-like region in this virus also had a leucine zipper just C-terminal to the proline. Therefore, designed N-terminal proline-leucine zipper motifs were designed to analyze whether any other viruses might contain this same pattern. The motifs are summarized in FIG. 19.

The PC/Gene protein database contains 5879 viral amino acid sequences (library file PVIRUSES; CD-ROM release 11.0). Of these, 1092 are viral envelope or glycoprotein sequences (library file PVIRUSE1). Tables V through X contain lists of protein sequence names and motif hit locations for all the motifs searched.

10. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION
OF DP-107 AND DP-178-LIKE SEQUENCES
IN HUMAN IMMUNODEFICIENCY VIRUS

FIG. 20 represents search results for HIV-1 BRU isolate gp41 (PC/Gene protein sequence PENV_HV1BR). Notice that the hybrid motif which crosses DP-107 and

DP-178 (named 107x178x4; the same motif as found in FIG. 16 found three hits including amino acids 550-599, 636-688, and 796-823. These areas include DP-107 plus eight N-terminal and four C-terminal amino acids; DP-178 plus seven N-terminal and ten C-terminal amino acids; and an area inside the transmembrane region (cytoplasmic). FIG. 20 also contains the results obtained from searching with the motif named ALLMOTI5, for which the key is found in FIG. 17 ({CDGHP}{CFP}x5). This motif also found three hits including DP-107 (amino acids 510-599), DP-178 (615-717), and a cytoplasmic region (772-841). These hits overlap the hits found by the motif 107x178x4 with considerable additional sequences on both the amino and carboxy termini. This is not surprising in that 107x178x4 is a subset of the ALLMOTI5 hybrid motif. Importantly, even though the stringency of ALLMOTI5 is considerably less than 107x178x4, it still selectively identifies the DP-107 and DP-178 regions of gp41 shown to contain sequences for inhibitory peptides of HIV-1. The results of these two motif searches are summarized in Table V under the PC/Gene protein sequence name PENV HV1BR. The proline-leucine zipper motifs also gave several hits in HIV-1 BRU including 503-525 which is at the very C-terminus of gp120, just upstream of the cleavage site (P7LZIPC and P12LZIPC); and 735-768 in the cytoplasmic domain of gp41 (P23LZIPC). These results are found in Tables VIII, IX, and X under the same sequence name as mentioned above. Notice that the only area of HIV-1 BRU which is predicted by the Lupas algorithm to contain a coiled-coil region, is from amino acids 635-670. This begins eight amino acids N-terminal to the start and ends eight amino acids N-terminal to the end of DP-178. DP-107, despite the fact that it is a known coiled coil, is

not predicted to contain a coiled-coil region using the Lupas method.

11. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION
OF DP-107-LIKE AND DP-178-LIKE
SEQUENCES IN HUMAN RESPIRATORY
SYNCYTIAL VIRUS

FIG. 21 represents search results for Human Respiratory Syncytial Virus (RSV; Strain A2) fusion glycoprotein F1 (PC/Gene protein sequence name PVGLF_HRSVA). Motif 107x178x4 finds three hits including amino acids 152-202, 213-243, and 488-515. The arrangement of these hits is similar to what is found in HIV-1 except that the motif finds two regions with similarities to DP-178, one just downstream of what would be called the DP-107 region or amino acids 213-243, and one just upstream of the transmembrane region (also similar to DP-178) or amino acids 488-515. Motif ALLMOTI5 also finds three areas including amino acids 116-202, 267-302, and 506-549. The proline-leucine zipper motifs also gave several hits including amino acids 205-221 and 265-287 (P1LZIPC 265-280, P12LZIPC), and 484-513 (P7LZIPC and P12LZIPC 484-506, P23LZIPC). Notice that the PLZIP motifs also identify regions which share location similarities with DP-178 of HIV-1.

12. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF
DP-107-LIKE AND DP-178-LIKE SEQUENCES
IN SIMIAN IMMUNODEFICIENCY VIRUS

Motif hits for Simian immunodeficiency Virus gp41 (AGM3 isolate; PC/Gene protein sequence name PENV_SIVAG) are shown in FIG. 22. Motif 107x178x4 finds three hits including amino acids 566-593, 597-624, and 703-730. The first two hits only have three amino acids between them and could probably be combined into one hit from 566-624 which would

represent a DP-107-like hit. Amino acids 703 to 730 would then represent a DP-178-like hit. ALLMOTI5 also finds three hits including amino acids 556-628 (DP-107-like), 651-699 (DP-178-like), and 808-852 which represents the transmembrane spanning region. SIV
5 also has one region from 655-692 with a high propensity to form a coiled coil as predicted by the Lupas algorithm. Both 107x178x4 and ALLMOTI5 motifs find the same region. SIV does not have any PLZIP motif hits in gp41.
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13. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF DP-107-LIKE AND DP-178 LIKE SEQUENCES IN CANINE DISTEMPER VIRUS

Canine Distemper Virus (strain Onderstepoort)
15 fusion glycoprotein F1 (PC/Gene Protein sequence name PVGLF_CDVO) has regions similar to Human RSV which are predicted to be DP-107-like and DP-178-like (FIG. 23). Motif 107x178x4 highlights one area just C-terminal to the fusion peptide at amino acids 252-293. Amino
20 acids 252-286 are also predicted to be coiled coil using the Lupas algorithm. Almost 100 amino acids C-terminal to the first region is a DP-178-like area at residues 340-367. ALLMOTI5 highlights three areas of interest including: amino acids 228-297, which
25 completely overlaps both the Lupas prediction and the DP-107-like 107x178x4 hit; residues 340-381, which overlaps the second 107x178x4 hit; and amino acids 568-602, which is DP178-like in that it is located just N-terminal to the transmembrane region. It also
30 overlaps another region (residues 570-602) predicted by the Lupas method to have a high propensity to form a coiled coil. Several PLZIP motifs successfully identified areas of interest including P6 and P12LZIPC which highlight residues 336-357 and 336-361
35 respectively; P1 and P12LZIPC which find residues 398-

414; and P12 and P23LZIPC which find residues 562-589 and 562-592 respectively.

14. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF
DP-107-LIKE AND DP-178-LIKE SEQUENCES
IN NEWCASTLE DISEASE VIRUS

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FIG. 24 shows the motif hits found in Newcastle Disease Virus (strain Australia-Victoria/32; PC Gene protein sequence name PVGLF_NDVA). Motif 107x178x4 finds two areas including a DP-107-like hit at amino acids 151-178 and a DP-178-like hit at residues 426-512. ALLMOTI5 finds three areas including residues 117-182, 231-272, and 426-512. The hits from 426-512 include a region which is predicted by the Lupas method to have a high coiled-coil propensity (460-503). The PLZIP motifs identify only one region of interest at amino acids 273-289 (P1 and 12LZIPC).

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15. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION
OF DP-107-LIKE AND DP-178-LIKE
SEQUENCES IN HUMAN PARAINFLUENZA VIRUS

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Both motifs 107x178x4 and ALLMOTI5 exhibit DP-107-like hits in the same region, 115-182 and 117-182 respectively, of Human Parainfluenza Virus (strain NIH 47885; PC/Gene protein sequence name PVGLF_p13H4; (FIG. 25). In addition, the two motifs have a DP-178-like hit just slightly C-terminal at amino acids 207-241. Both motifs also have DP-178-like hits nearer the transmembrane region including amino acids 457-497 and 462-512 respectively. Several PLZIP motif hits are also observed including 283-303 (P5LZIPC), 283-310 (P12LZIPC), 453-474 (P6LZIPC), and 453-481 (P23LZIPC). The Lupas algorithm predicts that amino acids 122-176 have a propensity to form a coiled-coil.

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16. EXAMPLE: COMPUTER-ASSISTED IDENTIFICATION OF
DP-107-LIKE AND DP-178-LIKE SEQUENCES OF
INFLUENZA A VIRUS

FIG. 26 illustrates the Lupas prediction for a coiled coil in Influenza A Virus (strain A/Aichi/2/68) at residues 379-436, as well as the motif hits for 107x178x4 at amino acids 387-453, and for ALLMOTI5 at residues 380-456. Residues 383-471 (38-125 of HA2) were shown by Carr and Kim to be an extended coiled coil when under acidic pH (Carr and Kim, 1993, Cell 73: 823-832). The Lupas algorithm predicts a coiled-coil at residues 379-436. All three methods successfully predicted the region shown to actually have coiled-coil structure; however, ALLMOTI5 predicted the greatest portion of the 88 residue stretch.

17. EXAMPLE: RSV ANTIVIRAL COMPOUNDS

In the Example presented herein, respiratory syncytial virus (RSV) peptide sequences identified by utilizing the computer-assisted coiled-coil peptide sequence searches described in Example 9, above, are shown to encode peptide domains that exhibit structural similarity to actual, known coiled-coil peptides, and are, additionally found to exhibit antiviral activity.

17.1 MATERIALS AND METHODS

Structural analyses consisted of circular dichroism (CD) studies, which were conducted according to the methods described in the Applicants' co-pending U.S. Patent Application Ser. No 08/073,028.

Anti-RSV antiviral activity was assayed as described in Pringle, C.R. et al., 1985, J. Medical Vir. 17:377-386.

35

A 48 amino acid RSV F2 peptide and a 53 amino acid RSV T67 peptide are utilized which span sequences that were identified via the computer assisted peptide sequence search strategies described in Example 9, above. See FIG. 21 for the exact position of these sequences and for the motifs utilized.

17.2 RESULTS

35-mer oligopeptides were synthesized which constituted portions of the 48 amino acid RSV F2 peptide sequence (FIG. 27) and portions of the 53 amino acid RSV T67 peptide sequence (FIG. 28). The oligopeptides were assayed, via CD analysis, for structural similarity to known coiled-coil structures, and for anti-RSV activity. As shown in FIGS. 27 and 28, a number of these oligopeptides exhibited substantial coiled-coil structural similarity and/or antiviral activity.

Thus, the computer assisted searches described, herein, in Example 9, for example, successfully identified viral peptide domains that represent highly promising anti-RSV antiviral compounds.

18. EXAMPLE: HPF3 ANTIVIRAL COMPOUNDS

In the Example presented herein, human parainfluenza virus 3 (HPF3) peptide sequences identified by utilizing the computer-assisted coiled-coil peptide sequence searches described in Example 9, above, are shown to encode peptide domains that exhibit structural similarity to actual, known coiled-coil peptides, and are, additionally found to exhibit antiviral activity.

18.1 MATERIALS AND METHODS

Structural analyses consisted of circular dichroism (CD) studies, which were conducted according

to the methods described in the Applicants' co-pending U.S. Patent Application Ser. No 08/073,028.

Anti-HPF3 antiviral activity was assayed as described in Pringle, C.R. et al., 1985, J. Medical Vir. 17:377-386.

5 A 56 amino acid and 70 amino acid HPF3 peptide are utilized which span sequences that were identified via the computer assisted peptide sequence search strategies described in Example 9, above. See FIG. 25 for the exact positions of these sequences and for the
10 motifs utilized.

18.2 RESULTS

35-mer oligopeptides were synthesized which constituted portions of the 56 amino acid HPF3 peptide
15 sequence (FIG. 29) and portions of the 70 amino acid HPF3 peptide sequence (FIG. 30). The oligopeptides were assayed, via CD analysis, for structural similarity to known coiled-coil structures, and for
20 anti-HPF3 activity. As shown in FIGS. 29 and 30, a number of these oligopeptides exhibited substantial coiled-coil structural similarity and/or antiviral activity.

Thus, the computer assisted searches described, herein, in Example 9, for example, successfully
25 identified viral peptide domains that represent highly promising anti-HPF3 antiviral compounds.

The present invention is not to be limited in scope by the specific embodiments described which are
30 intended as single illustrations of individual aspects of the invention, and functionally equivalent methods and components are within the scope of the invention. Indeed, various modifications of the invention, in addition to those shown and described herein will
35 become apparent to those skilled in the art from the

foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

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WHAT IS CLAIMED IS:

1. A peptide having an amino acid sequence corresponding to an α -helix region of an extracellular domain of a viral envelope protein, which interacts
5 with and binds to a second α -helix region of the viral envelope protein containing a leucine-zipper domain having a coiled-coil structure.
- 10 2. The peptide of Claim 1 wherein the peptide is recognized by a computer-assisted peptide sequence search utilizing an ALLMOTI5, 107x178x4 motif, or a PLZIP motif.
- 15 3. The peptide of Claim 1 in which the enveloped virus is a retrovirus.
4. The peptide of Claim 3 in which the retrovirus is a human retrovirus.
- 20 5. The peptide of Claim 4 in which the human retrovirus is HIV-1 or HIV-2.
- 25 6. The peptide of Claim 4 in which the human retrovirus is HTLV-I or HTLV-II
7. The peptide of Claim 1 in which the enveloped virus is a non-human retrovirus.
- 30 8. The peptide of Claim 6 in which the non-human retrovirus is bovine leukosis virus, feline sarcoma virus, feline leukemia virus, simian immunodeficiency virus, simian sarcoma virus, and sheep progress pneumonia virus.

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9. The peptide of Claim 1 in which the enveloped virus is a non-retroviral virus.

10. The peptide of Claim 9 in which the virus is respiratory syncytial virus, influenza virus,
 5 parainfluenza virus, canine distemper virus, or newcastle disease virus.

11. A peptide having a formula selected from the group consisting of:

10 X-YTS-Z
 X-YTSL-Z
 X-YTSLI-Z
 X-YTSLIH-Z
 X-YTSLIHS-Z
 X-YTSLIHSL-Z
 X-YTSLIHSLI-Z
 15 X-YTSLIHSLIE-Z
 X-YTSLIHSLIEE-Z
 X-YTSLIHSLIEES-Z
 X-YTSLIHSLIEESQ-Z
 X-YTSLIHSLIEESQN-Z
 X-YTSLIHSLIEESQNNQ-Z
 X-YTSLIHSLIEESQNNQE-Z
 20 X-YTSLIHSLIEESQNNQEK-Z
 X-YTSLIHSLIEESQNNQEKNE-Z
 X-YTSLIHSLIEESQNNQEKNEQ-Z
 X-YTSLIHSLIEESQNNQEKNEQE-Z
 X-YTSLIHSLIEESQNNQEKNEQEL-Z
 X-YTSLIHSLIEESQNNQEKNEQELL-Z
 25 X-YTSLIHSLIEESQNNQEKNEQELLE-Z
 X-YTSLIHSLIEESQNNQEKNEQELLEL-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELD-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELDK-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELDKW-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELDKWA-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELDKWA-S-Z
 30 X-YTSLIHSLIEESQNNQEKNEQELLELDKWA-SL-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELDKWA-SLW-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELDKWA-SLWN-Z
 X-YTSLIHSLIEESQNNQEKNEQELLELDKWA-SLWNW-Z and
 X-YTSLIHSLIEESQNNQEKNEQELLELDKWA-SLWNWF-Z (SEQ ID:1), or

35

5 X-NWF-Z
 X-WNWF-Z
 X-LWNWF-Z
 X-SLWNWF-Z
 X-ASLWNWF-Z
 X-WASLWNWF-Z
 X-KWASLWNWF-Z
 X-DKWASLWNWF-Z
 X-LDKWASLWNWF-Z
 X-ELDKWASLWNWF-Z
 X-LELDKWASLWNWF-Z
 X-LLELDKWASLWNWF-Z
 X-ELLELDKWASLWNWF-Z
 X-QELLELDKWASLWNWF-Z
 10 X-EQELLELDKWASLWNWF-Z
 X-NEQELLELDKWASLWNWF-Z
 X-KNEQELLELDKWASLWNWF-Z
 X-EKNEQELLELDKWASLWNWF-Z
 X-QEKNEQELLELDKWASLWNWF-Z
 X-QQEKNEQELLELDKWASLWNWF-Z
 X-NQQEKNEQELLELDKWASLWNWF-Z
 X-QNQQEKNEQELLELDKWASLWNWF-Z
 15 X-SQNQQEKNEQELLELDKWASLWNWF-Z
 X-ESQNQQEKNEQELLELDKWASLWNWF-Z
 X-EESQNQQEKNEQELLELDKWASLWNWF-Z
 X-IEESQNQQEKNEQELLELDKWASLWNWF-Z
 X-LIEESQNQQEKNEQELLELDKWASLWNWF-Z
 X-SLIEESQNQQEKNEQELLELDKWASLWNWF-Z
 X-HSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
 20 X-IHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
 X-LIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
 X-SLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z
 and X-TSLIHSLIEESQNQQEKNEQELLELDKWASLWNWF-Z

in which:

25 amino acid residues are presented by the single-
 letter code;
 X comprises an amino group, an acetyl group, a 9-
 fluorenylmethoxy-carbonyl group, a
 hydrophobic group, or a macromolecule
 30 carrier group;
 Z comprises a carboxyl group, an amido group, a
 hydrophobic group, or a macromolecular
 carrier group.

35 12. A peptide having a formula selected from the
 group consisting of:

X-LEA-Z
 X-LEAN-Z
 X-LEANI-Z
 X-LEANIS-Z
 X-LEANISQ-Z
 X-LEANISQS-Z
 X-LEANISQSL-Z
 5 X-LEANISQSLE-Z
 X-LEANISQSLEQ-Z
 X-LEANISQSLEQA-Z
 X-LEANISQSLEQQA-Z
 X-LEANISQSLEQAQI-Z
 X-LEANISQSLEQAQIQ-Z
 X-LEANISQSLEQAQIQQ-Z
 10 X-LEANISQSLEQAQIQQE-Z
 X-LEANISQSLEQAQIQQEK-Z
 X-LEANISQSLEQAQIQQEK-N-Z
 X-LEANISQSLEQAQIQQEKNM-Z
 X-LEANISQSLEQAQIQQEKNMY-Z
 X-LEANISQSLEQAQIQQEKNMYE-Z
 X-LEANISQSLEQAQIQQEKNMYEL-Z
 X-LEANISQSLEQAQIQQEKNMYELQ-Z
 15 X-LEANISQSLEQAQIQQEKNMYELQK-Z
 X-LEANISQSLEQAQIQQEKNMYELQKL-Z
 X-LEANISQSLEQAQIQQEKNMYELQKLN-Z
 X-LEANISQSLEQAQIQQEKNMYELQKLNS-Z
 X-LEANISQSLEQAQIQQEKNMYELQKLNSW-Z
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWD-Z
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDV-Z
 20 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFT-Z
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTN-Z
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNW-Z and
 X-LEANISQSLEQAQIQQEKNMYELQKLNSWDVFTNWL-Z (SEQ ID:7), or

X-NWL-Z
 X-TNWL-Z
 25 X-FTNWL-Z
 X-VFTNWL-Z
 X-DVFTNWL-Z
 X-WDVFTNWL-Z
 X-SWDVFTNWL-Z
 X-NSWDVFTNWL-Z
 X-LNSWDVFTNWL-Z
 30 X-KLNSWDVFTNWL-Z
 X-QKLNSWDVFTNWL-Z
 X-LQKLNSWDVFTNWL-Z
 X-ELQKLNSWDVFTNWL-Z
 X-YELQKLNSWDVFTNWL-Z
 X-MYELQKLNSWDVFTNWL-Z
 X-NMYELQKLNSWDVFTNWL-Z
 X-KNMYELQKLNSWDVFTNWL-Z
 35 X-EKNMYELQKLNSWDVFTNWL-Z
 X-QEKNMYELQKLNSWDVFTNWL-Z

X-QQEKMYELQKLNSWDVFTNWL-Z
 X-IQQEKMYELQKLNSWDVFTNWL-Z
 X-QIQQEKMYELQKLNSWDVFTNWL-Z
 X-AQIQQEKMYELQKLNSWDVFTNWL-Z
 X-QAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-EQAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-LEQAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-SLEQAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-QKSLEQAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-SQSLEQAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-ISQSLEQAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-NISQSLEQAQIQQEKMYELQKLNSWDVFTNWL-Z
 X-ANISQSLEQAQIQQEKMYELQKLNSWDVFTNWL-Z
 and X-EANISQSLEQAQIQQEKMYELQKLNSWDVFTNWL-Z

10 in which:

amino acid residues are presented by the single-letter code;

15 X comprises an amino group, an acetyl group, a 9-fluoromethoxymethyl-carbonyl group, a hydrophobic group, or a macromolecule carrier group;

20 Z comprises a carboxyl group, an amido group, a hydrophobic group, or a macromolecular carrier group.

13. A peptide having a formula selected from the group consisting of:

X-YTS-Z
 X-YTSV-Z
 25 X-YTSVI-Z
 X-YTSVIT-Z
 X-YTSVITI-Z
 X-YTSVITIE-Z
 X-YTSVITIEL-Z
 X-YTSVITIELS-Z
 X-YTSVITIELSN-Z
 30 X-YTSVITIELSNI-Z
 X-YTSVITIELSNIK-Z
 X-YTSVITIELSNIKE-Z
 X-YTSVITIELSNIKEN-Z
 X-YTSVITIELSNIKENK-Z
 X-YTSVITIELSNIKENKC-Z
 X-YTSVITIELSNIKENKCN-Z
 X-YTSVITIELSNIKENKCNCG-Z
 35 X-YTSVITIELSNIKENKCNGT-Z
 X-YTSVITIELSNIKENKCNGTD-Z

X-YTSVITIELSNIKENKCNGTDA-Z
 X-YTSVITIELSNIKENKCNGTDAK-Z
 X-YTSVITIELSNIKENKCNGTDAKV-Z
 X-YTSVITIELSNIKENKCNGTDAKVK-Z
 X-YTSVITIELSNIKENKCNGTDAKVKL-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLI-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIK-Z
 5 X-YTSVITIELSNIKENKCNGTDAKVKLIQ-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQE-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQEL-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELD-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDK-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKY-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYK-Z
 10 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKN-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNA-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAV-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTE-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTEL-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELO-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELOL-Z
 15 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELOLL-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELOLLM-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELOLLMQ-Z
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELOLLMQS-Z and
 X-YTSVITIELSNIKENKCNGTDAKVKLIQELDKYKNAVTELOLLMQST-Z, or

X-QST-Z
 X-MQST-Z
 20 X-LMQST-Z
 X-LLMQST-Z
 X-QLLMQST-Z
 X-LQLLMQST-Z
 X-ELQLLMQST-Z
 X-TELQLLMQST-Z
 X-VTELQLLMQST-Z
 25 X-AVTELQLLMQST-Z
 X-NAVTELQLLMQST-Z
 X-KNAVTELQLLMQST-Z
 X-YKNAVTELQLLMQST-Z
 X-KYKNAVTELQLLMQST-Z
 X-DKYKNAVTELQLLMQST-Z
 X-LDKYKNAVTELQLLMQST-Z
 30 X-ELDKYKNAVTELQLLMQST-Z
 X-QELDKYKNAVTELQLLMQST-Z
 X-KQELDKYKNAVTELQLLMQST-Z
 X-IKQELDKYKNAVTELQLLMQST-Z
 X-LIKQELDKYKNAVTELQLLMQST-Z
 X-KLIKQELDKYKNAVTELQLLMQST-Z
 X-VKLIKQELDKYKNAVTELQLLMQST-Z
 X-KVKLIKQELDKYKNAVTELQLLMQST-Z
 35 X-AKVLIKQELDKYKNAVTELQLLMQST-Z
 X-DAKVLIKQELDKYKNAVTELQLLMQST-Z

X-TDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-GTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-NGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-CNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-KCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-NKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-ENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 5 X-KENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-IKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-NIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-SNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-LSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-ELSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-IELSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 10 X-TIELSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-ITIELSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-VITIELSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-SVITIELSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z
 X-TSVITIELSNIKENKCNGTDAKVLIKQELDKYKNAVTELQLLMQST-Z

in which:

15 amino acid residues are presented by the single-
 letter code;
 X comprises an amino group, an acetyl group, a 9-
 fluoromethyoxymethyl-carbonyl group, a
 hydrophobic group, or a macromolecule
 carrier group;
 20 Z comprises a carboxyl group, an amido group, a
 hydrophobic group, or a macromolecular
 carrier group.

25 14. A peptide having a formula selected from the
 group consisting of:

X-FYD-Z
 X-FYDP-Z
 X-FYDPL-Z
 X-FYDPLV-Z
 X-FYDPLVF-Z
 30 X-FYDPLVFP-Z
 X-FYDPLVFPS-Z
 X-FYDPLVFPSD-Z
 X-FYDPLVFPSDE-Z
 X-FYDPLVFPSDEF-Z
 X-FYDPLVFPSDEFD-Z
 X-FYDPLVFPSDEFDA-Z
 35 X-FYDPLVFPSDEFDAS-Z
 X-FYDPLVFPSDEFDASI-Z

X-FYDPLVFPSEFDASIS-Z
 X-FYDPLVFPSEFDASISQ-Z
 X-FYDPLVFPSEFDASISQV-Z
 X-FYDPLVFPSEFDASISQVN-Z
 X-FYDPLVFPSEFDASISQVNE-Z
 X-FYDPLVFPSEFDASISQVNEK-Z
 X-FYDPLVFPSEFDASISQVNEKI-Z
 5 X-FYDPLVFPSEFDASISQVNEKIN-Z
 X-FYDPLVFPSEFDASISQVNEKINQ-Z
 X-FYDPLVFPSEFDASISQVNEKINQS-Z
 X-FYDPLVFPSEFDASISQVNEKINQSL-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLA-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAF-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAFI-Z
 10 X-FYDPLVFPSEFDASISQVNEKINQSLAFIR-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRK-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKS-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSD-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSDE-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSDEL-Z
 X-FYDPLVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z, or

 15 X-DELL-Z
 X-SDELL-Z
 X-KSDELL-Z
 X-RKSDELL-Z
 X-IRKSDELL-Z
 X-FIRKSDELL-Z
 X-AFIRKSDELL-Z
 20 X-LAFIRKSDELL-Z
 X-SLAFIRKSDELL-Z
 X-QSLAFIRKSDELL-Z
 X-NQSLAFIRKSDELL-Z
 X-INQSLAFIRKSDELL-Z
 X-KINQSLAFIRKSDELL-Z
 X-EKINQSLAFIRKSDELL-Z
 X-NEKINQSLAFIRKSDELL-Z
 25 X-VNEKINQSLAFIRKSDELL-Z
 X-QVNEKINQSLAFIRKSDELL-Z
 X-SQVNEKINQSLAFIRKSDELL-Z
 X-ISQVNEKINQSLAFIRKSDELL-Z
 X-SISQVNEKINQSLAFIRKSDELL-Z
 X-ASISQVNEKINQSLAFIRKSDELL-Z
 X-DASISQVNEKINQSLAFIRKSDELL-Z
 30 X-FDASISQVNEKINQSLAFIRKSDELL-Z
 X-EFDASISQVNEKINQSLAFIRKSDELL-Z
 X-DEFDASISQVNEKINQSLAFIRKSDELL-Z
 X-SDEFDASISQVNEKINQSLAFIRKSDELL-Z
 X-PSDEFDASISQVNEKINQSLAFIRKSDELL-Z
 X-FPSDEFDASISQVNEKINQSLAFIRKSDELL-Z
 X-VFPSDEFDASISQVNEKINQSLAFIRKSDELL-Z
 X-LVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z
 35 X-PLVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z
 X-DPLVFPSEFDASISQVNEKINQSLAFIRKSDELL-Z

X-YDPLVFPSEDFDASISQVNEKINQSLAFIRKSDELL-Z

in which:

amino acid residues are presented by the single-letter code;

5

X comprises an amino group, an acetyl group, a 9-fluoromethoxymethyl-carbonyl group, a hydrophobic group, or a macromolecule carrier group;

10

Z comprises a carboxyl group, an amido group, a hydrophobic group, or a macromolecular carrier group.

15. A peptide having a formula selected from the group consisting of:

15

X-ITL-Z

X-ITLN-Z

X-ITLNN-Z

X-ITLNNS-Z

X-ITLNNSV-Z

X-ITLNNSVA-Z

20

X-ITLNNSVAL-Z

X-ITLNNSVALD-Z

X-ITLNNSVALDP-Z

X-ITLNNSVALDPI-Z

X-ITLNNSVALDPID-Z

X-ITLNNSVALDPIDI-Z

X-ITLNNSVALDPIDIS-Z

X-ITLNNSVALDPIDISI-Z

25

X-ITLNNSVALDPIDISIE-Z

X-ITLNNSVALDPIDISIEL-Z

X-ITLNNSVALDPIDISIELN-Z

X-ITLNNSVALDPIDISIELNK-Z

X-ITLNNSVALDPIDISIELNKA-Z

X-ITLNNSVALDPIDISIELNKAK-Z

X-ITLNNSVALDPIDISIELNKA-KS-Z

30

X-ITLNNSVALDPIDISIELNKA-KSD-Z

X-ITLNNSVALDPIDISIELNKA-KSDL-Z

X-ITLNNSVALDPIDISIELNKA-KSDLE-Z

X-ITLNNSVALDPIDISIELNKA-KSDLEE-Z

X-ITLNNSVALDPIDISIELNKA-KSDLEES-Z

X-ITLNNSVALDPIDISIELNKA-KSDLEESK-Z

X-ITLNNSVALDPIDISIELNKA-KSDLEESKE-Z

X-ITLNNSVALDPIDISIELNKA-KSDLEESKEW-Z

35

X-ITLNNSVALDPIDISIELNKA-KSDLEESKEWIR-Z

X-ITLNNSVALDPIDISIELNKA-KSDLEESKEWIR-Z

X-ITLNNVALDPIDISIELNKA KSDLEESKEWIRR-Z
 X-ITLNNVALDPIDISIELNKA KSDLEESKEWIRRS-Z, or

5 X-RRS-Z
 X-IRRS-Z
 X-WIRRS-Z
 X-EWIRRS-Z
 X-KEWIRRS-Z
 X-SKEWIRRS-Z
 X-ESKEWIRRS-Z
 X-EESKEWIRRS-Z
 X-LEESKEWIRRS-Z
 X-DLEESKEWIRRS-Z
 X-SDLEESKEWIRRS-Z
 10 X-KSDLEESKEWIRRS-Z
 X-AKSDLEESKEWIRRS-Z
 X-KAKSDLEESKEWIRRS-Z
 X-NKAKSDLEESKEWIRRS-Z
 X-LNKA KSDLEESKEWIRRS-Z
 X-ELNKA KSDLEESKEWIRRS-Z
 X-IELNKA KSDLEESKEWIRRS-Z
 X-SIELNKA KSDLEESKEWIRRS-Z
 15 X-ISIELNKA KSDLEESKEWIRRS-Z
 X-DISIELNKA KSDLEESKEWIRRS-Z
 X-IDISIELNKA KSDLEESKEWIRRS-Z
 X-PIDISIELNKA KSDLEESKEWIRRS-Z
 X-DPIDISIELNKA KSDLEESKEWIRRS-Z
 X-LDPIDISIELNKA KSDLEESKEWIRRS-Z
 X-ALDPIDISIELNKA KSDLEESKEWIRRS-Z
 20 X-VALDPIDISIELNKA KSDLEESKEWIRRS-Z
 X-SVALDPIDISIELNKA KSDLEESKEWIRRS-Z
 X-NSVALDPIDISIELNKA KSDLEESKEWIRRS-Z
 X-NNSVALDPIDISIELNKA KSDLEESKEWIRRS-Z
 X-LNNSVALDPIDISIELNKA KSDLEESKEWIRRS-Z
 X-TLNNVALDPIDISIELNKA KSDLEESKEWIRRS-Z

in which:

25 amino acid residues are presented by the single-letter code;
 X comprises an amino group, an acetyl group, a 9-fluoromethoxymethyl-carbonyl group, a hydrophobic group, or a macromolecule
 30 carrier group;
 Z comprises a carboxyl group, an amido group, a hydrophobic group, or a macromolecular carrier group.

35

16. A peptide having a formula selected from the group consisting of:

- X-ALG-Z
 X-ALGV-Z
 X-ALGVA-Z
 X-ALGVAT-Z
 5 X-ALGVATS-Z
 X-ALGVATSA-Z
 X-ALGVATSAQ-Z
 X-ALGVATSAQI-Z
 X-ALGVATSAQIT-Z
 X-ALGVATSAQITA-Z
 X-ALGVATSAQITAA-Z
 10 X-ALGVATSAQITA-AV-Z
 X-ALGVATSAQITA-AVA-Z
 X-ALGVATSAQITA-AVAL-Z
 X-ALGVATSAQITA-AVALV-Z
 X-ALGVATSAQITA-AVALVE-Z
 X-ALGVATSAQITA-AVALVEA-Z
 X-ALGVATSAQITA-AVALVEAK-Z
 X-ALGVATSAQITA-AVALVEAKQ-Z
 15 X-ALGVATSAQITA-AVALVEAKQA-Z
 X-ALGVATSAQITA-AVALVEAKQAR-Z
 X-ALGVATSAQITA-AVALVEAKQARS-Z
 X-ALGVATSAQITA-AVALVEAKQARSD-Z
 X-ALGVATSAQITA-AVALVEAKQARSDI-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIE-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIEK-Z
 20 X-ALGVATSAQITA-AVALVEAKQARSDIEKL-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLK-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKE-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEA-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEAI-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEAIR-Z
 X-ALGVATSAQITA-AVALVEAKQARSDIEKLKEAIRD-Z, or

 25 X-IRD-Z
 X-AIRD-Z
 X-EAIRD-Z
 X-KEAIRD-Z
 X-LKEAIRD-Z
 X-KLKEAIRD-Z
 X-EKLKEAIRD-Z
 X-IEKLKEAIRD-Z
 30 X-DIEKLKEAIRD-Z
 X-SDIEKLKEAIRD-Z
 X-RSDIEKLKEAIRD-Z
 X-ARSDIEKLKEAIRD-Z
 X-QARSDIEKLKEAIRD-Z
 X-KQARSDIEKLKEAIRD-Z
 X-AKQARSDIEKLKEAIRD-Z
 35 X-EAKQARSDIEKLKEAIRD-Z
 X-VEAKQARSDIEKLKEAIRD-Z

X-LVEAKQARSDIEKLKEAIRD-Z
 X-ALVEAKQARSDIEKLKEAIRD-Z
 X-VALVEAKQARSDIEKLKEAIRD-Z
 X-AVALVEAKQARSDIEKLKEAIRD-Z
 X-AAVALVEAKQARSDIEKLKEAIRD-Z
 X-TAAVALVEAKQARSDIEKLKEAIRD-Z
 X-ITAAVALVEAKQARSDIEKLKEAIRD-Z
 5 X-QITAVALVEAKQARSDIEKLKEAIRD-Z
 X-AQITAVALVEAKQARSDIEKLKEAIRD-Z
 X-SAQITAVALVEAKQARSDIEKLKEAIRD-Z
 X-TSAQITAVALVEAKQARSDIEKLKEAIRD-Z
 X-ATSAQITAVALVEAKQARSDIEKLKEAIRD-Z
 X-VATSAQITAVALVEAKQARSDIEKLKEAIRD-Z
 X-GVATSAQITAVALVEAKQARSDIEKLKEAIRD-Z
 10 X-LGVATSAQITAVALVEAKQARSDIEKLKEAIRD-Z

in which:

amino acid residues are presented by the single-letter code;

X comprises an amino group, an acetyl group, a 9-fluoromethoxymethyl-carbonyl group, a hydrophobic group, or a macromolecule carrier group;

Z comprises a carboxyl group, an amido group, a hydrophobic group, or a macromolecular carrier group.

17. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein X is a hydrophobic group.

25 18. The peptide of Claim 17 wherein the hydrophobic group X is carbobenzoxyl, dansyl, or t-butyloxycarbonyl.

19. The peptide of Claim 11, 12, 13, 14, 15 or 30 16 wherein Z is a hydrophobic group.

20. The peptide of Claim 19 wherein the hydrophobic group Z is t-butyloxycarbonyl.

35

21. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein X is a macromolecular carrier group.

22. The peptide of Claim 21 wherein the macromolecular carrier group is a lipid-fatty acid conjugate, a polyethylene glycol, or a carbohydrate moiety.

23. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein Z is a macromolecular carrier group.

24. The peptide of Claim 23 wherein the macromolecular carrier group Z is a lipid-fatty acid conjugate, a polyethylene glycol, or a carbohydrate moiety.

25. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein at least one bond linking adjacent amino acid residues is a non-peptide bond.

26. The peptide of Claim 25 wherein the non-peptide bond is an inino, ester, hydrazine, semicarbazide, or azo bond.

27. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein at least one amino acid residue is in a D-isomer configuration.

28. The peptide of Claim 11, 12, 13, 14, 15 or 16 further comprising at least one amino acid insertion.

29. The peptide of Claim 11, 12, 13, 14, 15 or 16 wherein the amino acid insertion is between 1 and 15 amino acid residues.

30. The peptide of Claim 11, 12, 13, 14, 15 or 16 having at least one less amino acid residue, wherein the amino acid residue(s) represents an amino acid deletion, and wherein the peptide comprises at least three amino acid residues.

5

31. The peptide of Claim 11, 12, 13, 14, 15 or 16 further comprising at least one amino acid substitution wherein a first amino acid residue is substituted for a second, different amino acid residue.

10

32. The peptide of Claim 31 wherein the amino acid substitution is a conserved substitution.

15

33. The peptide of Claim 31 wherein the amino acid substitution is a non-conserved substitution.

34. A method for the inhibition of transmission of an enveloped virus to a cell, comprising contacting the cell with an effective concentration of the peptide of Claim 1 for an effective period of time so that no infection of the cell by the virus occurs.

20

35. A method for neutralizing an enveloped virus in a host, comprising administering to the host an effective concentration of the peptide of Claim 1 so that the host raises an immune response sufficient to neutralize the virus, and viral infection of uninfected cells in the host is inhibited.

25

30

36. A method for neutralizing an enveloped virus in a host, comprising administering to the host an effective concentration of an antibody raised against the peptide of Claim 1 so that viral infection of uninfected cells in the host is inhibited.

35

37. A method for the detection of an enveloped virus comprising:

contacting a viral isolate with an effective concentration of the peptide of Claim 1 for an effective amount of time so that viral infectivity is inhibited; and

assaying the viral isolate for viral enzyme activity.

38. A method for the inhibition of transmission of an HIV retrovirus to a cell, comprising contacting the cell with an effective concentration of the peptide of Claim 11 or 12 for an effective period of time so that no infection of the cell by the retrovirus occurs.

39. A method for neutralizing an HIV retrovirus in a host, comprising administering to the host an effective concentration of the peptide of Claim 11 or 12 so that the host raises an immune response sufficient to neutralize the HIV retrovirus, and HIV infection of uninfected cells in the host is inhibited.

40. A method for neutralizing an HIV retrovirus in a host, comprising administering to the host an effective concentration of an antibody raised against the peptide of Claim 11 or 12 so that HIV infection of uninfected cells in the host is inhibited.

41. A method for the detection of HIV, comprising:

contacting a viral isolate with an effective concentration of the peptide of Claim 11 or 12 for an effective amount of time so that HIV viral infectivity is inhibited; and

assaying the viral isolate for retroviral enzyme activity.

42. A method for the inhibition of transmission of a respiratory syncytial virus to a cell, comprising
5 contacting the cell with an effective concentration of the peptide of Claim 13 or 14 for an effective period of time so that no infection of the cell by the virus occurs.

10 43. A method for neutralizing a respiratory syncytial virus in a host, comprising administering to the host an effective concentration of the peptide of Claim 13 or 14 so that the host raises an immune
15 response sufficient to neutralize the virus, and respiratory syncytial virus infection of uninfected cells in the host is inhibited.

20 44. A method for neutralizing a respiratory syncytial virus in a host comprising administering to the host an effective concentration of an antibody raised against the peptide of Claim 13 or 14 so that
respiratory syncytial virus infection of uninfected cells in the host is inhibited.

25 45. A method for the detection of respiratory syncytial virus comprising:
contacting a viral isolate with an effective concentration of the peptide of Claim 13 or 14 for an
effective amount of time so that respiratory syncytial
30 viral infectivity is inhibited; and
assaying the viral isolate for respiratory syncytial virus enzyme activity.

35 46. A method for the inhibition of transmission of a parainfluenza virus to a cell comprising,

contacting the cell with an effective concentration of the peptide of Claim 15 or 16 for an effective period of time so that no infection of the cell by the virus occurs.

5 47. A method for neutralizing a parainfluenza virus in a host, comprising administering to the host an effective concentration of the peptide of Claim 15 or 16 so that the host raises an immune response
10 sufficient to neutralize the virus, and parainfluenza infection of uninfected cells in the host is inhibited.

 48. A method for neutralizing a parainfluenza virus in a host comprising administering to the host
15 an effective concentration of an antibody raised against the peptide of Claim 15 or 16 so that parainfluenza infection of uninfected cells in the host is inhibited.

20 49. A method for the detection of parainfluenza virus comprising:

 contacting a viral isolate with an effective concentration of the peptide of Claim 15 or 16 for an effective amount of time so that parainfluenza viral
25 infectivity is inhibited; and

 assaying the viral isolate for parainfluenza virus enzyme activity.

30

35

HIV1LAI (DP-178; SEQ ID:1)	YTSLIHSLIEESQNQEKNEQELLELDKWASLWNWF
HIV1SF2 (DP-185; SEQ ID:3)	YTNTIYNLLEESQNQEKNEQELLELDKWASLWNWF
HIV1RF (SEQ ID:4)	YTGIIYNLLEESQNQEKNEQELLELDKWANLWNWF
HIV1MN (SEQ ID:5)	YTSLIYSLLEKSQTQEKNEQELLELDKWASLWNWF
HIV2ROD (SEQ ID:6)	LEANISKSLEQAQIQEKNMYELQKLSWDIFGNWF
HIV2NIHZ (SEQ ID:7)	LEANISQSLEQAQIQEKNMYELQKLSWDVFTNWL
DP180 (SEQ ID:2)	SSSFSTLLEQNNWKLQAEQMLEQINEKHYLEDIS
DP118 (SEQ ID:10)	QQLLDVWKRQQEMRLTWGTKNLQARVTAIEKYLKDQ
DP125 (SEQ ID:8)	CCGNNLLRAIEAQQHLLQLTWG IKQLQARILAVERYLKDQ
DP116 (SEQ ID:9)	LQARILAVERYLKDQQQ

FIG.1

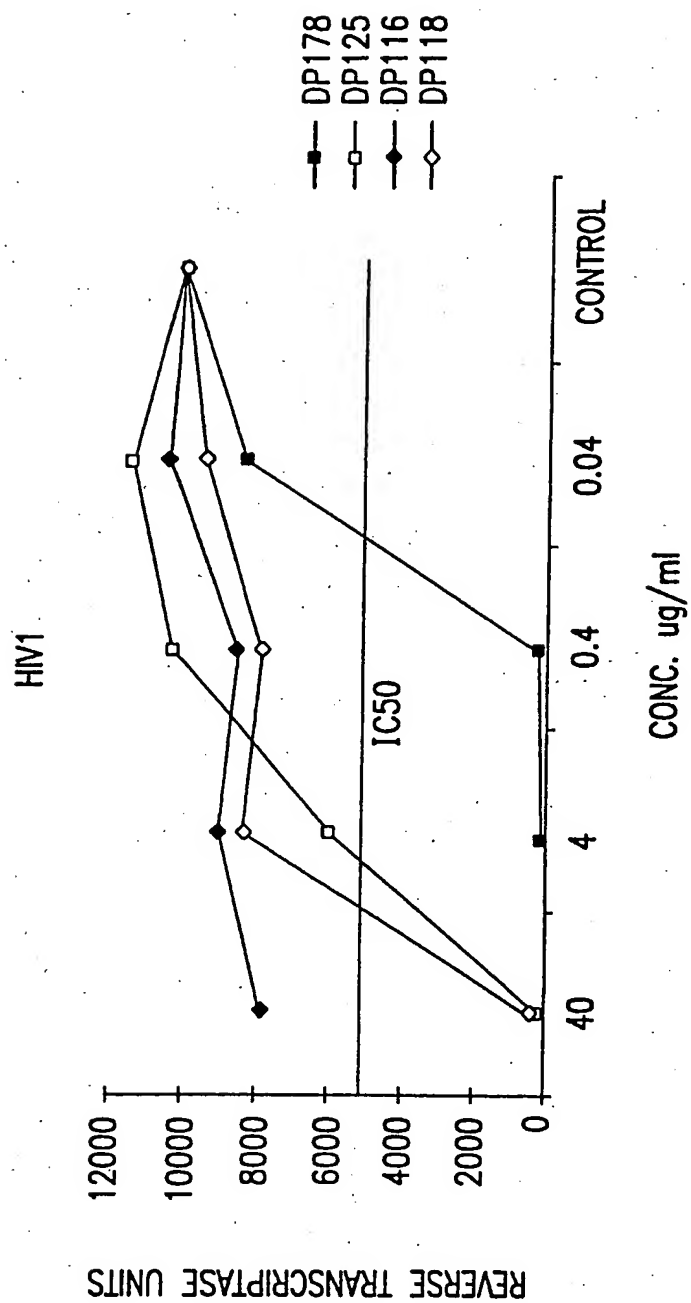


FIG.2

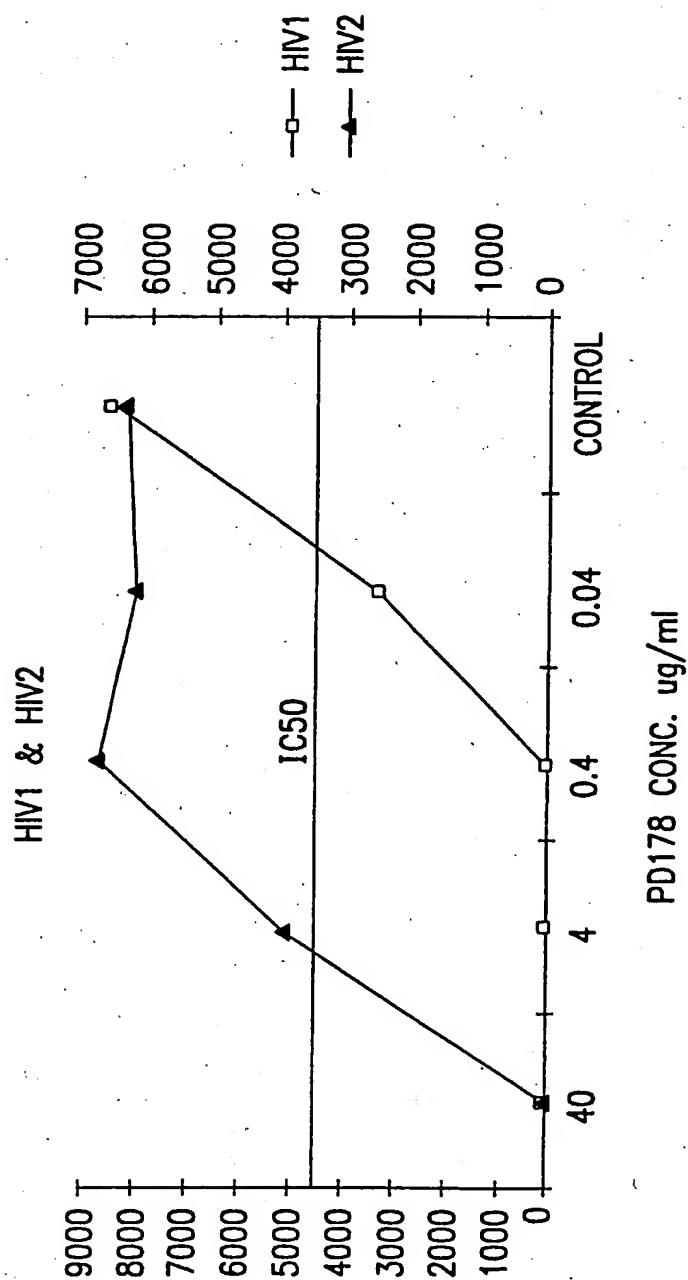


FIG.3

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Number of Syncytia/well: concentration in $\mu\text{g/ml}$ (micrograms/ml)									
DP178	10	5	1	0.2	0.1	0.05	0.025	0.0125	Control
<i>Syncytia</i>									
HIV1LAT	0	0	0	0	0	0	0	0	67
HIV1MN	0	0	0	0	0	ND	ND	ND	34
HIV1RF	0	0	0	0	0	ND	ND	ND	65
HIV1SF2	0	0	0	0	0	ND	ND	ND	58
DP125	10	5	1	0.2	0.1	0.05	0.025	0.0125	Control
<i>Syncytia</i>									
HIV1LAT	0	0	54	69	80	75	79	82	67
HIV1MN	0	0	30	36	ND	ND	ND	ND	34
HIV1RF	0	0	67	63	ND	ND	ND	ND	65
HIV1SF2	0	0	9	66	ND	ND	ND	ND	58
DP116	10	5	1	0.2	0.1	0.05	0.025	0.0125	Control
<i>Syncytia</i>									
HIV1LAT	75	ND	ND	ND	ND	ND	ND	ND	67
HIV1MN	35	ND	ND	ND	ND	ND	ND	ND	34
HIV1RF	81	ND	ND	ND	ND	ND	ND	ND	65
HIV1SF2	81	ND	ND	ND	ND	ND	ND	ND	58

FIG.4A

DP180	40	20	10	5	2.5	1.25	0.625	0.3125	Control
<i>Syncytia</i>									
HIV1LAT	50	>45	>45	>45	>45	>45	>45	>45	58
DP185	40	20	10	5	2.5	1.25	0.625	0.3125	Control
<i>Syncytia</i>									
HIV1LAT	0	0	0	0	0	0	0	ND	60

FIG.4B

<u>HIV1</u>								
Number of Syncytia/well: concentration in ng/ml (nanograms/ml)								
DP178	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV1	0	0	0	0	0	14	20	48
DP116	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV1	ND	48	ND	ND	ND	ND	ND	ND
<u>HIV2</u>								
Number of Syncytia/well: concentration in μ g/ml (micrograms/ml)								
DP178	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV2	50	54	55	57	63	77	78	76
DP116	20	10	5	2.5	1.25	0.625	0.3125	Control
<u>Syncytia</u>								
HIV2	ND	58	ND	ND	ND	ND	ND	ND

FIG.5

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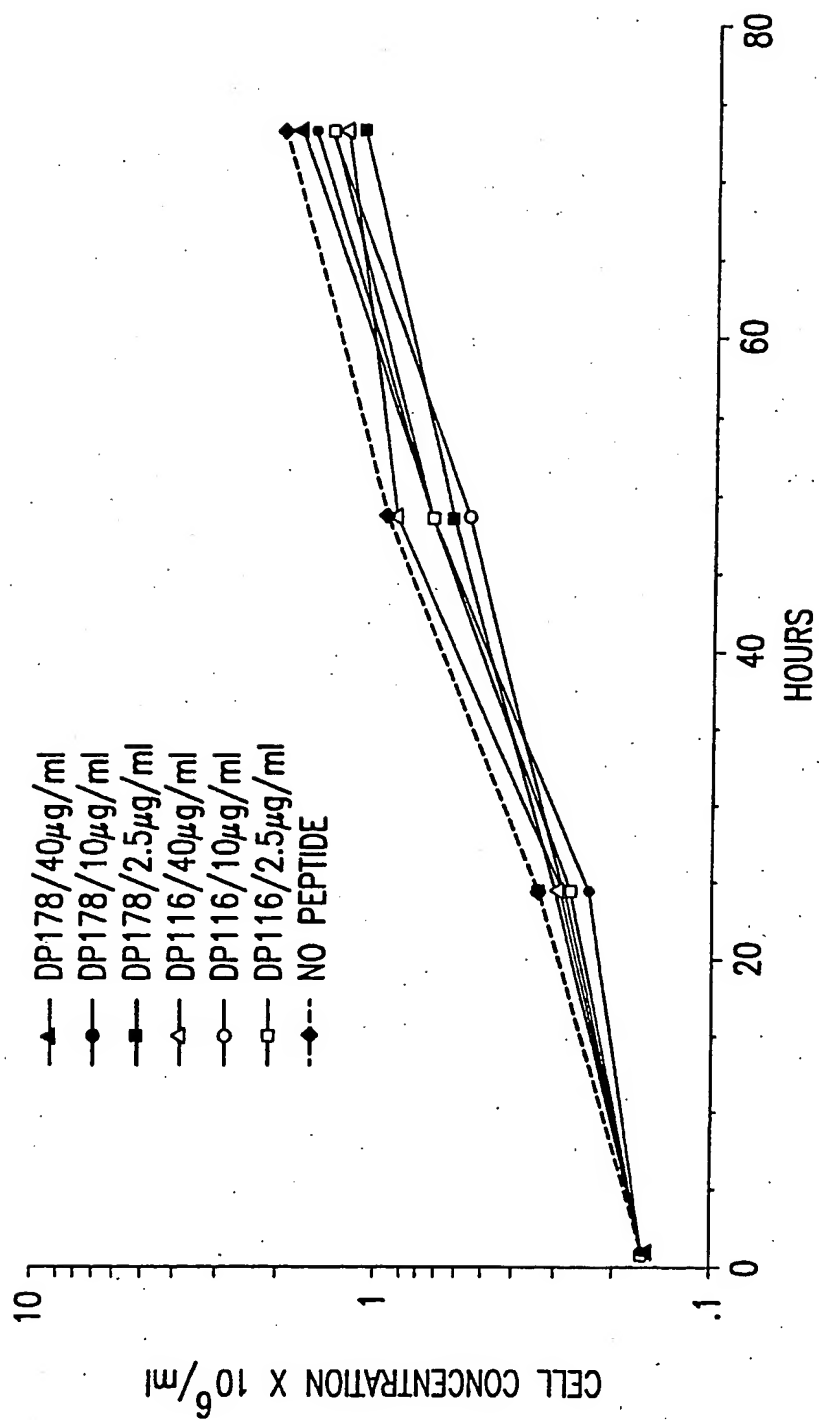


FIG. 6

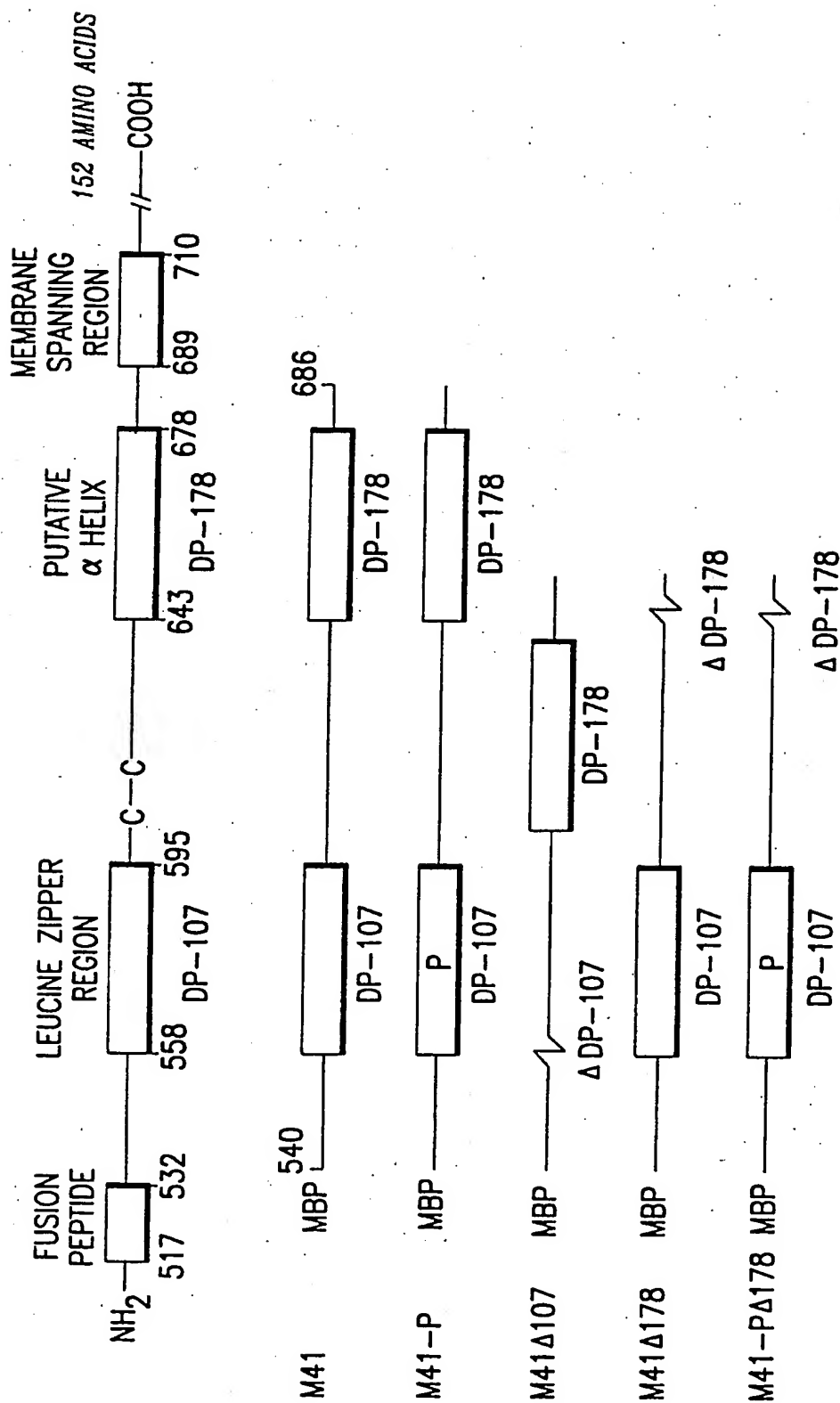


FIG.7

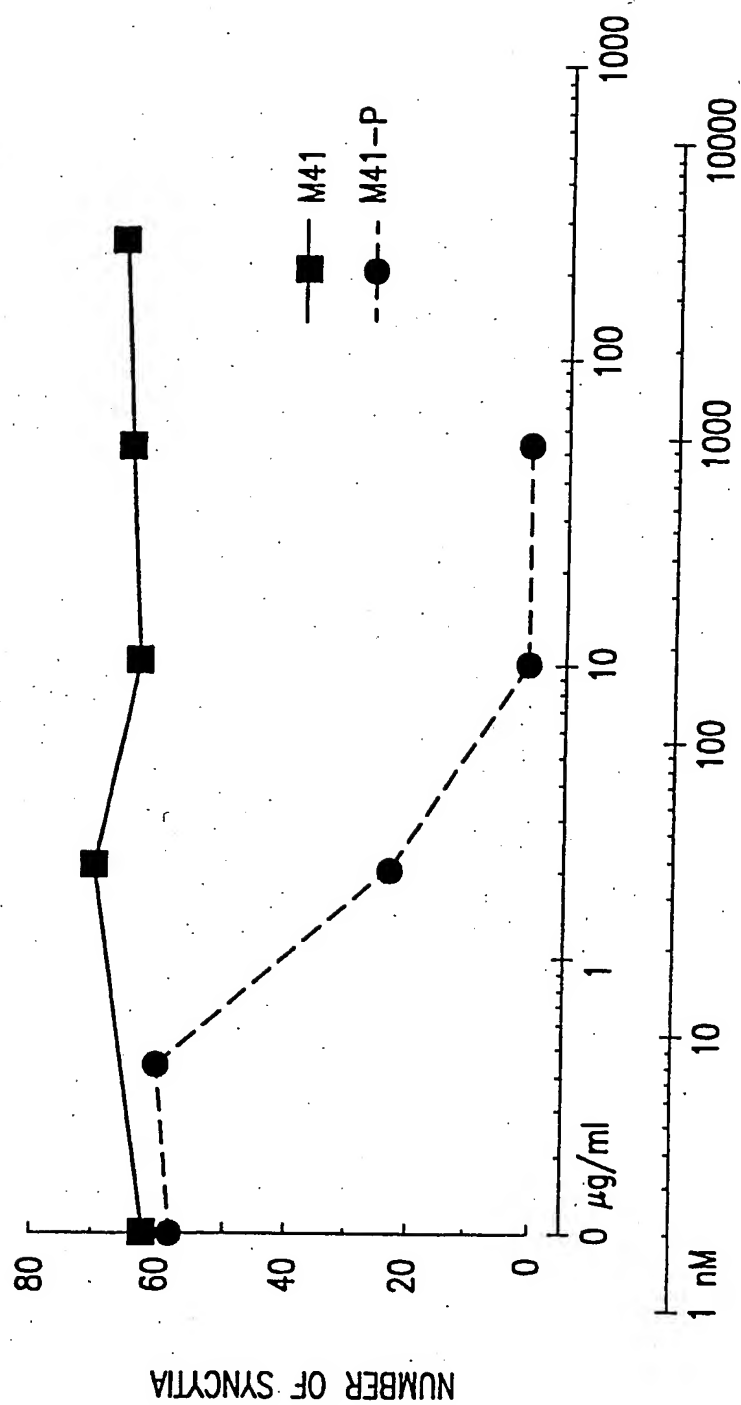


FIG.8

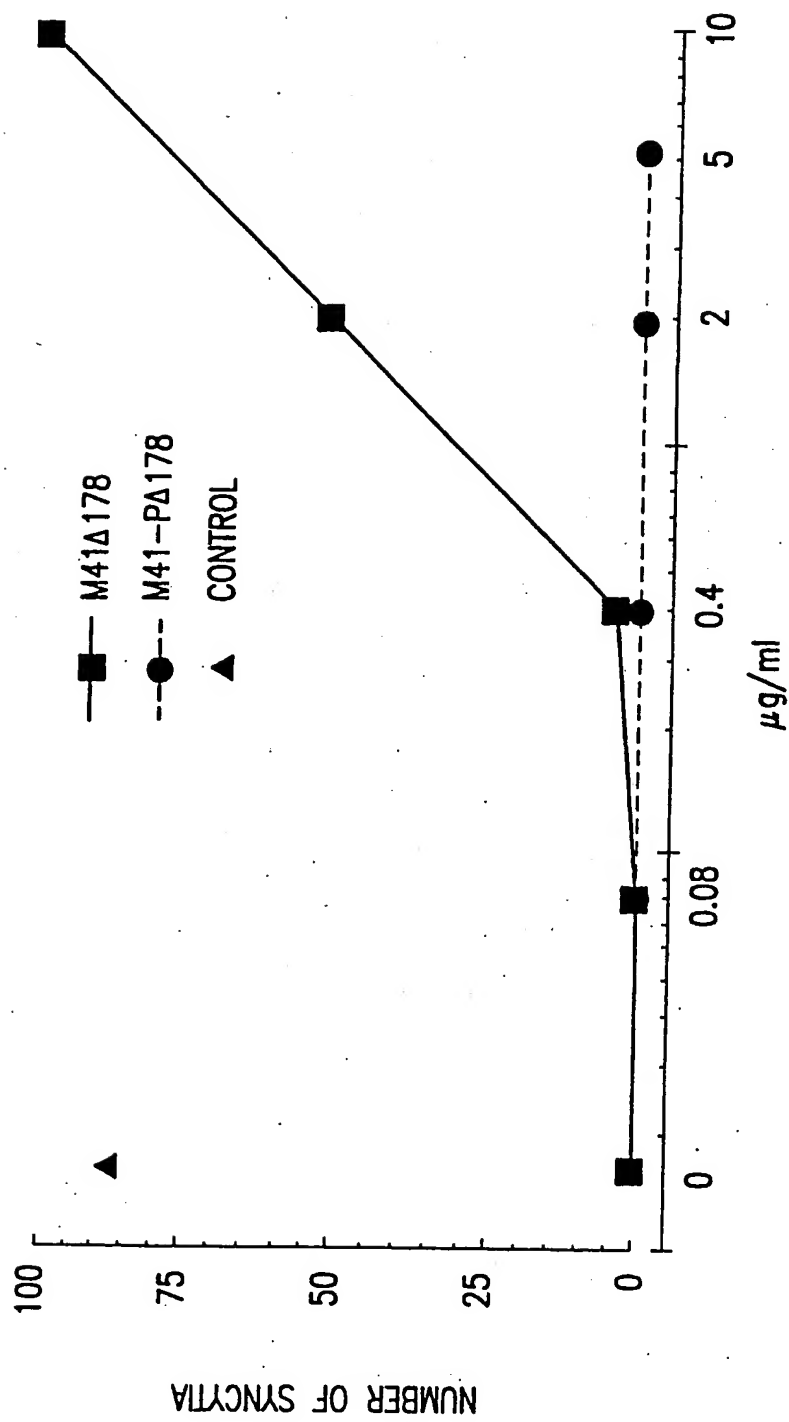


FIG.9

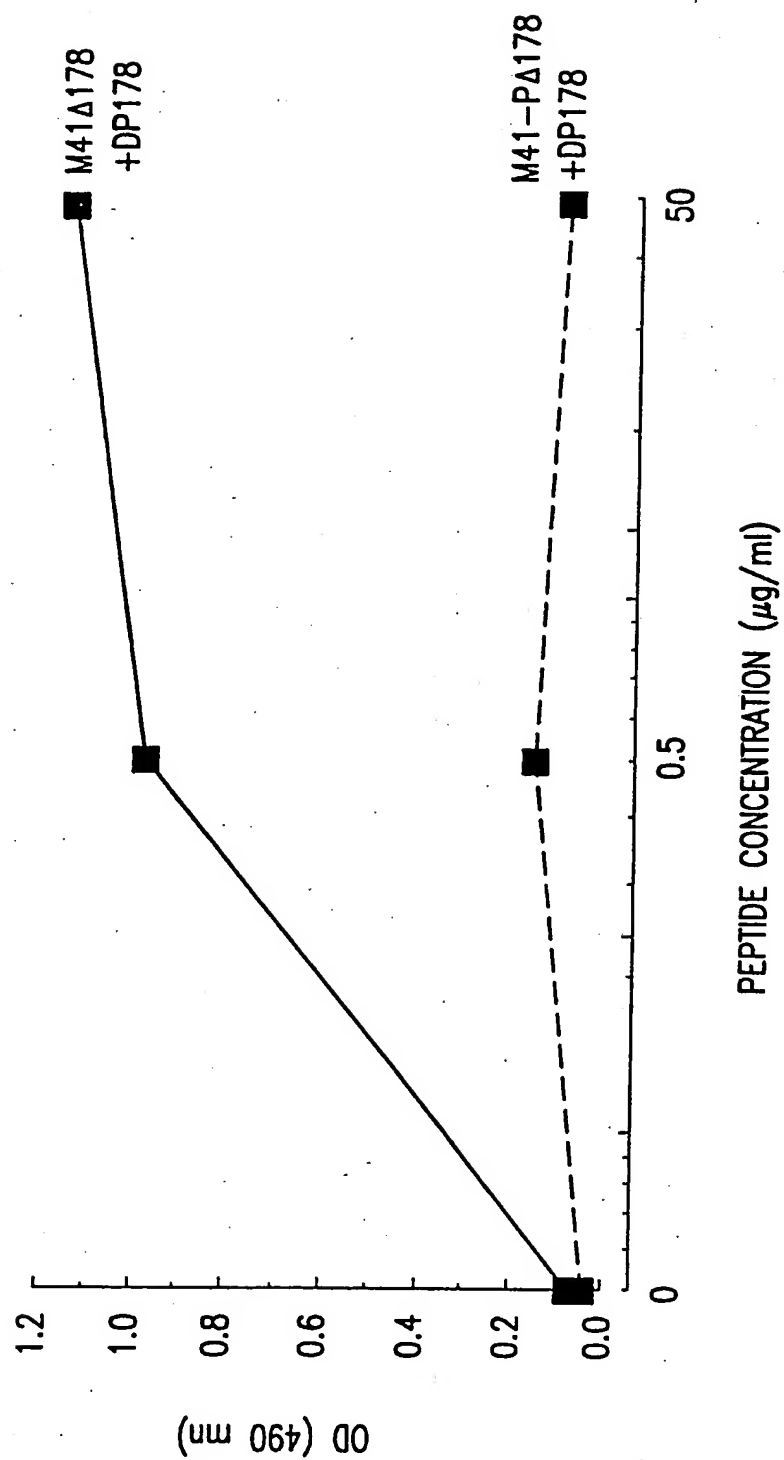


FIG.10

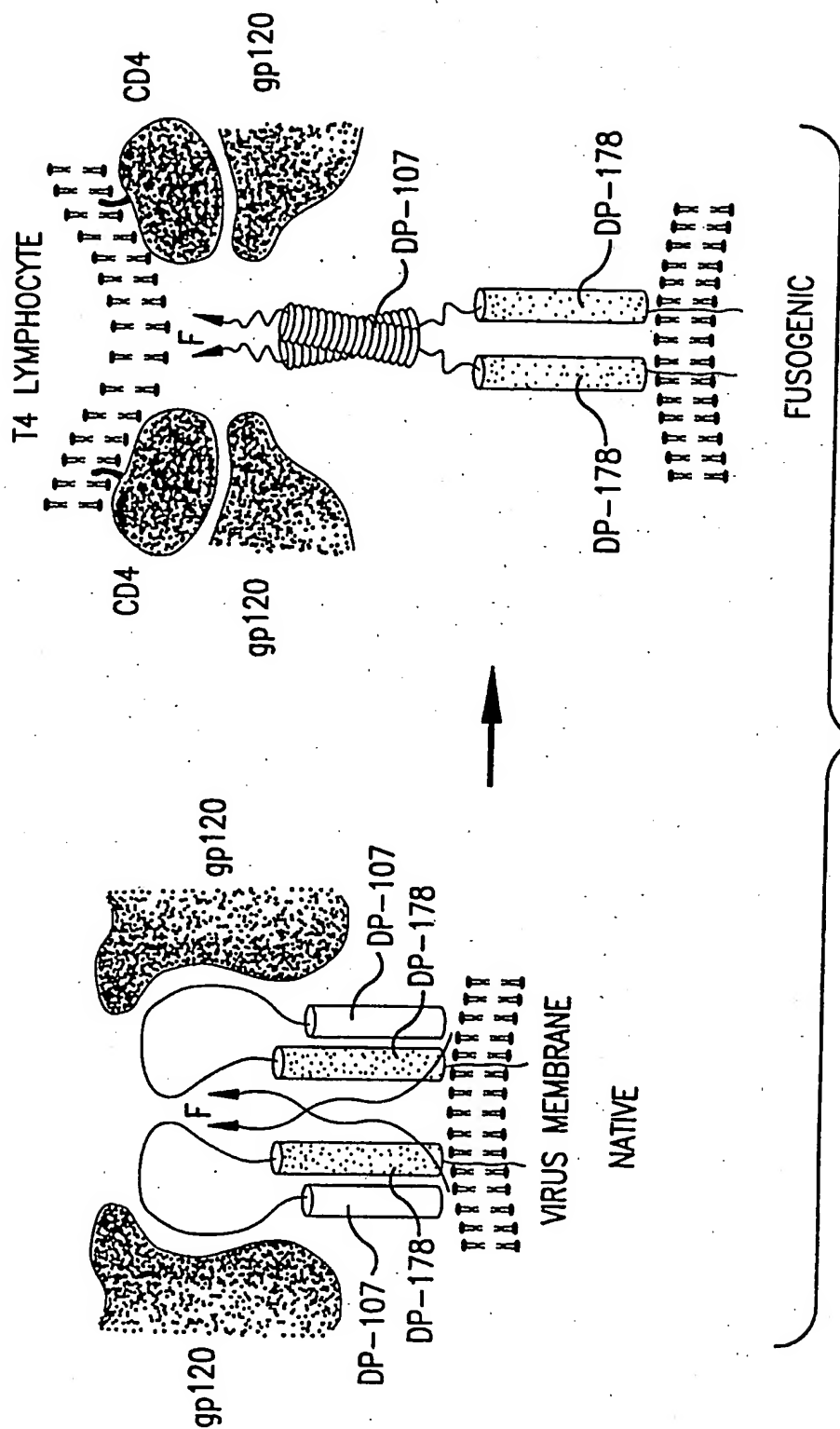


FIG.11A

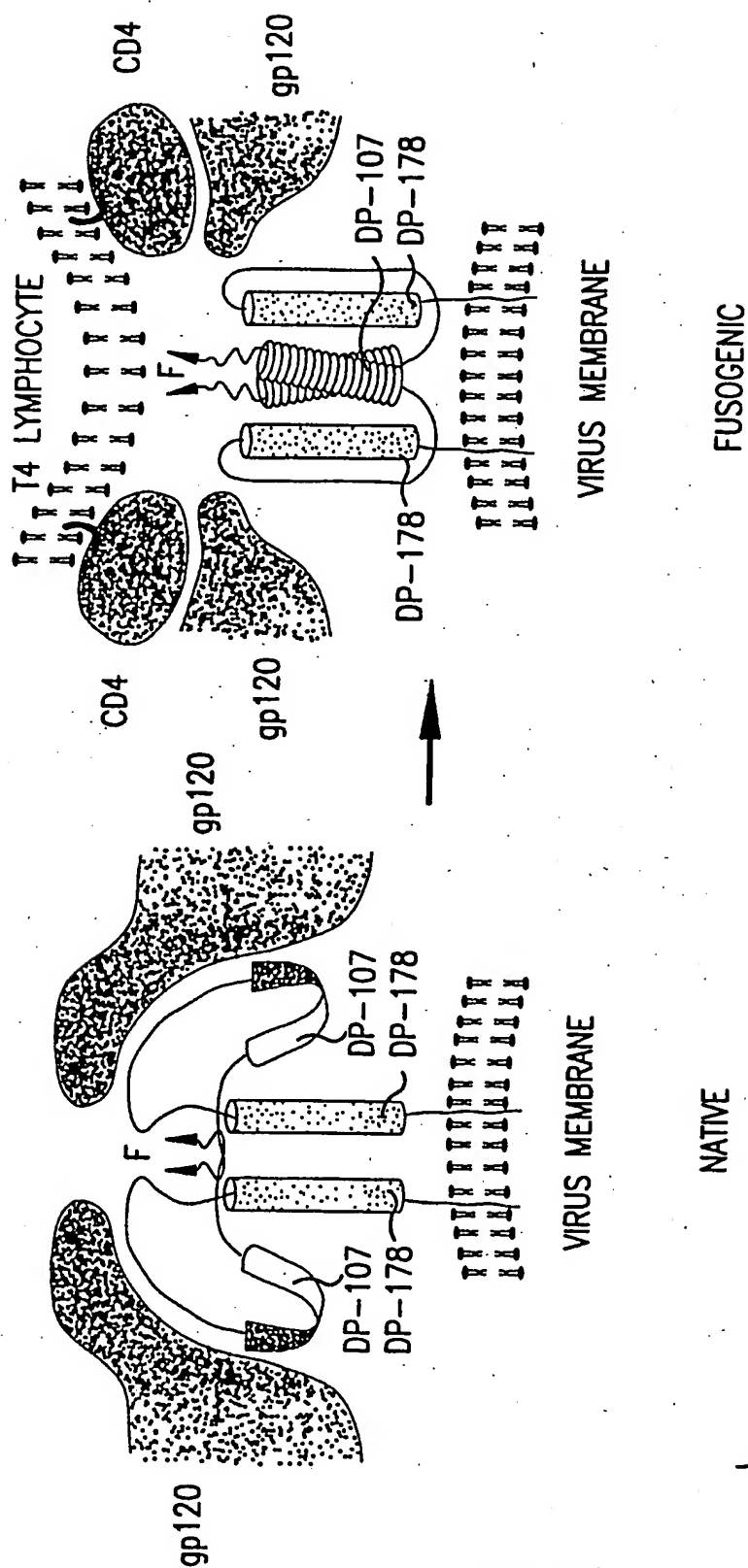


FIG.11B

Sequence	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	Motifs																		
DP-107 (env_hv1bru) L1=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	A	V	E	R	Y	L	K	D	Q	{ILQT} {CFIMPSTY}	
DP-107 (env_hv1bru) L1=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	A	V	E	R	Y	L	K	D	Q	{ILQTV} {CDFIMPST}	
DP-107 (env_hv1bru) L1=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	A	V	E	R	Y	L	K	D	Q	{ILQTV} {CDFIMPST}	
DP-107 (env_hv1bru) L2=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	A	V	E	R	Y	L	K	D	Q	{EKLQV} {CDFKMPSTV}	
DP-107 (env_hv1bru) L2=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	A	V	E	R	Y	L	K	D	Q	{EKLQV} {CFKMPST}	
DP-107 (env_hv1bru) L2=D	N	N	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	A	V	E	R	Y	L	K	D	Q	{EKLQV} {CFKMPST}	
DP-178 (env_hv1bru) Y1=A	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	E	L	D	K							{EKLOY} {ACFGMPRVWY}		
DP-178 (env_hv1bru) Y1=A	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	W	{EKLQWY} {CFGMPRVY}	
DP-178 (env_hv1bru) Y1=A	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	W	F	{EFKLQWY} {CFGMPRVY}
DP-178 (env_hv1bru) Y1=D	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	E	L	D	K							{EILNQSY} {ACFGMPRVWY}		
DP-178 (env_hv1bru) Y1=D	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	W	{EILNQSWY} {CFGMPRVY}	
DP-178 (env_hv1bru) Y1=D	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	W	F	{EFILNQSWY} {CFGMPRVY}

FIG.13

Sequence	Positions																								Parent Motif	Hybrid Motif													
	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D																	
GCN4 (gcN4 yeast)	M	K	Q	L	E	D	K	V	E	E	L	L	S	K	N	Y	H	L	E	N	E	V	A	R	L	K	K	L		[LMNV] {CFGIMPSTW}									
DP-107 (env_hv1bru) L1=D	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I			[ILOQ] {CFIMPSTY}	[ILMNQTV] {CFIMPT}								
DP-107 (env_hv1bru) L1=D	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L	[ILOQV] {CDFIMPST}	[ILMNQTV] {CFIMPT}			
DP-107 (env_hv1bru) L1=D	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L	K	D	Q	[ILOQV] {CDFIMPST}	[ILMNQTV] {CFIMPT}
DP-107 (env_hv1bru) L2=D	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I									[EKLNOV] {CDFKMPSTV}	[EKLNOV] {CFNP}		
DP-107 (env_hv1bru) L2=D	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L		[EKLNOV] {CFKMPST}	[EKLNOV] {CFNP}		
DP-107 (env_hv1bru) L2=D	N	N	L	L	R	A	I	E	A	Q	H	L	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L	K	D	Q	[EKLNOV] {CFKMPST}	[EKLNOV] {CFNP}

FIG. 14

Sequence	Positions																								Parent Motif	Hybrid Motif							
	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D											
GCN4 (gcn4 yeast)	M	K	Q	L	E	D	K	V	E	L	L	S	K	N	Y	H	L	E	N	V	A	R	L	K	K	L			[LMNV] {CFGIMPTW}				
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	E	K	N	E	Q	E	L	L	E	L	D	K			[EKLOY] {ACFGMPRVWY}	[EKLNDVY] {CFGMPH}			
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	[EKLNDVY] {CFGMPH}
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	[EKLNDVY] {CFGMPH}
DP-178 (env_hv1bru)Y1=D	Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	[EKLNDVY] {CFGMPH}
DP-178 (env_hv1bru)Y1=D	Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	[EKLNDVY] {CFGMPH}
DP-178 (env_hv1bru)Y1=D	Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	[EKLNDVY] {CFGMPH}

FIG. 15

Sequence	Positions																				Parent Motif		Hybrid Motif																	
	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D																				
DP-107 (env_hv1bru)L1=D	N	N	L	R	A	I	E	A	Q	H	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L	K	D	Q		[ILQTV] {CDFIMPST}			
DP-107 (env_hv1bru)L2=D	N	N	L	R	A	I	E	A	Q	H	L	Q	L	T	V	W	G	I	K	Q	L	Q	A	R	I	L	A	V	E	R	Y	L	K	D	Q		[EKLNGV] {CFKAPS}			
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	E	L	D	K	W	A	S	L	W	N	W	F		[EFKLQWY] {CFGMPRVY}			
DP-178 (env_hv1bru)Y1=D					Y	T	S	L	I	H	S	L	I	E	S	Q	N	Q	Q	E	K	N	E	Q	E	L	L	D	K	W	A	S	L	W	N	W	F		[EFILNGSWY] {CFGMPRVY}	[EFILNGSTVWY] {CFMP}
FLU LOOP 36	I	E	K	T	N	E	K	F	H	Q	I	E	K	E	F	S	E	V	E	G	R	I	Q	D	L	E	K	Y									[FILTV] {ACFLMPTVW}			

Parent Motif	Hybrid Motif
[ILOTV] {CDFIMPST}	
[EKLNOV] {CFKMPST}	
[EFKLOWY] {CFGMPRVY}	
[EFILNOSWY] {CFGMPRVY}	[EFIKLNDSTWY] {CFMP}
[FILTIV] {ACFLMPTVH}	

FIG.16

Sequence	Positions															Parent Motif	Hybrid Motif
GCN4 (gcN4 yeast)	A	D	A	D	A	D	A	D	A	D	A	D	A	D		[LMNV] {CFGIMP TH }	
DP-107 (env_hv1bru) L1=D	MKQL	EDKVEEL	LSKN	YHL	ENE	VAR	LKKL									[ILQTV] {CDFIMP ST }	
DP-178 (env_hv1bru) Y1=A	NNLL	RAIEAQ	HLQL	TLV	WGI	KQL	QAR	I	L	A	V	E	R	Y	L	K	D
	YTS	LIMS	LIEES	QNQ	QEK	NEQ	ELLE	D	K	W	A	S	L	W	N	W	F
GCN4 (gcN4 yeast)	MKQL	EDKVEEL	LSKN	YHL	ENE	VAR	LKKL									[LMNV] {CFGIMP TH }	
DP-107 (env_hv1bru) L1=D	NNLL	RAIEAQ	HLQL	TLV	WGI	KQL	QAR	I	L	A	V	E	R	Y	L	K	D
DP-178 (env_hv1bru) Y1=D	YTSL	IHS	LIEES	QNQ	QEK	NEQ	ELLE	D	K	W	A	S	L	W	N	W	F
GCN4 (gcN4 yeast)	MKQL	EDKVEEL	LSKN	YHL	ENE	VAR	LKKL									[LMNV] {CFGIMP TH }	
DP-107 (env_hv1bru) L2=D	NNLL	RAIEAQ	HLQL	TLV	WGI	KQL	QAR	I	L	A	V	E	R	Y	L	K	D
DP-178 (env_hv1bru) Y1=A	YTSL	IHS	LIEES	QNQ	QEK	NEQ	ELLE	D	K	W	A	S	L	W	N	W	F
GCN4 (gcN4 yeast)	MKQL	EDKVEEL	LSKN	YHL	ENE	VAR	LKKL									[LMNV] {CFGIMP TH }	
DP-107 (env_hv1bru) L2=D	NNLL	RAIEAQ	HLQL	TLV	WGI	KQL	QAR	I	L	A	V	E	R	Y	L	K	D
DP-178 (env_hv1bru) Y1=D	YTSL	IHS	LIEES	QNQ	QEK	NEQ	ELLE	D	K	W	A	S	L	W	N	W	F

FIG.17

Sequence	Positions																Parent Motif	Hybrid Motif
	A	D	A	D	A	D	A	D	A	D	A	D	A	D	A	D		
GCN4 (gcn4 yeast)	M	K	Q	L	E	D	K	V	E	E	L	L	S	K	N	Y	H	[LMNV] {CFGIMPTW}
DP-107 (env_hv1bru)L1=D	N	N	L	R	A	I	E	A	Q	Q	H	L	L	Q	L	T	V	[ILOTV] {CDFIMPST}
DP-107 (env_hv1bru)L2=D	N	N	L	R	A	I	E	A	Q	Q	H	L	L	Q	L	T	V	[EKLNV] {CFKIPS}
DP-178 (env_hv1bru)Y1=A	Y	T	S	L	I	H	S	L	I	E	E	S	Q	N	Q	Q	E	[EFKLOWY] {CFGMPRVY}
DP-178 (env_hv1bru)Y1=D																		[EFILNDSWY] {CFGMPRVY}
C-FOS (fos_human)	T	D	T	L	Q	A	E	T	D	Q	L	E	D	E	K	S	A	[IKLT] {CFGHIMPRVWY}
C-JUN (top1_human)	I	A	R	L	E	E	K	V	K	T	L	K	A	Q	N	S	E	[AILNV] {CDFGHILPWY}
C-MYC (myo_human)	E	Q	K	L	I	S	E	E	D	L	L	E	K	R	R	E	Q	[ELR] {ACFGMPVWY}
FLU LOOP 36	I	E	K	T	N	E	K	F	H	Q	I	E	K	E	F	S	E	[FILTV] {ACFLMPTWH}

[AEFIKLNDQSTVWY] {CFP}

= {CDGHP} {CFP}

1

FIG.18

$P-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(1)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(2)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(3)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(4)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(5)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(7)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(8)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(9)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-\{P\}(10)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-X(1,12)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$
 $P-X(13,23)-[LIV]-\{P\}(6)-[LIV]-\{P\}(6)-[LIV]$

FIG.19

Fusion ♡ALLMOTI5♡
 Peptide ♡107x178x4♡
 ♡.....ELGELG A AGSTMGARSM TLTVQARQ ♡LLSGIVQOO DP107-NNL

LRAIEAQOHL LOLTWGIKO LOARILAYER YLKDO-DP107 QLLG ♡ ♡ I WGC

 ♡107x178x4♡
 ♡ALLMOTI5♡ *LVS Coiled-Coil*
 SGKLICT TAVP ♡WNASWS NKSLEQIWNN MTWM *E ♡WDREINN DP178-

YTSLIHSL IEESONQOEK NEOELLELDK* WASLWNWF-DP178 NI

 ♡Transmembrane Region ♡
 TNWLWYIK ♡ ♡IEIMIVGGLVGLRIVEAVLSIV NRVROGYS ♡ PL

 ♡P23LZIPC ♡
 SFQTHLPTPR GPDR ♡PEGIEE EGGERDRDRS IRLVNGSLAL IWDDLRLS ♡ CL

 ♡ALLMOTI5♡ ♡107x178x4♡
 F ♡SYHRLRDLL LIVTRIVELL GRRGW ♡EALKY WWNLLQYWSQ

ELKNSAVSLLNAT ♡ AIAVAEG TDRVIEVVQGA ♡ CRAIRHIPR

RIRQGLERIL L

FIG. 20

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SUBSTITUTE SHEET (RULE 26)

Fusion ♡ALLMOTIS♡
 Peptide ♡107x178x4♡
 ♡.....FLGFL LGVGSALAS GVA ♡VSKVLHL EGEVNKIKSA

 ♡P1&12LZIPC♡
LLSTNKAYVS LSNGVSVLTS KVLDLKNYID KQ ♡ ♡ LL ♡PIVNKQ

 ♡107x178x4♡
 SC ♡SISNIETV I ♡ EFQOKNNRLLEITREFSYNAG ♡ VTPVSTMLTNSSELLSL

 ♡P1&12LZIPC♡
 ♡ALLMOTIS♡
 INDM ♡PI ♡TNDQ KKLMSNNVQI V ♡ RQSYSI ♡ MS IKKEEVLAYV

VQ ♡ LPLYGVID TPCWKLHTSP LCTTNTKEGS NICLTRTRDG WYCDNAGSVS

FFPQAETCKV QSNRVFCDTM NSLTLPSEIN LCNVDIFNPK

YDCKIMTSKT DVSSSVITSL GAIVSCYGKT KCTASNKNRG

IIKTFSNGCDYVSNKGMDTV SVGNTLYYVN KQEGKSLYVK G

 ♡P7, 12, & 23LZIPC♡
 ♡107x178x4♡ ♡ALLMOTIS♡
 EPIINFYDPLVF ♡PSDE ♡EDASISQVNEKINOSLAF ♡I ♡ RKSDELL ♡

 ♡Transmembrane Region ♡
HNVNA ♡ GK STTN ♡IMITLIIIVIIIVILLS LIAVGLLLY ♡ C ♡

KARSTPVTLS KDQLSGINNI AFSN

FIG. 21

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Fusion
 Peptide ♡ALLMOTI5♡ ♡107x178x4♡
FLGFLG ♡AAGTAMGAAA ♡TALTYQSOHLLAGILOQOKNLLAAY

♡107x178x4♡
EAQ ♡ QQM ♡ LKLTIWGVKNLNARVTALEKYLEDOARLN ♡ AWG ♡ CA

LVS Coiled-Coil
 ♡ALLMOTI5♡ ♡107x178x4♡
 WKQVCHTTVP WQWNNRTPDW ♡NNMT *WLE ♡WEROISYLEGNTT

♡107x178x4♡
TOLEEARAQEEKNLD ♡ AYOKLSS* WSDFWSW ♡ FDF ♡SKWLN ♡ILK

♦Transmembrane Region♦
IGELDYLGIGLRLLYTV ♦ YS ♡ CIARVRQGYS PLSPQIHHP WKGQPDNAEG

PGEKGDKRKN SSEPWQKESG TAEWKS NWCK RL TNWCSISS IWL YNS

♡ALLMOTI5♡
 ♡CLTL LVHLRSAFQY IQYGLGELKA AAQEAVVALA RLAQNAGYQIWL♡

ACRSAYRA IINSPRRVRQ GLEGILN

FIG. 22

Fusion ♣107x178x4♣
 Peptide ♡ALLMOTIS♡ *LVS Coiled-Coil*
FAG ♡VYL AGVALGVATA AQITAGIALHQ ♣*SNLNAQAIQ

SLRTSLEOSNKAIEEIREATOETVIA* YQGVODY♣ VNNEL♡ VP

♡ALLMOTIS♡
 ♣107x178x4♣
 ♣P6 & 12LZIPC♣

AMQHMSCELVGQRLGLRLLRYYTELLSIFGPSLRD ♣PISA ♣♡EISIQALIVAL

GGEEHKILEKLGYSGSD♣ MIAILES RGIKTKI♡ THVDLP GKF ILSISY

♣P1 & 12LZIPC♣
 ♣PTLSEVKGVIVHRLEAV♣ SYNIGSQEWYTTVP RYIATNGYLISNFD ESSCVFVS

ESAIC SQNSL YPMSPLLQQC IRGDTSSCAR TLVSGTMGNK FILSKGNIVA

NCASILCKCY STSTINQSP DKLLTFIASD TCPLVEIDGA TIQVGGRQYP

LVS Coiled-Coil
 ♡ALLMOTIS♡
 ♣P12 & 23LZIPC♣

DMVYEGKVAL G ♣PAISLD ♡RL*DVGTNLGNALKKLDDAKVLI♣

♦Transmembrane Region♦

DSS♣ NOILETVR RS♡* SFN ♦FGSLL SVPILSCTAL ALLLLIYCC♦

K RRYQQTLKQH TKVDPAFKPD LTGTSKSYVR SL

FIG. 23

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SUBSTITUTE SHEET (RULE 26)

Fusion ♥ALLMOTI5♥
 Peptide ♣107x178x4♣
 ♥.....FIGAI IGSVALGVA TAAQITAASA LIQANQNAAN ♣ILRLKETTA

TIEAVHEVTDGLSOLAVA♣ VG KM♥ QQFVNDQFNNTAQELDCIKITQQV

♥ALLMOTI5♥
 GVELNLYLTELT TV FGPQITSPAL ♥TQLTIQALYNAGGNMDYLLTKLGVG

♣P1 & 12LZIPC♣
 NNQLSSLIGSGLIT GN♥ ♣PILYDSQT QLLGIQVTLP SVGNLNNMRATYLET

LSVST TKGFASALVP KVVVTQVGSVI EELDTSYCIE TDL DLYCTRI VTFPMSPGIY

SCLNGNTSAC MYSKTEGALT TPYMTLKGSV IANCKMTTCR CADPPGIISQ

♥ALLMOTI5♥
 ♣107x178x4♣
 NYGEAVSLID RHSCN ♣♥VLSLD GITRLSGEF DATYQKNISI LDSQVIVTG

LVS Coiled-Coil ♠Trans-
 NLDISTELGNV NNSISNALDK LEESNSKLDK VNVKLTSTSA ♠LIT YIA

membrane Region♠
LT AISLVCGILSLV♥♣ LACYLMY♠ KQKAQKTLWLGNNTLGQMRATTKM

FIG. 24

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Fusion ♥ALLMOTIS♥
 Peptide ♠107x178x4♠ *LVS Coiled-Coil*
EEGGV ♠IG ♥TIALG *YATSAQITAAAVALVEAKOARSDIEKLKE

AIRDTNKAVQSVSSIGNLIVAIKSVQ* DYVNKE♥♠ IVPSIARLGCEAAG

♥ALLMOTIS♥
 ♠107x178x4♠
 LQLGIALTQH ♠♥YSELTNIFGDNIGSLQEKGIKLOGIASLYRTNITE♥♠

♠P5 & 12LZIPC♠
 IFTTSTVDKYDIYDLLFTESIKVRVIDVDLNDYSITLQVRL ♠PLLTRLNTQIYR

VDSISYNI♠ QNREWI♠ PLPSHIMTKGAFLGGADVKECIEAFSSYIC

PSDPGFVLNHEMESCLSGNISQCPRTVVKSDIVPRYAFVNGGVVANCITT

TCTCNGIGNRINQPPDQGVKIITHKECNTIGINGMLFNTNKEGTLAFYTP

♥ALLMOTIS♥
 ♠107x178x4♠
 ♠P6 & 23LZIPC♠
 NDITLNNSVALD ♠PIDI ♠SIELN ♥KAKSDLEESKEWI♠ RRSNOKL♠

♦Transmembrane Region♦
DSIGNWHOSSTT ♦IIIV♠ LIMIIILFIINVTII♦ IIAVKYY♥ R
 IQKRNRVDQN DKPYVLTNK

FIG. 25

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Fusion

Peptide

.....GLEGAI AGFIENGWEGMIDGWYGFRHQNSEGTG

♠107x178x4♠

♥ALLMOTIS♥

LVS Coiled-Coil

*Q ♥AADLKST ♠QAADQINGKLN RVIEKTNEKTHQIEKEESEVEGRIQ

DLEKYVEDTKIDL* WSYNAELLVALENQHTI♠ DLT♥ DSEMKNLFETR

RQLRENAEEMGNGCFKIYHKCDNACIESIRNGTYDHDVYRDEALNNRFQIKG

VELKSGYKDWILWISFAISCFLLCVVLLGFIMWACQQRGNIRCNICI

FIG. 26

AV	CD	RSV F2	YTSVITIELSNIKENKNGTDAKVKL IKOELDKYKNAVTELQLLMQST
+	+ / ++	T-142	YTSVITIELSNIKENKNGTDAKVKL IKOELDKYK
++	+ / +++	T-143	TSVITIELSNIKENKNGTDAKVKL IKOELDKYKN
+	+ / ++	T-144	SVITIELSNIKENKNGTDAKVKL IKOELDKYKNA
-	+ / +	T-145	VITIELSNIKENKNGTDAKVKL IKOELDKYKNAV
-	+ / -	T-146	ITIELSNIKENKNGTDAKVKL IKOELDKYKNAV
-	-	T-147	TIELSNIKENKNGTDAKVKL IKOELDKYKNAVTE
-	-	T-148	IELSNIKENKNGTDAKVKL IKOELDKYKNAVTEL
-	+ / -	T-149	ELSNIKENKNGTDAKVKL IKOELDKYKNAVTELO
-	-	T-150	LSNIKENKNGTDAKVKL IKOELDKYKNAVTELOL
-	+ / +	T-151	SNIKENKNGTDAKVKL IKOELDKYKNAVTELOLL
-	+ / ++	T-152	NIKENKNGTDAKVKL IKOELDKYKNAVTELOLLM
-	+ / +	T-153	IKENKNGTDAKVKL IKOELDKYKNAVTELOLLMQ
-	+ / ++	T-154	KENKNGTDAKVKL IKOELDKYKNAVTELOLLMQS
-	+ / +	T-155	ENKNGTDAKVKL IKOELDKYKNAVTELOLLMQST

FIG.27

AV	CD	RSV	
+++	+/ -	T-67	DEFDASISQVNEKINOSLAFIRKSDELL
+/ -		F1-178	GEPIINFYDPLVFPSPDEFDASISQVNEKINOSLAFIRKSDELLHNWAGKSTT
+/ -		T-104	IINFYDPLVFPSPDEFDASISQVNEKINOSLAFIRK
+/ -		T-105	INFYDPLVFPSPDEFDASISQVNEKINOSLAFIRKS
+/ -		T-106	NFYDPLVFPSPDEFDASISQVNEKINOSLAFIRKSD
+		T-107	FYDPLVFPSPDEFDASISQVNEKINOSLAFIRKSDE
++		T-108	YDPLVFPSPDEFDASISQVNEKINOSLAFIRKSDEL
+++		T-109	DPLVFPSPDEFDASISQVNEKINOSLAFIRKSDELL
+		T-110	PLVFPSPDEFDASISQVNEKINOSLAFIRKSDELLH
+++		T-111	LVFPSPDEFDASISQVNEKINOSLAFIRKSDELLHN
+++	+/ -	T-112	VFPSPDEFDASISQVNEKINOSLAFIRKSDELLHN
++	+/ -	T-113	FPSDEFDASISQVNEKINOSLAFIRKSDELLHN
+++	+/ -	T-114	PSDEFDASISQVNEKINOSLAFIRKSDELLHNNA
+++	+/ -	T-115	SDEFDASISQVNEKINOSLAFIRKSDELLHNWAG
++	+/ -	T-116	DEFDASISQVNEKINOSLAFIRKSDELLHNWAG
++	+/ -	T-117	EFDASISQVNEKINOSLAFIRKSDELLHNWAGK
++	+/ -	T-118	FDASISQVNEKINOSLAFIRKSDELLHNWAGKS
+++	+/ -	T-119	DASISQVNEKINOSLAFIRKSDELLHNWAGKSTT

(T-67 LIKE)

FIG.28

AV	CD	HPF3	178	YTPNDITLNNVALDPIDISIELNKAQSDLEESKEWIRRSNQKLDISIGNWHOSSTT
-	-	189		YTPNDITLNNVALDPIDISIELNKAQSDLEESKE
-	-	190		TPNDITLNNVALDPIDISIELNKAQSDLEESKEW
-	-	191		PNDITLNNVALDPIDISIELNKAQSDLEESKEWI
-	-	192		NDITLNNVALDPIDISIELNKAQSDLEESKEWIR
-	+/-	193		DITLNNVALDPIDISIELNKAQSDLEESKEWIRR
+/-	+/-	194		ITLNNVALDPIDISIELNKAQSDLEESKEWIRRS
+/-	+/+ +	195		TLNNVALDPIDISIELNKAQSDLEESKEWIRRSN
+	+/+	196		LNNVALDPIDISIELNKAQSDLEESKEWIRRSNQ
+	+/+	197		NNSVALDPIDISIELNKAQSDLEESKEWIRRSNQK
+++	+/+	198		NSVALDPIDISIELNKAQSDLEESKEWIRRSNQKL
++	+/+	199		SVALDPIDISIELNKAQSDLEESKEWIRRSNQKLD
-		200		VALDPIDISIELNKAQSDLEESKEWIRRSNQKLD
+++		201		ALDPIDISIELNKAQSDLEESKEWIRRSNQKLD
+++		202		LDPIDISIELNKAQSDLEESKEWIRRSNQKLD
+++		203		DPIDISIELNKAQSDLEESKEWIRRSNQKLD
+++		204		PIDISIELNKAQSDLEESKEWIRRSNQKLD
+++		205		IDISIELNKAQSDLEESKEWIRRSNQKLD
+		206		DISIELNKAQSDLEESKEWIRRSNQKLD
+		207		ISIELNKAQSDLEESKEWIRRSNQKLD
+		208		SIELNKAQSDLEESKEWIRRSNQKLD
++		209		IELNKAQSDLEESKEWIRRSNQKLD
++		210		ELNKAQSDLEESKEWIRRSNQKLD

FIG.29

CD	HPF3 107	GT	ALGVATSAQITA AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNLIVA I KSVQDYVNKE IVP
+/+	157		ALGVATSAQITA AVALVEAKQARSDIEKLKEAIRD
+/+	158		LGVATSAQITA AVALVEAKQARSDIEKLKEAIRDT
+/-	159		GVATSAQITA AVALVEAKQARSDIEKLKEAIRDTN
+/+	160		VATSAQITA AVALVEAKQARSDIEKLKEAIRDTNK
+/+	161		ATSAQITA AVALVEAKQARSDIEKLKEAIRDTNKA
+/-	162		TSAQITA AVALVEAKQARSDIEKLKEAIRDTNKAV
+/+	163		SAQITA AVALVEAKQARSDIEKLKEAIRDTNKAVQ
+/+++	164		AQITA AVALVEAKQARSDIEKLKEAIRDTNKAVQS
+/+	165		QITA AVALVEAKQARSDIEKLKEAIRDTNKAVQSV
+/-	166		ITA AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQ
+/-	167		TA AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQS
+/-	168		AAVALVEAKQARSDIEKLKEAIRDTNKAVQSVQSS
+/-	169		AVALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSI
+/-	170		VALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIG
+/-	171		ALVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGN
+/-	172		LVEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNL
+/-	173		VEAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNL I
+/++	174		EAKQARSDIEKLKEAIRDTNKAVQSVQSSIGNL IV
	T-40		AKQARSDIEKLKEAIRDTNKAVQSVQSSIGNL IVA
+/+	175		KQARSDIEKLKEAIRDTNKAVQSVQSSIGNL IVAI
+/+++	176		QARSDIEKLKEAIRDTNKAVQSVQSSIGNL IVAIK
+/-	177		ARSDIEKLKEAIRDTNKAVQSVQSSIGNL IVAIKS
+/-	178		RSDIEKLKEAIRDTNKAVQSVQSSIGNL IVAIKSV
-	179		SDIEKLKEAIRDTNKAVQSVQSSIGNL IVAIKSVQ
-	180		DIEKLKEAIRDTNKAVQSVQSSIGNL IVAIKSVQD
-	181		IEKLKEAIRDTNKAVQSVQSSIGNL IVAIKSVQDY
-	182		EKLKEAIRDTNKAVQSVQSSIGNL IVAIKSVQDYV
+/++	183		KLKEAIRDTNKAVQSVQSSIGNL IVAIKSVQDYVN
+/+++	184		LKEAIRDTNKAVQSVQSSIGNL IVAIKSVQDYVNK
-	185		KEAIRDTNKAVQSVQSSIGNL IVAIKSVQDYVNKE
-	186		EAIRDTNKAVQSVQSSIGNL IVAIKSVQDYVNKEI
-	187		AIRDTNKAVQSVQSSIGNL IVAIKSVQDYVNKEIV
-	188		IRDTNKAVQSVQSSIGNL IVAIKSVQDYVNKEIVP

FIG.30

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INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/05739

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) : A61K 37/02, 39/12; C12Q 1/70; G01N 33/53

US CL : 424/88, 89; 435/5, 7.1, 7.92-7.95, 974; 530/324-331, 333, 334

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/88, 89; 435/5, 7.1, 7.92-7.95, 974; 530/324-331, 333, 334

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, Biosis

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
NONE	NONE	NONE

☐ Further documents are listed in the continuation of Box C.☐ See patent family annex.

* Special categories of cited documents:	*T later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Z* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search

07 SEPTEMBER 1994

Date of mailing of the international search report

26 SEP 1994

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INTERNATIONAL SEARCH REPORT

International application No.

PCT/US94/05739

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☒ Claims Nos.: 2
because they relate to subject matter not required to be searched by this Authority, namely:

that the claimed subject matter is directed to mental processes.
2. ☒ Claims Nos.: 13-16 and 42-49
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

because the sequences have not been submitted to the International Searching Authority in electronic form.
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

☐
☐

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.